Rituximab in people living with HIV affected by immune-mediated renal diseases: a case-series

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Abstract
Over the last two decades, rituximab (RTX) has played an important role in the treatment of some lymphoproliferative malignancies and immune-mediated diseases. RTX administration is generally safe and well-tolerated, but side effects including late-onset neutropenia, hypogammaglobulinemia, hepatitis B reactivation and rare cases of progressive multifocal leukoencephalopathy have been observed after its administration. Although there are no absolute contraindications regarding its use in people living with HIV (PLWH), the prescription of this drug has been principally limited in patients with oncohematological diseases. In this report, we described the outcome of four PLWH who underwent RTX therapy after the diagnosis of immune-mediated renal disease. The main RTX-associated adverse effects were leukopenia, late-onset neutropenia and decline of CD4+ and CD8+ T-cell counts. In addition, two of the four patients experienced pneumonia requiring hospitalization within six months from the last RTX infusion. We suggest that RTX should be used with caution in PLWH until further evidence emerges on its safety profile in this vulnerable population.

Keywords
HIV, rituximab, antiretroviral therapy, kidney transplantation, neutropenia, rejection

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Introduction
Rituximab (RTX) is a monoclonal chimeric antibody used to target the antigen CD20 expressed on both normal and neoplastic B-cells. Originally, it was included in non-Hodgkin lymphoma and chronic lymphocytic leukemia therapies, whereas now it has also been approved to treat several autoimmune diseases such as rheumatoid arthritis, anti-neutrophil cytoplasmic autoantibody (ANCA)-associated vasculitis and pemphigus vulgaris.1 In the last decade, off-label use of RTX has been extended to immune-mediated renal diseases including cryoglobulinemia-associated nephropathy, membranous glomerulonephritis (MGN) and lupus erythematosus. In addition, the promising results of RTX in induction immunosuppressive therapy, desensitization programs and treatment of antibody-mediated rejection have broadened its therapeutic application in the field of kidney transplantation.2,3

The mechanism of action of drug is only partially understood. Besides the rapid depletion of pre-B and mature B-cells expressing CD20 with subsequent decrease of antibody production, RTX has an immunomodulatory effect on both CD4+ and CD8+ T-cells.4–6

Unfortunately, the beneficial effects of RTX-based therapy are offset by a series of unwanted adverse effects including late-onset neutropenia,7–9

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hypogammaglobulinemia and probably an early decrease in CD4+ T-cell count. RTX seems also associated with an increasing rate of infectious complications due to transient immunosuppression. Recently, Trivin et al. reported that in a retrospective cohort of 98 patients treated with RTX for glomerulonephropathy, 25 of them (21.6%) experienced infectious complications, most frequently bacterial in origin.

The tendency of RTX to increase the susceptibility to infections has likely limited its use in people living with HIV (PLWH). As a result, the drug has been principally used as chemotherapeutic agent to treat lymphoproliferative diseases including non-Hodgkin lymphoma and Castleman disease. The available information about safety of RTX from these sources is scarce and rather conflicting. Indeed, whereas a randomized trial comparing R-CHOP (RTX plus cyclophosphamide, doxorubicin, vincristine and prednisone) with the CHOP regimen showed a higher occurrence of fatal infections in patients treated with RTX, a recent observational study found no differences in the rate of death for infections in patients who received RTX for HIV-associated lymphoma. In light of these opposing results and the little information available on RTX in PLWH, we report our experience in the use of RTX to target B-cells involved in the pathogenesis of immune-mediated renal diseases, focusing on its safety profile.

Cases presentation

Case 1


On presentation, laboratory tests were notable for serum creatinine of 1.4 mg/dl, albumin 2.2 mg/dl and spot urine protein/creatinine ratio of 8.3 mg/mg. She had a normal CD4+ T-cells count (1070 cells/µl) with undetectable HIV-1 viral load (VL) (<40 copies). Antiretroviral therapy (ART) consisted of three-drug combination therapy with lamivudine, efavirenz and abacavir.

The renal biopsy performed revealed a histological pattern compatible with MGN (stage 2), confirmed by serum levels of anti-phospholipase-A2-receptor autoantibodies (anti-PLA2R) of 237.64 UR/ml. The patient was to receive intravenous (IV) infusion of RTX 375 mg/m² once weekly for four consecutive weeks in July 2017 without antimicrobial prophylaxis.

After three doses of RTX administered at a dosage of 750 mg, the patient experienced moderate neutropenia (770 vs. 3250 cells/µL) in the absence of fever or other signs of infections. A single dose of granulocyte colony-stimulating factor (G-CSF) was used to raise the neutrophil count that reached a normal value (3260 cells/µL) after 19 days from its nadir. After RTX treatment, HIV VL remained unchanged, but T CD4+ cell count decreased slightly to 611 cells/µL, even though it remained in the reference range throughout the observation period. Despite the decrease of anti-PLA2R antibody levels (1.9 UR/ml), proteinuria remained steadily in the nephrotic range. In December 2017, worsening renal function (creatinine 2.63 mg/dl vs. a baseline creatinine of 1.5 mg/dl) prompted a course of second-line immunosuppressive therapy consisting of steroids and cyclosporine. In March 2018, she was admitted for fever and dyspnea due to pneumonia that was treated with a course of antibiotic therapy. At the end of one-year follow-up, renal function remained stable (creatinine 1.76 mg/dl, eGFR 30 ml/min) in the absence of proteinuria.

Case 2

A 24-year-old African woman living with HIV underwent kidney transplantation in 2010 after four years spent on dialysis for end-stage renal disease (ESRD) secondary to HIV-associated nephropathy. Her medical history included also multinodular goitre. Her immunosuppression consisted of everolimus in combination with reduced-dose cyclosporine whereas ART was based on dolutegravir and rilpivirine. According to our protocol, prophylaxis against cytomegalovirus and Pneumocystis carinii infection was discontinued at 6 and 12 months after transplantation, respectively.

In May 2017, her post-transplant course was complicated by a slight increase in serum creatinine (1.5 mg/ml vs. a baseline creatinine of 1.3 mg/ml) associated with proteinuria greater than 400 mg/dl.

Renal biopsy revealed a histological pattern compatible with antibody-mediated transplant rejection. The diagnosis was confirmed by the detection of a donor-specific antibody (DSA) titer of 9068 mean fluorescent intensity (MFI).

In light of these histological findings, a course of immunosuppressive therapy including three boluses of methylprednisolone and plasmapheresis was started. Additionally, IV infusion of RTX 375 mg/m² once weekly for four consecutive weeks was administered in order to reduce antibody production, and maintenance immunosuppressive therapy was potentiated with the switch from cyclosporine and everolimus to tacrolimus and mycophenolic acid. Prophylaxis with trimethoprim-sulfamethoxazole was administered for six months.
After 3.1 months from the last RTX dose, a fall in leukocyte and neutrophil counts was seen with a nadir of 220 neutrophils/μL without infectious sequelae. Four doses of G-CSF restored the neutrophil count (2200 cells/μL) three days after its nadir. In April 2019, despite the disappearance of DSAs, a second graft biopsy was performed for elevation in the serum creatinine concentration (2.7 mg/dl) associated with significant proteinuria (1.5 g/daily). Histological evaluation identified progression to the chronic stage of the underlying humoral rejection.

**Case 3**

A 54-year-old African woman living with HIV-2 infection underwent kidney transplantation in 2012 after 12 years spent on dialysis for ESRD caused by malignant hypertension. She also had a medical history of type 2 diabetes mellitus and tertiary hyperparathyroidism. Her immunosuppression consisted of everolimus in combination with reduced-dose cyclosporine, mycophenolic acid and steroids. ART was based on lamivudine, maraviroc and raltegravir. HIV VL was <40 copies and T-CD4+ cell count was 1080 cells/μL.

Post-transplant was complicated by two episodes of biopsy-proven T-cell-mediated graft rejection that were successfully treated with a short course of high-dose corticosteroid.

In August 2013, a rapid increase in serum creatinine level from baseline 1.8 mg/ml to 2.78 mg/dl prompted renal graft biopsy.

The histological evaluation showed a pattern of antibody-mediated rejection that was confirmed by a high titer of DSA (>10,000 MFI both class I and II).

A course of combined immunosuppressive therapy including three boluses of methylprednisolone, four cycles of IV immunoglobulins and a single dose of 700 mg of RTX was administered. The poor experience of RTX in PLWH limited further infusions of the drug. Prophylaxis against *Pneumocystis jirovecii* pneumonia for six months started after the administration of anti-rejection therapy. After five months from RTX treatment, neutrophils reached a nadir of 230 cells/μL in the absence of signs of systemic infection. Treatment with G-CSF and mycophenolic acid withdrawal resulted in a normalization of the WBC count one month later. CD4+ T-cell count reached a nadir of 256 cells/μL a few days after RTX treatment but returned to over 800 cells/μL one month after. Despite DSA becoming negative and HIV VL remaining undetectable, a relentless worsening of renal function continued in the following months and the patient started hemodialysis in May 2014.

**Case 4**

A 43-year-old Caucasian man living with HIV since 2013 underwent kidney transplantation in 2016 after three years spent on peritoneal dialysis for ESRD caused by MGN. His medical history also included failure of a living-donor kidney transplant, HCV infection, hyperthyroidism, and previous syphilis, human herpesvirus 6 and 7 infections. His immunosuppressive therapy was a combination of tacrolimus, mycophenolic acid and steroid, whereas ART was based on dolutegravir and rilpivirine.

Two years after kidney transplant, the patient had a decline in kidney function (serum creatinine raised from a baseline of 1.7 mg/dl to 2.6 mg/dl) in the absence of HIV replication (<40 copies).

A graft biopsy was performed. Detection of high levels of DSA (>10,000 MFI for class II) and histologic evidence of C4d staining and chronic tissue injury lead to the diagnosis of chronic antibody-mediated rejection.

A course of immunosuppressive therapy consisting of a single dose of IV RTX (750 mg) and IV immunoglobulin (0.1 g/kg) was administered in October 2019.

After one week from the second infusion of RTX (750 mg), the patient developed severe *Legionella* pneumonia manifesting with acute dyspnoea and fever. RTX therapy was discontinued and the infection was effectively treated with levofloxacin without sequelae. In March 2019, his CD4+ T-cell count dropped below 100 cells/μL and prophylaxis against *Pneumocystis carinii* pneumonia was started. DSA showed a moderate decline but renal function worsened quickly reaching a pre-dialysis stage with a creatinine value of 5.17 mg/dl in April 2019.

**Discussion**

This case-series showed that RTX should be used cautiously in PLWH because it is associated with some serious post-treatment side effects that may increase the risk of infections. We described the outcome of four PLWH who were treated with RTX after the diagnosis of renal disease triggered by immune-mediated mechanisms. RTX was used as single agent for the treatment of idiopathic MGN in a non-transplanted subject and in combination with other immunosuppressive agents for the treatment of antibody-mediated graft rejection in three kidney transplant recipients (Table 1). RTX was administered uniformly at dose of 375 mg/m² once weekly with the total number of infusions varying from one to four per patient. The difference in RTX dose prescription was ascribed to the early onset of significant adverse effects leading to RTX withdrawal (case 1 and 4) and to the prudential
use of RTX as a new antirejection drug in HIV+ KT recipients (case 3). However, Despite the reduced dosage compared to the standard protocol (once weekly for four consecutive weeks), RTX consistently decreased the level of serum anti-PLA2R antibodies and DSA, serological markers of MGN and antibody-mediated rejection, respectively.

In our patients, RTX was associated with clinically relevant adverse effects such as leukopenia, neutropenia, transient decline in CD4+ and CD8+ T-cell count that potentially increased the susceptibility to infections (Table 1). After RTX therapy, we recognized a decline in the neutrophil count which was clinically relevant only in three patients (75%). Neutropenia developed after a period ranging from 0.7 to 4.8 months from the first RTX infusion in the absence of evident signs of infection or sepsis. Neutropenia was responsive to the administration of G-CSF in all three cases.

Two patients experienced bacterial pneumonia that necessitated hospital admission. In both cases, pneumonia was treated with intravenous antibiotics and resolved without sequelae. We also noticed a tendency toward the reduction of CD4+ and CD8+ T-cell count within 6 months from RTX therapy. Unfortunately, we have few data to trace all changes in immunoglobulin levels in these patients. However, the decline of serum immunoglobulin levels was moderate and probably multifactorial.

Late-onset neutropenia is a delayed adverse effect of RTX. The incidence varies between 3 and 27% and it seems to occur more frequently among patients treated for hematological cancers than autoimmune diseases. The mechanism by which RTX determines neutropenia is still unclear. It has been suggested that the aberrant reconstitution of B-cells after therapy cessation may disrupt neutrophil homeostasis through formation of anti-neutrophil antibodies or production of chemokines. There is compelling evidence supporting the theory of polymorphism in the immunoglobulin (Ig)G Fc receptor (FcγR).

Table 1. Main laboratory and clinical findings in people living with HIV who underwent rituximab therapy.

<table>
<thead>
<tr>
<th>Case 1</th>
<th>Case 2</th>
<th>Case 3</th>
<th>Case 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>65</td>
<td>24</td>
<td>54</td>
</tr>
<tr>
<td>Immune-mediated kidney disease</td>
<td>MGN</td>
<td>AMR</td>
<td>AMR</td>
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<tr>
<td>Total number RTX infusions</td>
<td>3</td>
<td>4</td>
<td>1</td>
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<tr>
<td>RTX dose at each infusion (mg)</td>
<td>750</td>
<td>700</td>
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</tr>
<tr>
<td>Leucocytes count (cells/μL)</td>
<td></td>
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<td></td>
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<tr>
<td>Pre-RTX</td>
<td>8890</td>
<td>5280</td>
<td>7450</td>
</tr>
<tr>
<td>Nadir post-RTX</td>
<td>3336</td>
<td>1720</td>
<td>2120</td>
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<tr>
<td>Lymphocyte count (cells/μL)</td>
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<tr>
<td>Pre-RTX</td>
<td>2820</td>
<td>1600</td>
<td>1900</td>
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<tr>
<td>Nadir post-RTX</td>
<td>2140</td>
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<td>1004</td>
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<tr>
<td>Neutrophil count (cells/μL)</td>
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<td>Pre-RTX</td>
<td>3250</td>
<td>2240</td>
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<tr>
<td>Nadir post-RTX</td>
<td>770</td>
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<td>230</td>
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<tr>
<td>Months from RTX to neutropenia</td>
<td>0.7</td>
<td>3.1</td>
<td>4.8</td>
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<td>Platelet count (10^12 cells/μL)</td>
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<tr>
<td>Pre-RTX</td>
<td>244</td>
<td>360</td>
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<td>Nadir post-RTX</td>
<td>206</td>
<td>298</td>
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<tr>
<td>CD4+ T-cell count (cells/μL)</td>
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<td>Pre-RTX</td>
<td>1070</td>
<td>758</td>
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<td>Nadir post-RTX</td>
<td>611</td>
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<td>CD8+ T-cell count (cells/μL)</td>
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<td>Pre-RTX</td>
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<td>IgG/IgM/IgA (mg/dl)</td>
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<tr>
<td>Pre-RTX</td>
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<td>1310/268/158</td>
<td>797/168/200</td>
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<tr>
<td>Nadir post-RTX</td>
<td>244/106/94</td>
<td>1113/208/179</td>
<td>587/65/167</td>
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<tr>
<td>G-CSF administration</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
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<tr>
<td>Post-RTX infectious complications</td>
<td>Pneumonia</td>
<td>None</td>
<td>None</td>
</tr>
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Abnormal lab test results were considered secondary to the pharmacological effect of rituximab if they occurred within six months from the last rituximab infusion and other potential causes (e.g., infections, drugs, plasmapheresis, etc.) were excluded.

AMR: antibody-mediated rejection; RTX: rituximab; G-CSF: granulocyte colony-stimulating factor; MGN: membranous glomerulonephritis; Ig: immunoglobulin.
Notably, subjects harboring the specific polymorphism in the IgG Fc receptor FCGR3 position 158 V/F have a higher incidence of RTX-induced neutropenia. Likely RTX interacts with high-affinity FCGR3 158 on neutrophils to induce a high apoptotic response in these cells. Late-onset neutropenia remains an unpredictable event and occurs after a median of 38–175 days from last RTX dose. The severity of neutropenia is variable, but in case of neutropenic fever, G-CSF may be given to enhance the recovery of neutrophils.

In the literature, there are no data about the incidence of RTX-induced neutropenia in PLWH. In this group of patients, the drug has been mainly used for HIV-associated lymphoproliferative disease and, to a lesser extent, for rejections in solid organ transplant, anti-glomerular basal membrane disease and autoimmune thrombocytopenia. The majority of information regarding safety and tolerability of the drug has been gleaned from trials on patients with hematologic malignancies where an alarmingly high rate of fatal infections has been found. Only a single episode of fatal progressive multifocal leukoencephalopathy was reported following therapy with RTX in one heart transplant recipient.

It is unclear if PLWH are more prone to developing RTX-related side effects compared to other subsets of the population. CD20 lymphopenia probably aggravates CD4 lymphopenia, and consequently, can worsen the state of immunodeficiency. However, in absence of further randomized trials comparing RTX with other chemotherapy or immunosuppressive regimens, it is difficult to weigh the effects of RTX implicated in late-onset neutropenia and infectious complications because several confounding factors may be involved in this cohort of patients. First, the exposition to combination chemotherapy does not allow us to differentiate the effect of each single agent. Second, the use of protease inhibitors may increase the risk of chemotherapy-induced neutropenia. Lastly, low CD4 T-cell count (<450 cells/μL), a condition commonly found in PLWH, is a risk factor for infections and early death.

The principal limitation of our study is the small number of patients that does not allow us to generalize our findings. Further studies should be carried out in a much broader cohort of PLWH in order to clearly weigh the risks and benefits of RTX therapy in this population. When the use of RTX is inevitable, we cautiously suggest avoiding the drug in neutropenic patients with altered CD4 T-cell count. Given the risk of delayed adverse effects, close follow-up is warranted with frequent and careful monitoring of blood count, CD4+ and CD8+ T-cell counts until further evidence is found on the infectious risk associated with RTX therapy.

Conclusions
In our series, RTX induced serious laboratory abnormalities that may have increased the susceptibility to infections. Based on this observation, the use of this drug deserves special attention in PLWH. Close follow-up with careful monitoring of complete blood count, CD4+ and CD8+ T-cells – even a long time after RTX therapy cessation – is needed in all PLWH undergoing RTX therapy in order to identify and potentially treat life-threatening infections.

Authors’ contributions
Gaetano Alfano and Francesco Giaroni have contributed equally to this work.

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