This is the peer reviewd version of the followng article:
This is the peer reviewd version of the following article.
Spitz naevi and melanomas with similar dermoscopic pattern: can confocal microscopy differentiate? / Guida, Stefania; Pellacani, Giovanni; Cesinaro, A. M; Moscarella, Elvira; Argenziano, Giuseppe; Farnetani, Francesca; Bonamonte, Domenico; Longo, Caterina In: BRITISH JOURNAL OF DERMATOLOGY ISSN 0007-0963 STAMPA 174:3(2016), pp. 610-616. [10.1111/bjd.14286]
Terms of use: The terms and conditions for the reuse of this version of the manuscript are specified in the publishing
policy. For all terms of use and more information see the publisher's website.
10/04/2024 19:54

(Article begins on next page)

## SPITZ NAEVI AND MELANOMAS WITH SIMILAR DERMOSCOPIC

PATTERN: CAN CONFOCAL MICROSCOPY DIFFERENTIATE?

Running head: SNs and MMs with similar dermoscopic pattern: the role of RCM

Stefania Guida<sup>1</sup>, MD, Giovanni Pellacani<sup>1</sup>, MD, Anna Maria Cesinaro<sup>2</sup>, MD, Elvira Moscarella<sup>3</sup>, MD, Giuseppe Argenziano<sup>4</sup>, MD, Francesca Farnetani<sup>1</sup>, MD, Domenico Bonamonte<sup>5</sup>, MD, PhD, Caterina Longo<sup>3</sup>, MD, PhD

#### Affiliations:

- 1.Dermatology Unit, University of Modena and Reggio Emilia, Italy
- 2. Pathology Unit, University of Modena and Reggio Emilia, Italy
- 3. Skin Cancer Unit, Arcispedale S. Maria Nuova-IRCCS, Reggio Emilia, Italy
- 4. Dermatology Unit, Second University of Naples, Italy
- 5. Dermatology Unit, University of Bari "Aldo Moro", Bari, Italy

Word count: 1770

Figures: 6

Tables: 5-2 (1 Supplementary)

References: 24

Conflict of interest: none declared

Funding source: none

Corresponding Author:
Caterina Longo, MD, PhD
Dermatology and Skin Cancer Unit,
Arcispedale Santa Maria Nuova-IRCCS, Reggio Emilia, Italy
Viale Risorgimento, 80
42100 Reggio Emilia, Italy
E-mail: longo.caterina@gmail.com

#### Abstract

Background Differentiating Spitz naevi (SNs) and melanomas (MMs) can be difficult both clinically and dermoscopically. In fact, it is common, in clinical practice, to find SNs simulating MMs, but also MMs simulating SNs. Previous studies reported the potential role of *in vivo* reflectance confocal microscopy (RCM) in increasing diagnostic accuracy.

Objectives To define RCM criteria that can differentiate "false twins", namely SNs and MMs sharing similar dermoscopic appearance.

Methods Lesions histopathologically-diagnosed as SNsSpitz naevi or MMsmelanomas were retrospectively retrieved and they been selected to cover all dermoscopy types and put in «couple». Lesions were classified in three main dermoscopic categories: globular and starburst pattern; spitzoid with dotted vessels; multicomponent or aspecific pattern.

Results RCM findings revealed that marked striking cell pleomorphism within epidermis, widespread atypical cells at the dermo-epidermal junction and marked pleomorphism within nests were significantly associated with the diagnosis of MM while spindled cell and peripheral clefting were exclusively found and pathognomonic of SNs. Furthermore, the analysis of dermoscopic subgroup highlight the importance of striking pleomorphism and spindled cells as clues to differentiate "false twins" with globular or starbust pattern.

Conclusions The current study highlights the role of RCM in discriminating "false twins" of SNsSpitz naevi and MMsmelanomas in adults, in particular

for lesions showing starburst and globular patterns upon dermoscopy whereas RCM is not useful in the other dermoscopic subgroups.

.

#### **Bulleted statements**

what's already known about this topic?

The differential diagnosis between SNsSpitz naevi and MMsmelanomas can be challenging. RCM represents an adds-on tool that improves diagnostic accuracy for skin cancer.

what does this study add?

RCM is useful to discriminate SNsSpitz naevi and MMsmelanomas sharing the same dermoscopic pattern, namely globular or starburst pattern while it's not helpful in other dermoscopic subgroups.

#### INTRODUCTION

In 1948 Sophie Spitz described 13 cases of what she called "juvenile melanoma," underlining its presumably good prognosis because only one case of her series had proven fatal.<sup>1</sup> During the following forty years, however, the entity described by Sophie Spitz was thought to be completely benign, with metastasizing cases being intuitively considered as cases of melanomas simulating Spitz naevi (Spitzoid melanoma). <sup>2,3</sup>

Several attempts have been made to establish objective criteria that can reliably differentiate Spitz Naevi (SNs) and melanomas (MMs) on clinical, histopathological and molecular ground. <sup>2-7</sup>

Dermoscopy significantly improved the recognition of SNsSpitz naevi<sup>3</sup> that can be roughly grouped in 4 main patterns, namely starburst, globular, homogeneous, reticular and inverse network.<sup>3</sup> starburst or globular; negative network or superficial black network; reticular, homogeneous, complex, or multicomponent pattern and nonspecific pattern.<sup>47</sup> <sup>8</sup> As a general rule, a "typical" SNSpitz naevus reveals a symmetric arrangement of colours and structures. Conversely, asymmetrically distribution of dermoscopic structures such as peripheral streaks, pseudopods, or globules are thought to be melanoma criteria.

However, SNsSpitz naevi and MMsmelanomas may show overlapping dermoscopic pattern rendering them as "false twins". Reflectance confocal microscopy (RCM) is a pivotal imaging tool that is used as an adjunct to dermoscopy for the diagnosis of melanocytic tumors.<sup>8-14</sup> 9-15A correlation between dermoscopic findings, histopathologic

aspects and *in vivo* RCM features of <del>SNs</del>Spitz naevi has been previously identified.<sup>8,15-17</sup> <sup>8-9, 16-17</sup>

In our study, we sought to identify distinctive RCM features that can potentially differentiate between "false twins", namely, SNs and MMs typified by a similar dermoscopic pattern.

#### MATERIALS AND METHODS

Study population

Histopathologically-verified SNsSpitz naevi and MMsmelanomas excised between 2010 and 2014 were retrieved from databases of 2 tertiary skin cancer centers (Modena and Reggio Emilia, Italy). Exclusion criteria were lesions located on special body sites such as acral and mucosal areas.

#### Instruments and criteria

#### Dermoscopy

Dermoscopic pictures were collected with DermLite Photo 3Gen (San Juan Capistrano, CA, US). Lesions were selected in order to put them in "couple" that shares similar dermoscopic aspect. Then they were classified, according to dermoscopy, in three main categories: 1. Globular and starburst pattern; 2. spitzoid with dotted vessels; 3. multicomponent or non-specific pattern. These two latter entities were grouped into the third category in order to obtain a larger group.

#### RCM evaluation

After the selection of couples of SNSpitz naevus and MMsmelanoma with similar dermoscopic appearance, an accurate analysis of RCM was performed on the entire lesion area. RCM evaluation was carried out by means of a wide-probe confocal microscope (Vivascope 1500®, MAVIG GmbH, Munich, Germany), obtaining mosaics (Vivablock®) of high-resolution within the epidermis, at the dermal–epidermal junction (DEJ) and within the upper dermis; Vivastack® were also performed, following previously described procedures.<sup>8,17,8-9</sup>

One expert Investigator, different from the one who performed the image acquisition, analyzed all cases according to a list of RCM features<sup>8</sup>
14,17-248-15,18-24 (**Table 1 Supplementary**) blinded from histopathologic diagnosis or other information with the exception of body site. A second investigator was involved in case of uncertainty.

#### Histopathologic evaluation

Histopathologic evaluation was focused on the following criteria: epithelioid and/or spindled cell morphology, cell pleomorphism, pagetoid infiltration, epidermal hyperplasia, epidermal flattening and sharp borders. Then, a side by side comparison between RCM and hematoxylin and eosin slides has been done according to our workup.<sup>25</sup>

#### Statistical analysis

Statistical evaluation was carried out with the SPSS® statistical package (SPSS Inc., Chicago, IL, USA).

Outcome dichotomous variable was set to definite RCM and histopathologic clues of MMmelanoma or SNSpitz nevus. Demographic, clinical, and dermoscopic variables were included in the analysis. Absolute and relative frequencies of observations in benign and malignant lesions were described. Fisher exact test  $\chi^2$  was used to compare RCM and histopathology criteria among different type of lesions. A stratified analysis within each dermoscopic group was also carried out. Cohen's kappa coefficient was calculated in order to find association between confocal features and histology findings.

A *p*-value <0.05 was considered significant.

#### **RESULTS**

A total of 34 cases were included (10 were males and 24 females). Mean age was 49,59±16,42 for melanoma patients and 31±10,72 for subjects with Spitz naevi, ranging from 13 to 81 years.

Of 17 MMsmelanomas, 8 (47%) were *in situ* while 9 were invasive with a mean Breslow's thickness of 0.37 mm. None of the melanoma included was a spitzoid melanoma histologically. Lower limbs were the most frequent body location (18/34 lesions, 52,9%), followed by upper limbs (8/34, 23,5%), trunk (7/34, 20,6%) and only one case located on the head. Analytic results of demographic information and

anatomic site of the tumor are reported in **Table 2**. The dermoscopic appearance of the lesion of the head justified the enrollment in this study. No significant difference was found between MMsmelanomas and SNsSpitz naevi for sex, age and body location.

Based on the dermoscopic analysis, 8 couples of SNs/MMsSpitz naevi/melanomas (16/34 lesions, 47%) were identified within group 1 (**Figs 1-2**), 2 couples (4/34 lesions, 11,8%) within the second group (**Figs 3-4**) and 7 couples (14/34 lesions 41,2%) belonging to group 3 (**Figs 5-6**).

#### RCM evaluation

Several features were evaluated according to different skin depth (**Table 3**2). No significant differences were found for epidermal patterns between SNsSpitz naevi and MMsmelanomas. Pagetoid cells were a common finding in both lesion type and were represented by dendritic/spindled cells more frequently than roundish in SNsSpitz naevi (76,5% *versus* 23,5%, respectively). Hyporeflective pagetoid cells were observed in amelanotic tumors. An higher quantity of pagetoid cells was found in 9 MMs compared to 5 SNs. Similarly, a A widespread pagetoid infiltration occurred in 12 lesions, 9 malignant and 3 benign. A correlation between striking cell pleomorphism and MMmelanoma was established (p=0,010).

Evaluating the DEJ level, no significant association was observed between DEJ architecture and specific lesions group. Atypical cells were present in 14 MMsmelanomas *versus* 9 SNsSpitz naevi. SNsSpitz naevi

were often characterized by the presence of few atypical cells, predominantly spindled (p=0,003), showing numerous markedly atypical cells throughout the DEJ level in only 5 cases. Conversely, MMsmelanomas predominantly showed numerous (58,8%) atypical and pleomorphic cells, distributed throughout the entire lesion (11/17 p=0,037). Sharp borders were related with the diagnosis of benign lesions (p=0,044).

At the level of upper dermis, inflammation and dermal nests (dense, dense non-homogeneous) presented a similar distribution among MMsmelanomas and SNsSpitz naevi while dense and sparse nests were exclusively described in MMsmelanomas (3/17). A marked cell pleomorphism within nests was significantly associated with MMs melanoma (p=0,003).

#### Histopathology evaluation

Histopathologic findings were are reported in **Table 4**. A statistically significant association between cell pleomorphism (p=0,007), pagetoid infiltration (p=0,026) and epidermal flattening (p=0,044) and MMsmelanoma was found while epidermal hyperplasia (p=0,044) and sharp borders (p=0,018) were significantly associated with SNsSpitz naevus.

Other interesting findings were the presence of spindled melanocyte in 47,1% of SNsSpitz naevi vs 11,8% of MMsmelanomas while epithelioid cells characterized 70,6% of SNsSpitz naevi vs 52,9% of MMsmelanomas.

# Dermoscopic group analysis RCM analysis within dermoscopic groups

Group 1 revealed the presence of helpful RCM clues to differentiate MMsmelanomas and SNsSpitz naevi. In details, a marked striking cell pleomorphism within epidermis was significantly associated with the diagnosis of MMmelanoma (p=0,041) while a few dendritic cells, with a prevalence of spindled cells, not involving the entire lesion area, were typically found in SNsSpitz naevi (p=0,026). Furthermore, malignant lesions showed epidermal disruption, roundish pagetoid cells, many dendritic and atypical cells distributing throughout the entire lesion, junctional nests. A ringed pattern at the DEJ and peripheral clefting (being this latter exclusively found in SNsSpitz naevi) were represented in one half of SNsSpitz naevi.

RCM results regarding groups 2 and 3 were not statistically significant.

Within group 2 and 3, epidermal disruption, junctional nests, many dendritic and atypical cells distributed throughout the entire lesion were suggestive for a diagnosis of MMmelanoma while a few dendritic cells not involving the whole lesion represented the expression of a SNSpitz naevus.

Additionally group 3 was typified by the presence of roundish pagetoid cells and marked pleomorphism within nests in MMsmelanomas and ringed pattern and peripheral clefting in SNsSpitz naevi.

Acanthosis, dendritic cells and inflammation were equally represented in both categories.

Histopathology analysis within different dermoscopic groups revealed the presence of cell pleomorphism, significantly associated with MMmelanoma diagnosis for group 1 (p=0,007). Of note, analyzing lesions of the same group, pagetoid infiltration characterized MMsmelanomas whereas spindled cells and sharp boundary were commonly observed in SNsSpitz naevi.

In group 2, pagetoid infiltration and epidermis flattening were suggestive for the diagnosis of a malignant lesions while spindled cells, epidermal hyperplasia and sharp boundary (being these two latter observed exclusively in SNsSpitz naevi) characterized SNsSpitz naevi.

Similarly to previous findings, in MMsmelanomas belonging to group 3, pagetoid infiltration and epidermis flattening could be found and spindled cells in SNsSpitz naevi.

Specific RCM criteria were significantly correlated with histopathologic criteria, namely the presence of sharp borders cut-off (5 and 6 Spitz naevi in RCM and histopathology, respectively) ( $\hat{k}=0,675$ ; p<0,05), spindled cells (8 Spitz naevi in RCM and 2 melanomas and 8 Spitz naevi in histology) ( $\hat{k}=0,398$ ; p<0,05) and striking pleomorphism (9 melanomas and 2 Spitz naevi in RCM and 7 melanomas in histology) ( $\hat{k}=0,502$ ; p<0,05) (Table 5).

#### DISCUSSION

The differential diagnosis between SNsSpitz naevi and MMsmelanomas may represent a diagnostic pitfall both for Dermatologists and Pathologists.

In vivo reflectance confocal microscopy (RCM) is a novel imaging technique enabling the visualization of epidermis, DEJ and papillary dermal structures at a *quasi*-histologic resolution, improving diagnostic accuracy for different skin lesions. 8-14,17-248-15,18-24 Previous studies showed that the presence of sharp border cut-off, junctional nests and melanophages were the most relevant features for differentiating SNsSpitz naevi from MMsmelanomas. 478 However, a low specificity in differentiating MMsmelanomas and SNsSpitz naevi was identified, due to the impossibility to explore the skin in depth, thus hampering the evaluation of mitoses and maturation with depth. 8,15-117 8-9,16-17

In the present study, specific RCM features statistically differed between MMsmelanomas and SNsSpitz naevi, highlighting some differences within each dermoscopic group. The presence of a striking pleomorphism within epidermis and nests and atypical cells scattered throughout the whole lesion area were statistically significant and independently correlated with malignancy. In particular, cell pleomorphism within epidermis was observed within MMsmelanomas belonging to group 1, including starburst and globular lesions. On the other hand, SNsSpitz naevi were characterized by the presence of sharp borders cut-off and spindled cells, these latter being prevalent in group 1. A previous study<sup>17</sup>

<sup>8</sup>showed a similar rate of pleomorphism between SNsSpitz naevi and MMsmelanomas while and the presence of atypical spindled cells in 30% of MMsmelanomas examined. These differences can be justified by lesions included and interobserver variation.

Ulceration and cerebriform clusters were not found because of the recruitment method of MMsmelanomas. In fact, 47% of MMsmelanomas included in this study were *in situ* and the mean Breslow's thickness was less than 1 mm (0.37 mm). Specifically, early spitzoid-looking MMsmelanomas are more likely to show dermoscopic characteristics enabling a dermoscopic match with SNsSpitz naevi.

No RCM criteria were identified for group 2 and 3, including Spitzoid-dotted lesions and lesions with multicomponent or non-specific patterns, respectively. This can be related to the thickness of these lesions, impairing the evaluation of the deeper part of the tumor. Translated into clinical practice, it means that lesions displaying dotted vessels or non-specific pattern do not benefit of RCM imaging and thus, other factors, such as clinical context and dermoscopy, should be considered for their proper management. 4617

Results of histopathologic revision supported in part RCM findings. In fact, within the first group cell pleomorphism characterized MMsmelanomas while sharp borders were associated with SNsSpitz naevi. Furthermore, clues that cannot be easily studied with RCM were found. In particular, epidermal hyperplasia in SNsSpitz naevi and pagetoid infiltration, epidermal flattening in MMsmelanomas. Some of these

histology criteria showed an overlap with RCM findings: sharp borders, cell pleomorphism and spindled cells.

Demographic data revealed that all the lesions included in the study were found in patients older than 12 years of age. Intuitively, studying lesions analyzed by pathologists means including doubtful spitzoid lesions that are most common in patients older than 12 year-old. <sup>17</sup>

The main limitation of this study was the low number of cases selected that is related to the careful and detailed matching of difficult-to-diagnose tumors revealing spitzoid-looking pattern.

Clinical practice highlights the high variability of spitzoid lesions. Our study underlines the need of a careful examination of all spitzoid-looking lesions by combining dermoscopy with RCM to establish an accurate diagnosis for "false twins" and manages them accordingly. It supports the role of RCM in discriminating spitzoid lesions revealing a globular and starburst pattern on dermoscopy, representing the most common dermoscopic patterns of SNs.<sup>5</sup> Further studies with a larger number of lesions are warranted in order to confirm our preliminary results.

#### **REFERENCES**

>9

- 1. Spitz S. Melanoma of childhood. *Am J Pathol* 1948; **24**:591–609.
- 2. Peris K, Ferrari A, Argenziano G *et al.* Dermatoscopic classification of Spitz/Reed naevi. *Clin Dermatol* 2002; **20**:259-62.
- 3. Ferrara G, Argenziano G, Soyer HP *et al.* The spectrum of Spitz naevi. A clinicopathologic study of 83 cases. *Arch Dermatol* 2005; **141**:1381-7.
- 4. Casso EM, Grin-Jorgensen CM, Grant-Kels JM. Spitz naevi. *J Am Acad Dermatol* 1992; **27**:901–13.
- 5. Argenziano G, Scalvenzi M, Staibano S *et al.* Dermatoscopic pitfalls in differentiating pigmented Spitz naevi from cutaneous melanomas. *Br J Dermatol* 1999; **141**:788-93.
- 6. Pellacani G, Cesinaro AM, Seidenari S. Morphological features of Spitz naevus as observed by digital videomicroscopy. *Acta Derm Venereol* 2000; **80**:117-21.
- 7. Requena C, Rubio L, Traves V *et al.* Fluorescence in situ hybridization for the differential diagnosis between Spitz naevus and spitzoid melanoma. *Histopathology* 2012; **61**:899-909.
- 8. Pellacani G, Guitera P, Longo C *et al.* The impact of in vivo reflectance confocal microscopy for the diagnostic accuracy of melanoma and equivocal melanocytic lesions. *J Invest Dermatol* 2007; **127**:2759-65.
  - 9. Guitera P, Menzies SW, Longo C et al. In vivo confocal

microscopy for diagnosis of melanoma and basal cell carcinoma using a twostep method: analysis of 710 consecutive clinically equivocal cases. *J Invest Dermatol* 2012; **132**:2386-9. ->10

- 10. Longo C, Farnetani F, Ciardo S *et al.* Is confocal microscopy a valuable tool in diagnosing nodular lesions? A study of 140 cases. *Br J Dermatol* 2013; **169**:58-67. ->11
- 11. Pupelli G, Longo C, Veneziano L *et al.* Small-diameter melanocytic lesions: morphological analysis by means of in vivo confocal microscopy. *Br J Dermatol* 2013; **168**:1027-33. ->12
- 12. Longo C, Rito C, Beretti F *et al.* De novo melanoma and melanoma arising from pre-existing nevus: in vivo morphologic differences as evaluated by confocal microscopy. *J Am Acad Dermatol* 2011; **65**:604-14. ->13
- 13. Pellacani G, Pepe P, Casari A, Longo C. Reflectance confocal microscopi as a second-level examination in skin oncology improves diagnostic accuracy and saves unnecessary excisions: a longitudinal prospective study. *Br J Dermatol* 2014; **171**:1044-51. ->14
- 14. Alarcon I, Carrera C, Palou J *et al.* Impact of in vivo reflectance confocal microscopy on the number needed to treat melanoma in doubtful lesions. *Br J Dermatol* 2014; **170**:802-8. ->15
- 15. Broganelli P, Titli S, Lallas A *et al.* Spitz/Reed naevi: proposal of management recommendations by the Dermoscopy Study Group of the Italian Society of Dermatology (SIDeMaST). *G Ital Dermatol Venereol* 2014; **149**:601-6. ->16

- 16. Lallas A, Moscarella E, Longo C *et al.* Likelihood of finding melanoma when removing a Spitzoid-looking lesion in patients aged 12 years or older. *J Am Acad Dermatol* 2015; **72**:47-53. ->17
- 17. Pellacani G, Longo C, Ferrara G *et al.* Spitz naevi: in vivo confocal microscopic features, dermoscopic aspects, histopathologic correlates and diagnostic significance. *J Am Acad Dermatol* 2009; **60**:236–47. ->8
- 18. Guitera P, Pellacani G, Longo C *et al.* In vivo reflectance confocal microscopy enhances secondary evaluation of melanocytic lesions. *J Invest Dermatol* 2009; **129**:131–8.
- 19. Pellacani G, Cesinaro A, Seidenari S. In vivo assessment of melanocytic nests in naevi and melanomas by reflectance confocal microscopy. *Mod Pathol* 2005; **18**:469–74.
- 20. Pellacani G, Cesinaro A, Seidenari S. Reflectance-mode confocal microscopy for the in vivo characterization of pagetoid melanocytosis in melanomas and naevi. *J Invest Dermatol* 2005; **125**:532–7.
- 21. Longo C, Zalaudek I, Argenziano G, Pellacani G. New directions in dermatopathology: in vivo confocal microscopy in clinical practice. *Dermatol Clin* 2012; **30**:799–814.
- 22. Segura S, Puig S, Carrera C *et al.* Development of a two-step method for the diagnosis of melanoma by reflectance confocal microscopy. *J Am Acad Dermatol* 2009; **61**:216–29.
  - 23. Benati E, Zalaudek I, Piana S et al. In vivo detection of

peripheral clefting in melanocytic lesions. *Br J Dermatol* 2015 [Epub ahead of print].

- 24. Losi A, Longo C, Cesinaro AM *et al.* Hyporeflective pagetoid cells: a new clue for amelanotic melanoma diagnosis by reflectance confocal microscopy. *Br J Dermatol* 2014; **171**:48-54.
- 25. Pellacani G, Longo C, Malvehy J *et al.* In vivo confocal microscopic and histopathologic correlations of dermoscopic features in 202 melanocytic lesions. *Arch Dermatol* 2008; **144**:1597-608.

#### Figure legends

**Figure 1**. (a) Dermoscopy of a starburst melanoma, 0.63 mm Breslow thickness. (b) High magnification confocal microscopy showing several dendritic and roundish pagetoid cells at epidermal level (arrows circle). (c) Corresponding histology presenting pagetoid infiltration and pleomorphic cells (arrow circle) (hematoxylin-eosin stain; original magnification: 200X).

Figure 2. (a) Dermoscopy of a starburst Spitz nevus. (b) Confocal mosaic High magnification confocal microscopy at dermoepidermal junction showing sharp border cut-off (green line), a peripheral nest (asterisk) atypical cells with focal distribution in an acanthotic lesion. (c) Corresponding histology presenting a peripheral nest (asterisk) and sharply demarcated border (arrow) (hematoxylin-eosin stain; original magnification: 200X).

**Figure 3.** (a) Dermoscopy of a spitzoid-dotted melanoma, 0.5 mm Breslow thickness. (b) High magnification confocal microscopy showing hyporeflective, large and well-demarcated structures (arrows) within the honeycombed pattern epidermis. (c) Corresponding histology presenting non-pigmented pagetoid cells scattered throughout the epidermis (hematoxylin-eosin stain; original magnification: 200X).

**Figure 4**. (a) Dermoscopy of a spitzoid-dotted Spitz nevus. (b) High magnification confocal microscopy showing concentric honeycombed structures centered by thin dermal papillae (asterisks). (c) Corresponding

histology presenting an acanthotic epidermis with elongated cristae separated by thin papillae and hypopigmented melanocytes predominantly clustered in small nests at tip of epidermal cristae (hematoxylin-eosin stain; original magnification: 200X).

**Figure 5**. (**a**) Dermoscopy of a multicomponent melanoma, 1.06 Breslow thickness. (**b**) High magnification confocal microscopy showing nonhomogeneous nests with ill defined contours (asterisks). (**c**) Corresponding histology presenting pleomorphic melanocytes predominantly clustered in small nests (asterisks) (hematoxylin-eosin stain; original magnification: 200X).

**Figure 6**. (a) Dermoscopy of a multicomponent Spitz nevus. (b) High magnification confocal microscopy showing fusiform cells with bipolar elongations or dendrites (arrows). (c) Corresponding histology showing spindled melanocytes (arrows) (hematoxylin-eosin stain; original magnification: 200X).

### **TABLES**

**Table 1 Supplementary**. List and definition of the RCM features evaluated for MMs and SNs.

RCM criteria	Definition
Epidermis Regular honeycombed pattern	large polygonal cells with dark nuclei and bright cytoplasm and cell borders
Regular cobblestone pattern	small polygonal cells with refractive cytoplasm separated by a less refractive border
Atypical honeycombed pattern	irregularity in size of the cells and thickness of the contour within a honeycombed structure
Atypical cobblestone pattern	irregularity in size and/or refractivity of the cells within a cobblestone structure
Epidermal disruption	disarray of the normal architecture of superficial layers with unevenly distributed bright granular particles and cells, in absense of honeycombed or cobblestone pattern
Acanthosis	prominent bright epidermis intermingled with papillae
Ulceration	dark areas, usually with sharp borders and irregular contours, filled with amorphous material and/or clotted bright small particles
Pagetoid cells	large roundish nucleated cells, twice the size of basal keratinocytes, with a dark nucleus and bright cytoplasm
Dendritic pagetoid cells	large cells with bright cytoplasm and dark nucleus with clearly visible dendrites connected to the cell
Roundish pagetoid cells	large bright cells with well outlined border and dark nucleus within the epidermis, represent the most common finding for melanoma diagnosis
Number of pagetoid cells	more than 5 clearly visible pagetoid cells evaluated on five samples of 0.5x0.5mm image
Widespread pagetoid cells	pagetoid cells scattered throughout the whole lesion area
Striking pleomorphism	variability of the aspect of pagetoid cell and/or the presence of cells with bizarre shapes
DEJ	
Ringed pattern	densely packed bright rings corresponding to papillae surrounded by a rim of small bright cells sharply contrasting with the dark background
Junctional nests	compact, round to oval bright cell aggregates, connected with the basal layer of the epidermis

Sheet-like strucure cells distributed at the transition of the DEJ showing

loss of dermal papillae not aggregated in clusters but closely distributed in the same plane with the loss of

dermal papillae

Edged papillae demarcated by a rim of bright cells,

appearing as bright rings sharply contrasting with the

dark background

Non-edged papillae dermal papillae without a demarcating bright rim at the

DEJ

Atypical cells large cells showing a bright cytoplasm with clearly

outlined borders and sharply contrasted dark nucleus

inside, roundish to oval in shapes, sometimes

presenting dendritic-like structures

Spindled cells elongated cells oriented toward the same direction

Number of atypical cells more than 5 clearly visible atypical cells evaluated on

five samples of 0.5x0.5mm image

Widespread atypical cells atypical cells scattered throughout the whole lesion

area

Peripheral Clefting dark slit-like space observed between tumor and

surrounding skin

**Upper dermis** 

Dermal dense nests compact aggregates with sharp margin in which outline

of individual cells is indiscernible or similar in shape,

size and refractivity

Dermal dense nonhomogenous nests compact cell aggregates showing non homogeneity in

cell morphology and reflectivity

Dense and sparse dermal nests cell aggregates with irregular, discohesive margins

showing isolated nucleated cells at the periphery

Cerebriform clusters confluent amorphous brain-like aggregates of low

reflecting cells exhibiting granular cytoplasm without evident nuclei and ill-defined borders, showing a fine

hyporeflective "fissure" like appearance

Marked pleomorphism within nests nonreflecting structures with a well-demarcated border,

containing isolated round to oval cells with dark

nucleus and reflecting cytoplasm, with bizarre shapes

Inflammation large irregularly shaped bright cells with ill-defined

borders and usually no visible nucleus which can be

visible within dermal papillae

**Table 2**. Descriptive demographic and clinical characteristics of the study population.

	<del>MM (17)</del>	<del>SN (17)</del>	p-value
-	<del>N, (%)</del>	<del>N, (%)</del>	•
Age	-	_	-
Mean	4 <del>9,5±16,4</del>	31±10,7	-
-	-	-	-
Age group	-	-	-
<del>&lt;40</del>	<del>6 (35,3)</del>	<del>12 (70,6)</del>	0,084
<del>≥</del> 40	<del>11 (64,7)</del>	<del>5 (29,4)</del>	-
-	-	-	-
Sex	-	-	-
<del>female</del>	<del>10 (58,8)</del>	<del>14 (82,4)</del>	0,259
<del>Male</del>	<del>7 (41,2)</del>	<del>3 (17,6)</del>	-
-	-	-	-
Breslow	-	-	-
-	0,38±0,45	-	-
-	-	-	-
<b>Location</b>	-	-	-
head and neck	<del>1 (5,9)</del>	0	0,47
<del>Trunk</del>	<del>4 (23,5)</del>	<del>3 (17,6)</del>	-
<del>upper limbs</del>	<del>5 (29,4)</del>	<del>3 (17,6)</del>	-
<del>lower limbs</del>	<del>7 (41,2)</del>	<del>11 (64,8)</del>	-
-	-	-	-
Dermoscopic group	-	-	-
4	<del>8 (47,1)</del>	<del>8 (47,1)</del>	-
2	<del>2 (11,8)</del>	<del>2 (11,8)</del>	-
3	<del>7 (41,1)</del>	<del>7 (41,1)</del>	-

\*p-value<0,05

**Table 3** 2. List of RCM features discriminating MMs and SNs (p<0.05).

RCM criteria	MM (17) N, (%)	SN (17) N, (%)	<i>p</i> -value
Epidermis			
Regular honeycombed pattern	1 (5,9)	1 (5,9)	1
Regular cobblestone pattern	0	0	§
Atypical honeycombed	7 (41,2)	12 (70,6)	0,166
Atypical cobblestone	2 (11,8)	1 (5,9)	1,000
Epidemis disruption	7 (41,2)	3 (17,6)	0,259
Acanthosis	14 (82,4)	15 (88,2)	1,000
Ulceration	0	0	§
Dendritic pagetoid cells	14 (82,4)	13 (76,5)	1,000
Roundish pagetoid cells	10 (58,8)	4 (23,5)	0,080
Several pagetoid cells	9 (52,9)	5 (29,4)	0,296
Widespread pagetoid cells	9 (52,9)	3 (17,6)	0,071
Striking pleomorphism	9 (52,9)	2 (11,8)	0,010*
DEJ			
Ringed pattern	5 (29,4)	9 (52,9)	0,296
Junctional nests	13 (76,5)	9 (52,9)	0,282
Sheet-like structure	5 (29,4)	2 (11,8)	0,398
Edged papillae	11 (64,7)	9 (52,9)	0,728
Non-edged papillae	4 (23,5)	5 (29,4)	1,000
Atypical cells	14 (82,4)	9 (52,9)	0,141
Spindled cells	0	8 (47,1)	0,003*
Many atypical cells	10 (58,8)	7 (41,2)	0,494
Widespread atypical cells	11 (64,7)	5 (29,4)	0,037*
Peripheral clefting	0	5 (29,4)	0,044*
Upper dermis			
Regular dense dermal nests	6 (35,3)	6 (35,3)	0,601
Non-homogeneous dense nests	5 (29,4)	3 (17,6)	0,688
Dense and sparse dermal nests	3 (17,6)	0	0,227
Cerebriform clusters	0	0	§
Striking Marked pleomorphism	8 (47,1)	0	0,003*
within nests Inflammation	16 (94,1)	14 (82,4)	0,601

<sup>\*</sup>P-value <0,05

§no statistics was computed because the feature is a constant

**Table 4**. List of histopathologic criteria that discriminate MMs and SNs (p<0.05).

Histopathology criteria	<i>p</i> -value
epithelioid cells	<del>0,282</del>
spindled cells	<del>0,057</del>
pleomorphism	<del>0,007*</del>
<del>pagetoid spread</del>	<del>0,026*</del>
epidermal hyperplasia	<del>0,044*</del>
epidermis flattening	<del>0,044*</del>
sharp boundary	<del>0,018*</del>

<sup>\*</sup>p-value <0,05

**Table 5.** RCM findings correlated with histologic features for MM and SN diagnosis (p<0.05).

<b>Hystopathology</b>	RCM	<del>value</del>	<del>p</del>
sharp boundary	<del>peripheral clefting</del>	<del>0,675</del>	<mark>&lt;0.05</mark>
<del>cell pleomorphism</del>	striking cell pleomorphism	<del>0,502</del>	<mark>&lt;0.05</mark>
spindled cells	spindled cells	<del>0,398</del>	<del>&lt;0.05</del>