

This is the peer reviewed version of the following article:

Electronic eye for the prediction of parameters related to grape ripening / Orlandi, G.; Calvini, R.; Pigani, L.; Foca, G.; Vasile Simone, G.; Antonelli, A.; Ulrici, A.. - In: TALANTA. - ISSN 0039-9140. - 186:(2018), pp. 381-388. [10.1016/j.talanta.2018.04.076]

*Terms of use:*

The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

21/04/2025 23:16

(Article begins on next page)

# Electronic eye for the prediction of parameters related to grape ripening

G. Orlandi<sup>1</sup>, R. Calvini<sup>1,2</sup>, L. Pigani<sup>2,3</sup>, G. Foca<sup>1,2</sup>, G. Vasile Simone<sup>3</sup>, A. Antonelli<sup>1,2</sup>, A. Ulrici<sup>1,2\*</sup>

<sup>1</sup> Dipartimento di Scienze della Vita, Università di Modena e Reggio Emilia, Padiglione Besta, Via Amendola, 2 – 42122 Reggio Emilia

<sup>2</sup> Centro Interdipartimentale BIOGEST-SITEIA, Università degli Studi di Modena e Reggio Emilia, Piazzale Europa, 1 – 42122 Reggio Emilia

<sup>3</sup> Dipartimento di Scienze Chimiche e Geologiche, Università degli Studi di Modena e Reggio Emilia, Via G. Campi, 103 – 41125 Modena

[\\*alessandro.ulrici@unimore.it](mailto:alessandro.ulrici@unimore.it)

## Abstract

An electronic eye (EE) for fast and easy evaluation of grape phenolic ripening has been developed. For this purpose, berries of different grape varieties were collected at different harvest times from veraison to maturity, then an amount of the derived must was deposited on a white sheet of absorbent paper to obtain a sort of paper chromatography. Thus, RGB images of the must spots were collected using a flatbed scanner and converted into one-dimensional signals, named *colourgrams*, which codify the colour properties of the images. The dataset of colourgrams was used to build calibration models to relate the colour of the images with the phenolic composition of the samples – determined by reference analytical methods – and therefore to follow the ripening trend. Satisfactory calibration models were obtained for the prediction of the most important parameters related to phenolic ripening of grapes, such as colour index, tonality, total anthocyanins content, malvidin-3-O-glucoside and petunidin-3-O-glucoside.

## Keywords

Optical sensors; Image analysis; Multivariate calibration; Phenolic ripening; Grapes; Anthocyanins.

## 1 Introduction

1 The maturity level of grapes (*Vitis Vinifera*) at the harvest is the first factor that influences the  
2 quality of the resulting wine [1]. Among the various features, sugar content, pH and acidity levels  
3 are the parameters more frequently used to monitor the maturity level of grapes [2]. However, also  
4 the phenolic composition of grapes plays an important role on the development of several sensorial  
5 attributes of wine, such as colour, body, structure, bitterness and astringency [3].

6 The determination of the parameters related to phenolic ripening is performed by means of classical  
7 analytical procedures, mainly based on UV-Vis spectrophotometry and high-performance liquid  
8 chromatography [4-6]. These methods are very accurate but require expensive instrumentations and  
9 the involvement of skilled personnel. Alternatively, sensory analysis is frequently used to provide a  
10 description of colour, aroma, flavour and texture of the grapes, giving a global characterisation of  
11 the maturity level [7]. However, the sensory evaluation requires a lot of training and experience to  
12 minimize the inherent subjectivity and the variability of tasters [8].

13 For these reasons, nowadays increased efforts are devoted to develop easy-to-use, inexpensive and  
14 objective methods based on artificial sensors [9], known as electronic nose (EN) [10, 11], electronic  
15 tongue (ET) [12, 13] and electronic eye (EE) [14, 15]. These devices are generally used to analyse  
16 the sample 'as it is' or after a minimal manipulation. Then, the sensor output is processed by proper  
17 chemometric techniques that, following a blind-analysis approach, extract the sought information,  
18 such as the amount of specific analytes or the sensory features responsible of smell, taste and colour  
19 of the sample.

20 In a recently published paper, some of us have presented an ET aimed at monitoring grape ripening  
21 [16]. In that work it was shown that, by proper fusion of the data measured with two voltammetric  
22 sensors, it was possible to quantify the parameters related to the technological maturity of grapes,  
23 i.e., pH, total acidity and sugar content, in addition to the anthocyanins content. In the present work,  
24 which was conducted on the same grape samples and can be considered as a follow up from our  
25 previous investigation, we propose the development of an EE sensing system for the prediction of  
26 colour-related parameters, principally suitable to monitor the phenolic maturity.

27 To this aim, purple grape samples were collected at different harvest times from veraison to  
28 maturity, then a drop of the derived must was deposited on a paper sheet to obtain a sort of paper  
29 chromatography. The spots of must were imaged by means of a flatbed scanner, and the resulting  
30 RGB images were analysed using multivariate methods, in order to use the colour-related  
31 information content of the images to predict the phenolic composition of the samples, and therefore  
32 to follow the phenolic ripening trend.

1 To date, few research studies reported the use of image analysis for the evaluation of colour  
2 characteristics and colour changes of grape berries during ripening. For instance, in [17, 18] RGB  
3 images of grape berries have been categorised into different classes according to ripening. More in  
4 detail, considering the histograms of colour-related parameters such as hue or brightness, proper  
5 threshold values have been identified to distinguish grapes into different clusters according to their  
6 colour development (e.g., pre-veraison and post-veraison). In other research papers, the digital  
7 images of grape seeds have been used to investigate CIELAB parameters and morphological  
8 features, such as area, aspect, roundness, length, width and heterogeneity, in order to predict their  
9 maturity stage [19, 20]. Notwithstanding the satisfactory results obtained in these studies, the  
10 proposed methods were limited to the identification of correlations between colour and/or  
11 morphological features and the maturity level of grapes, which was determined based on harvest  
12 time or on visual inspection by expert assessors.  
13  
14  
15  
16  
17  
18  
19  
20

21 In the present work, we followed an alternative approach based on the use of RGB images for the  
22 quantitative prediction of specific analytical parameters related to the colour properties and the  
23 phenolic composition of the grape samples, which could allow to obtain a more detailed and  
24 comprehensive overview of the maturity level.  
25  
26  
27  
28

29 Basically, our method consists in converting each image into a one-dimensional signal, named  
30 *colourgram*, which codifies all the colour-related information and represents a sort of fingerprint of  
31 the corresponding RGB image [21]. The main advantage of this approach is that the whole dataset  
32 of colourgrams can be analysed in the same manner as any other dataset of signals. For instance,  
33 exploratory analysis tools like Principal Component Analysis (PCA) can be used to highlight the  
34 presence of trends, of clusters or of outlier images, whereas multivariate calibration or classification  
35 methods can be used to predict the value of specific parameters or to assign a sample to a specific  
36 class, based on its colour-related characteristics.  
37  
38  
39  
40  
41  
42  
43

44 The colourgram approach has been successfully applied for the automated solution of several  
45 colour-related issues concerning food industry, among which the quantification of defective maize  
46 kernels, related to the presence of mycotoxins [22], the detection of red skin defect of raw hams  
47 [23], the quantification of *Lactobacillus* in fermented milk [24], the prediction of the compositional  
48 and sensory characteristics of pesto sauce [25] and the classification of different pesto brands [21].  
49  
50  
51  
52

53 In this study, after an exploratory analysis of the colourgrams matrix by PCA, multivariate  
54 calibration models were developed using a feature selection/calibration algorithm, namely interval-  
55 Partial Least Squares (iPLS) [26]. The purpose was to define the correlation between the images of  
56 must spots and a series of twelve parameters related to the phenolic composition of the samples.  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 The use of iPLS was particularly profitable for identifying the colourgram features related to the  
2 changes in colour and phenolic composition of must samples during the grape berries ripening.  
3 Moreover, by using a proper algorithm [23] the selected features were displayed in the original  
4 image domain, allowing to evaluate the relevant colour features automatically selected by iPLS.  
5  
6  
7

## 8 **2 Materials and methods**

### 9 *2.1 Samples*

10 In this study, three Italian purple grape varieties were considered: *Ancellotta* (A), *Lambrusco*  
11 *Marani* (L) and *Malbo Gentile* (M). The samples were collected in Reggio Emilia (Italy) during  
12 vintage 2015.  
13  
14  
15  
16

17 For each variety, grape sampling was conducted on three grapevines (field replicates) in order to  
18 account for the vineyard variability. More in detail, about 100 individual grape berries per  
19 grapevine were randomly gathered for each one of 5 subsequent harvest times (T0, T1, T2, T3, T4)  
20 at about 10-day intervals, starting from veraison and ending at harvest of the mature grapes.  
21 Therefore, 45 grape samples were collected on the whole, resulting from (3 grape varieties × 3 field  
22 replicates × 5 harvest times) and were immediately carried to the laboratory under refrigerated  
23 conditions. Each grape sample was crushed into a falcon tube under nitrogen atmosphere to prevent  
24 the oxidation of phenolic compounds. The crushed berries were left to macerate for 60 minutes at  
25 4°C in the dark, then centrifugation at 4000 rpm for 15 minutes was performed (refrigerated  
26 centrifuge 4237R, ALC, Cologno Monzese, MI). The supernatant –called “must” from here  
27 onwards– was divided into two different aliquots to perform replicate determinations.  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37

38 Each aliquot was stored at -20 °C and unfrozen just before spectrophotometric and chromatographic  
39 analyses, that were performed in parallel with image acquisition. All the analyses were replicated  
40 twice for each must sample: in the first measurement session the acquisition order was randomized,  
41 and the first aliquot of each sample was analysed. Then, the order was shuffled again and the  
42 second aliquot of each sample was analysed. The overall number of analyses was therefore equal to  
43 90 (3 grape varieties × 5 harvest times × 3 field replicates × 2 analytical replicates).  
44  
45  
46  
47  
48  
49  
50

### 51 *2.2 Determination of parameters related to phenolic ripeness*

52 The following twelve parameters were measured by means of spectrophotometric and  
53 chromatographic assays:  
54  
55

- 56 - total flavonoids content (TF) was determined by UV spectroscopy as reported in the  
57 literature [27]: after a proper dilution, the absorbance of the sample was measured at 280 nm  
58 and TF was expressed as mg of (+) catechin/L. All the UV-Vis measurements were  
59  
60  
61  
62  
63  
64  
65

1 performed by means of a Perkin Elmer Lambda 650 spectrophotometer using a 10 mm  
2 quartz cuvette as sample holder. Before spectrophotometric analysis the samples were  
3 diluted 50 times in hydrochloric acid-ethanol solution (ethanol:H<sub>2</sub>O:HCl 70:30:1 v/v/v);  
4

- 5 - total anthocyanins content (TAnt) was determined by UV-Vis spectroscopy according to the  
6 method described in [28]: the absorbance value was measured at 540 nm and TAnt was  
7 expressed as mg of oenin chloride/L;  
8
- 9 - colour index (CI) was calculated as the sum of the absorbance values measured at 420 nm  
10 (corresponding to a yellow-orange sample colour), at 520 nm (corresponding to a red-purple  
11 sample colour) and at 620 nm (corresponding to a blue sample colour) [29]. In oenology, CI  
12 is used to evaluate the colour of red wines: if a wine needs a colour correction it is blended  
13 with a different wine up to the desired CI value;  
14
- 15 - optical density values (OD<sub>420%</sub>, OD<sub>520%</sub>, OD<sub>620%</sub>) defined as the percentage  
16 contribution of each absorbance, at 420, 520 and 620 nm, to the colour index [30];  
17
- 18 - tonality (Ton) was calculated as the ratio between the absorbance values measured at 420  
19 nm and at 520 nm. In oenology, Ton is a parameter frequently used to assess the oxidation  
20 of wine during aging [30]. In this case, Ton is used as a parameter suitable to describe the  
21 colour variation which occurs during grape ripening;  
22
- 23 - the five major anthocyanins in the form of 3-O-monoglucoside, i.e. malvidin (Mv-3-glc),  
24 petunidin (Pt-3-glc), peonidin (Pn-3-glc), delphinidin (Df-3-glc) and cyanidin (Cn-3-glc),  
25 were separated and quantified by reverse phase-high performance liquid chromatography  
26 with a diode array detector (RP-HPLC-DAD) following the chromatographic method  
27 described in [31] and adjusted to our equipment, as reported by Vasile Simone and co-  
28 authors [32]. The concentration of the anthocyanins was determined by measuring the  
29 absorbance at 520 nm by Total-Chrom Workstation version 6.2.1 chromatography system  
30 software (PerkinElmer, Inc.), and was expressed as malvidin-3-O-glucoside equivalents.  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45

### 46 2.3 Image acquisition

47 From each aliquot of must sample obtained as described in Section 2.1, 50 µl drops of must were  
48 deposited on A4-sized sheets of white absorbent paper. For the deposition of the must drops, a  
49 precise scheme was used: 8 drops of each must sample were put on each sheet of absorbent paper  
50 following a 4×4 chessboard scheme, alternated to 8 drops of another randomly chosen sample.  
51 Therefore, each absorbent paper sheet contained 16 spots of two samples, i.e., 8 spots per sample  
52 (as an example, see the image on the left side of Figure 1).  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

Several paper types were tested to choose the most proper one to be used as a substratum for spot deposition. Among them, it was found that the absorbent and coated paper (weight: 120 g/m<sup>2</sup>, thickness: 0.24 mm) gave more homogeneous spots due to the presence of two layers: an upper one, made of cellulose with a high liquid absorption capability, and a lower one, made of polyethylene with a high water resistance that allowed to prevent paper wrinkling.

RGB images of the paper sheets were acquired immediately after drops deposition using a CanoScan Lide 220 scanner and saved in JPEG format with a spatial resolution of 4960 × 7015 pixels and an average file size equal to 2 MB. The image scene included also a set of standard colour references used for the subsequent image standardization.

## 2.4 Image elaboration

The key steps followed for the elaboration of the RGB images are illustrated in the right side of [Figure 1](#) and can be summarized as follows:

### 2.4.1 Internal calibration

The RGB images were firstly standardized by means of an internal calibration procedure, in order to reduce possible variability among images over time, mainly due to instrumental instability. The correction procedure reported here is derived from the approach already developed by some of us for the elaboration of hyperspectral images [33]. For each image, eight coloured squares of the standard colour references were considered as regions of interest (ROIs), and a random sampling procedure was used to divide the pixels of each ROI into three groups, in order to account for possible spatial variability of the coloured squares. For each pixel group, the corresponding median values of the R, G and B channels were calculated, then they were stored in a matrix with size equal to {24, 3}, i.e., including the 24 median values (= 3 pixel groups × 8 ROIs) for each one of the 3 channels.

The internal calibration step was carried out by computing, for each channel  $c$ , a regression model between the median vector extracted from the first acquired image, that was considered as the master image ( $M$ ), and the median vector of each one of the remainder “slave” images ( $S$ ). To select the most appropriate order of the regression model, the values of the first- and second-order regression coefficients were estimated, together with the corresponding statistical significances. Based on the results, for each slave image the quadratic models were finally considered, which can be expressed as:

$$M(c) = b_0(i, c) + b_1(i, c) \times S_i(c) + b_2(i, c) \times S_i^2(c) \quad (1)$$

1 where  $b_0$ ,  $b_1$  and  $b_2$  are the regression coefficients calculated for the channel  $c$  of the  $i$ -th slave  
2 image,  $S_i$ .

3 Then, the regression coefficients calculated using the standard colour references were used to  
4 standardize each pixel of the corresponding image of the must spots, according to the equation:  
5  
6

$$7 \quad 8 \quad 9 \quad \text{img}_{corr}(i, c) = b_0(i, c) + b_1(i, c) \times \text{img}_{orig}(i, c) + b_2(i, c) \times [\text{img}_{orig}(i, c)]^2 \quad (2)$$

10 where  $\text{img}_{corr}(i, c)$  is the channel  $c$  of the  $i$ -th standardized image,  $\text{img}_{orig}(i, c)$  is the corresponding  
11 channel of the original image, and  $b_0(i, c)$ ,  $b_1(i, c)$ ,  $b_2(i, c)$  are the regression coefficients calculated  
12 by equation 1.  
13

#### 14 2.4.2 Cropping

15 After internal calibration, for each image each spot of must deposited on the paper was separately  
16 cropped to recover an image of size equal to  $900 \times 900$  pixels. The cropping procedure generated a  
17 dataset of 720 images (= 3 grape varieties  $\times$  3 field replicates  $\times$  5 harvest times  $\times$  8 spots  $\times$  2  
18 measurement sessions).  
19

#### 20 2.4.3 Spot segmentation

21 Before converting all the images into the corresponding colourgrams, a preliminary inspection of  
22 the frequency distribution curves of the parameters included in the colourgram was conducted on a  
23 representative subset of images. This inspection evidenced that the pixels with values of relative red  
24 (rR, see paragraph 2.4.4 for definition) lower than 0.34 correspond to the background (white paper  
25 sheet). Therefore, the colourgram of each image was calculated after background removal  
26 (segmentation), i.e., considering only the pixels of the spots which present rR values greater than  
27 0.34.  
28

#### 29 2.4.4 Conversion into colourgrams

30 After spot segmentation, the images of the individual spots were converted into the corresponding  
31 colourgrams, following the procedure reported in [21].  
32

33 Basically, the first step of this approach consists in unfolding each RGB image in a two-  
34 dimensional matrix with as many rows as the number of segmented pixels, and 3 columns,  
35 corresponding to the R, G and B channels. For each pixel, a series of additional variables directly  
36 derived by the R, G and B values are calculated: lightness (L), defined as the sum of R, G and B  
37 values; relative colours (rR, rG and rB), which are respectively the ratios between each channel and  
38 L; hue (H), saturation (S) and intensity (I), obtained by converting the RGB data into the HSI colour  
39 space, and the nine score vectors (three for each model) obtained by applying PCA to the raw,  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

meancentered and autoscaled unfolded image data. For each variable, the frequency distribution curve (i.e., a histogram with 256 bins) is calculated, then the histograms of the 19 variables are joined in sequence to form a unique one-dimensional signal, which is then divided by the number of segmented pixels. In the end, the loading vectors and the eigenvalues of the three PCA models are added to the signal, that reaches a final length equal to 4900 points ( $= 256 \times 19 + 36$ ).

The conversion of the image into a one-dimensional signal is useful to gain a considerable data compression, which results in a faster computation time necessary for data elaboration. Indeed, the obtained matrix of these fingerprint-like signals can be analyzed by means of common multivariate analysis techniques, such as PCA or PLS.

On the other hand, the colourgram does not account for the spatial location in the original image domain of the colour-related information extracted by multivariate analysis. To address this issue, a proper algorithm has been implemented to visualize directly in the original image domain the colourgram features of interest for the problem at hand. In other words, it is possible to have a reconstructed image where the pixels related to the colourgram features recognized as interesting by the user are displayed in false colours, while the remainder pixels are represented in black [23].

All the steps followed for image elaboration were performed with routines written *ad hoc* in MATLAB language (ver. 7.12, The Mathworks Inc., USA). In particular, the correction of the images and the subsequent conversion into colourgrams have been performed with specific Graphical User Interfaces (GUI): *RGB Image Correction GUI* and *Colourgrams GUI*, which are available at [www.chimslab.unimore.it](http://www.chimslab.unimore.it).

## 2.5 *Multivariate analysis of colourgrams*

PCA was firstly applied to the meancentered colourgrams matrix as an exploratory data analysis tool suitable to recognize similarities and differences among the images of must spots, also highlighting the possible presence of outliers.

Then, multivariate calibration models were calculated on the meancentered colourgrams matrix to quantitatively determine the values of the twelve considered parameters (Y variables). To this aim, the 720 colourgrams were split into a training set, used to calculate the models, and a test set, used for validation. In order to avoid overfitting, the splitting was conducted in a way to maintain together the colourgrams derived from replicate images: 480 colourgrams corresponding to 60 samples belonging to two field replicates were included in the training set, and the remainder 240 colourgrams corresponding to 30 samples belonging to the remaining field replicate were included in the test set.

1 The optimal number of Latent Variables (LVs) used to build each model was chosen by minimizing  
2 the Root Mean Square Error in Cross-Validation (RMSECV). A venetian-blinds cross-validation  
3 was performed considering 2 deletion groups, each one containing the signals corresponding to one  
4 field replicate.  
5

6  
7 The information brought by the different “peaks” (i.e., the frequency distribution curves) of the  
8 colourgram is partially redundant, since all those variables are derived from the RGB values of the  
9 pixels in the original image. Therefore, the selection of the most relevant descriptors from such a  
10 wide number of features could be useful to improve the performance of the calibration models and  
11 to reduce the time needed for image analysis in the perspective of an industrial application.  
12 Therefore, interval-PLS (iPLS) in the *forward* mode was used as a variable selection/calibration  
13 method on the meancentered colourgram matrix [26]. Basically, *forward* iPLS is an algorithm  
14 conceived to divide the signal into a certain number of intervals of equal width defined by the user.  
15 By iteratively adding one interval at a time, the algorithm selects the most effective intervals for  
16 calibration as the ones that, when added, produce a significant decrease of RMSECV. In this work,  
17 6 iPLS models for each Y variable were calculated considering different interval widths,  
18 corresponding to 256, 128, 64, 32, 16 and 8 variables.  
19

20 The performance of the calibration models are reported in terms of the Root Mean Square Error in  
21 calibration (RMSEC), in cross-validation (RMSECV), and in prediction (RMSEP), together with  
22 the coefficient of determination in calibration ( $R^2_{\text{Cal}}$ ), in cross-validation ( $R^2_{\text{CV}}$ ), and in prediction  
23 ( $R^2_{\text{Pred}}$ ). The RMSE and  $R^2$  values were calculated using the equations reported in [16].  
24

25 The PCA, PLS and iPLS models were calculated using the PLS Toolbox (ver 8.1.1, Eigenvector  
26 Research Inc., USA) running in the MATLAB environment (ver. 7.12, The Mathworks Inc., USA).  
27

## 28 **3 Results and discussion**

### 29 *3.1 Exploratory data analysis*

30 The PCA model calculated on the colourgrams matrix highlighted a trend according to harvest time  
31 that was clearly visible along the PC1 scores, as reported in Figure 2a. In general, it can be observed  
32 that PC1 values generally increase for the different grape varieties according to the harvest time.  
33 However, considering the single grape varieties, it is possible to recognise some minor clusters  
34 formed by samples picked in different time periods. The comparison of the PC1 score distribution  
35 with sample images collected at the five harvest times (Figure 2b), highlights that actually the  
36 observed clustering involves groups of spots having similar colour-related characteristics, i.e., that  
37

1 there is a separation between spots having a lighter colour and spots having a more intense colour.  
2 For instance, the colourgrams derived from *Ancellotta* samples gather in two clusters, the first one  
3 composed of the light colour samples collected at T0 and T1, and the second one composed of the  
4 darker colour samples collected at T2, T3 and T4.  
5

6  
7 Moreover, the colour evolution of the must spots during grape ripening is different for the distinct  
8 varieties.  
9

### 10 11 12 3.2 *interval-PLS calibration models*

13 For each Y variable 6 iPLS models were calculated considering different interval widths. [Table 1](#)  
14 reports the results of the best calibration models selected in cross-validation, considering the lowest  
15 RMSECV value.  
16

17  
18 The overall best calibration model was obtained for the total amount of anthocyanins ( $R^2_{\text{pred}} = 0.94$ ).  
19 This model has been built using only 416 variables (26 intervals made of 16 variables) out of 4900  
20 original variables. The selected regions include variables belonging to several distribution curves of  
21 the colourgram, i.e., R, G, B, L, Rr, S, V, the score vectors of PC1-raw, PC3-raw, PC1-  
22 meancentred, PC1-autoscaled and PC2-autoscaled. The actual values versus the predicted values of  
23 TAnt for the test set samples are reported in [Figure 3a](#).  
24

25  
26 The performance of the calibration model obtained for the prediction of TF were less good, even  
27 though acceptable ( $R^2_{\text{pred}} = 0.67$ ). As a matter of fact, flavonoids are a large family of hydroxylated  
28 polyphenolic compounds: the major group consists of anthocyanins (red-purple pigments), but  
29 many other compounds belonging to the family of flavonoids are weakly coloured or colourless,  
30 such as flavonols (pale yellow pigments) and flavanols (colourless pigments that become brown in  
31 case of oxidation) [34]. Therefore, RGB images are more suitable to accurately predict the  
32 anthocyanins content, which increases during the maturation, strongly influencing the red-purple  
33 grape colour and, consequently, the colour of the resulting must.  
34

35  
36 Colour index is the most useful attribute to evaluate the overall colour of this kind of samples. For  
37 CI, a high predictive performance ( $R^2_{\text{pred}} = 0.91$ ) was obtained considering only 576 variables, that  
38 were selected using an interval width of 32 variables. The most part of the selected regions belong  
39 to the same frequency distribution curves selected for the calibration of TAnt. In this case, variables  
40 from the histograms of R, G, B, L, H, S, V, score vectors of PC1-raw, PC3-raw, PC1-meancentred,  
41 PC2-meancentred, PC1-autoscaled and PC2-autoscaled were selected. This fact indicates that few  
42 regions of the colourgram are sufficient to have an accurate prediction of both TAnt and CI, which  
43 are among the most important parameters to monitor the phenolic ripening of grapes.  
44  
45

1 Satisfactory models were also obtained for the prediction of Ton ( $R^2_{\text{pred}} = 0.88$ ), OD420% ( $R^2_{\text{pred}} =$   
2 0.91) and OD520% ( $R^2_{\text{pred}} = 0.86$ ), whereas the estimate of OD620% was not even remotely  
3 possible. As it was already observed in [16], where analogous results were obtained for the  
4 prediction of this parameter also using the ET, this is likely due to the very low absorbance values  
5 measured at 620 nm, close to the baseline.  
6

7  
8 Concerning the calibration models of the single anthocyanins, the best result was obtained for the  
9 prediction of malvidin (Mv-3-glc,  $R^2_{\text{pred}} = 0.85$ ), which is the most abundant anthocyanin in the  
10 considered grape varieties, and consequently the most responsible one for the red-purple colour of  
11 the corresponding must. Similar results were obtained for the prediction of petunidin (Pt-3-glc,  
12  $R^2_{\text{pred}} = 0.84$ ) and peonidin (Pn-3-glc,  $R^2_{\text{pred}} = 0.73$ ). On the contrary, the calibration models for the  
13 prediction of delphinidin (Df-3-glc,  $R^2_{\text{pred}} = 0.59$ ) and cyanidin (Cn-3-glc,  $R^2_{\text{pred}} = 0.09$ ) showed a  
14 poor correlation between the amount of these anthocyanins and the colour variation of the images of  
15 must during the grape ripening.  
16

17  
18 Finally, it must be underlined that in this study the calibration models were calculated considering  
19 as separate items the eight colourgrams corresponding to the images of the eight spots obtained for  
20 each aliquot of must sample. In this manner, it was possible to evaluate the repeatability of the  
21 procedure through the estimate of the within-sample variability [22], i.e., of the variability of the  
22 spots of the same must sample. However, for an application on a real case scenario, the estimates of  
23 the parameters would be obtained as the average of the values predicted from images of multiple  
24 spots of the same sample. As an example, considering the best calibration model obtained for TAnt,  
25 **Figure 3a** reports the predictions for the single spots of the test set samples, while **Figure 3b** shows  
26 the corresponding average values.  
27

28  
29 Therefore, for completeness, **Table 1** also reports the values of aRMSEP and  $aR^2_{\text{Pred}}$ , which  
30 correspond to the RMSEP and  $R^2_{\text{Pred}}$  values calculated considering the average of the eight  $y$  values  
31 estimated from the eight spots of each sample. As one might expect, the predictive performance of  
32 the calibration models obtained after averaging the results are improved.  
33

### 3.3 *Spatial interpretation of the results by means of images reconstruction*

34  
35 When using PLS-based algorithms for calibration or classification purposes, it is a common practice  
36 to look at the Variable Importance in Projection (VIP) plots to identify the most relevant variables.  
37 Indeed, the variables having VIP scores higher than a threshold value, generally set to 1, are  
38 considered significant for the model [35]. The comparison of the VIP scores of the iPLS models  
39 calculated for the prediction of TAnt, CI and Mv-3-glc (**Figure 4**) shows that the most important  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 colourgram region is the same for the three models, and corresponds to the score vector of PC3  
2 obtained by applying PCA to the raw unfolded RGB matrix (SC3RAW).

3 In order to interpret the results of the models, the original images were reconstructed by considering  
4 only the pixels falling in the selected interval of SC3RAW. Figure 5 reports the reconstruction of  
5 five images for each grape variety at the different maturity levels, from T0 to T4, together with the  
6 corresponding original segmented images. It is possible to notice a common trend regardless of the  
7 grape variety. In fact, the number of reconstructed pixels increases in accordance with the degree of  
8 grape maturity: at T0 only few pixels belonging to the must spot appear in the reconstructed image,  
9 while at T4 most of the spot pixels of the original image are present in the corresponding  
10 reconstructed image. Therefore, the informative colour-related parameters, identified by means of a  
11 blind approach, correspond to the pixels related to high values of both colour index and total  
12 anthocyanins content, that is mainly due to malvidin-3-O-glucoside.

13 A final remark concerns the time needed for computation: the conversion of one image into the  
14 corresponding colourgram and the prediction of the considered parameters require only few  
15 seconds, and the whole procedure can be easily automated. Conversely, the determination of the  
16 same parameters related to grape ripening by means of classical analytical methods would require  
17 much longer times.

#### 31 **4 Conclusions**

32 In the present paper we investigated the possibility of using an electronic eye, consisting of a  
33 common flatbed scanner, to monitor the phenolic ripening of grapes. This electronic eye allowed to  
34 obtain acceptable to satisfactory predictions for nine out of twelve parameters related to phenolic  
35 ripening, based on multivariate analysis of the images of grape musts spots.

36 The most interesting outcome is that these results are complementary to those obtained on the same  
37 samples using an ET sensing system [16]: indeed, the EE is suitable to predict parameters related to  
38 the phenolic ripeness (anthocyanins), while the ET is suitable to predict parameters related to the  
39 technological ripeness (sugars and acidity). Therefore, the original idea behind these studies  
40 becomes realistic: to develop a fast, inexpensive and eco-friendly method, based on a combined  
41 device, which is able to easily quantify the parameters related to technological and phenolic  
42 maturity. To this aim, further investigations are being conducted to condense the information  
43 brought by EE and ET through the use of different data fusion strategies.

44 More in general, the EE-based analytical system developed in the frame of the present study could  
45 be easily adapted for the quantitative and/or qualitative assessment of colour-related properties of  
46 liquid samples. Indeed, the proposed approach could be extended to all those situations in which the  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 colour of a solution is strictly related to the presence of one or more analytes of interest. Therefore,  
2 further applications could be not necessarily limited to the food industry, e.g. for the analysis of  
3 other beverages like wine, fruit juice, beer and coffee, but also in other contexts; for example, after  
4 proper adjustments, this approach could be also helpful for the analysis of specimens of forensic or  
5 clinical interest, among others.  
6  
7  
8  
9

### 10 **Conflict of interest**

11 The authors declare that they have no conflict of interest.  
12  
13  
14  
15

### 16 **Acknowledgements**

17 The authors acknowledge financial support for this research by University of Modena and Reggio  
18 Emilia through FAR 2014. The Authors are grateful to “Istituto Superiore A. Zanelli”, Reggio  
19 Emilia – Italy, for providing grape samples.  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

## References

- [1] A.A. Kader, Fruit maturity, ripening, and quality relationships, *Acta Hortic.* 485 (1999) 203–208.
- [2] D.I. Jackson, P.B. Lombard, Environmental and Management Practices Affecting Grape Composition and Wine Quality - A Review, *Amer. J. Enol. Viticult.* 44 (1993) 409–430.
- [3] R. Gawel, Red wine astringency: a review, *Austr. J. Grape Wine R.* 4(2) (1998) 74–95.
- [4] E. Obreque-Sliera, A. Peña-Neiraa, R. López-Solísb, A. Cáceres-Mellaa, H. Toledo-Arayab, A. López-Riverab, Phenolic composition of skins from four Carmenet grape varieties (*Vitis vinifera* L.) during ripening, *LWT - Food Sci. Tech.* 54(2) (2013) 404–413.
- [5] E. Ferrari, G. Foca, M. Vignali, L. Tassi, A. Ulrici, Adulteration of the anthocyanin content of red wines: Perspectives for authentication by Fourier Transform-Near InfraRed and 1H NMR spectroscopies, *Anal. Chim. Acta* 701(2) (2011) 139–151.
- [6] R. Ferrer-Gallego, J.M. Hernández-Hierro, J.C. Rivas-Gonzalo, T.M. Rivas-Gonzalo, Determination of phenolic compounds of grape skins during ripening by NIR spectroscopy, *LWT – Food Sci. Tech.* 44(4) (2011) 847–853.
- [7] M. Le Moigne, C. Maury, D. Bertrand, F. Jourjon, Sensory and instrumental characterisation of Cabernet Franc grapes according to ripening stages and growing location, *Food Qual. Pref.* 19(2) (2008) 220–231.
- [8] J.M. Murraya, C.M. Delahuntyb, I.A. Baxtera, Descriptive sensory analysis: past, present and future, *Food Res. Int.* 34(6) (2001) 461–471.
- [9] M. Śliwińska, P. Wiśniewska, T. Dymerski, J. Namieśnik, W. Wardencki, Food Analysis using artificial senses, *J Agric Food Chem.* 62(7) (2014) 1423–48.
- [10] A. Loutfi, S. Coradeschi, G.K. Mani, P. Shankar, J.B.B. Rayappan, Electronic noses for food quality: A review, *J. Food Eng.* 144 (2015) 103–111.
- [11] J. Gutiérrez, M.C. Horrillo, Advances in artificial olfaction: Sensors and applications, *Talanta* 124 (2014) 95–105.
- [12] Y. Tahara, K. Toko, Electronic tongues-a review, *IEEE Sens. J.* 13(8) (2013) 6516019, 3001–3011.
- [13] L. Escuder-Gilabert, M. Peris, Review: Highlights in recent applications of electronic tongues in food analysis, *Anal. Chim. Acta* 665(1) (2010) 15–25.
- [14] D. Wu, D.-W. Sun, Colour measurements by computer vision for food quality control - A review, *Trends Food Sci. Tech.* 29(1) (2013) 5–20.
- [15] P. Jackman, D.-W. Sun, Recent advances in image processing using image texture features for food quality assessment, *Trends Food Sci. Tech.* 29(1) (2013) 35–43.
- [16] L. Pigani, G. Vasile Simone, G. Foca, A. Ulrici, F. Masino, L. Cubillana-Aguilera, R. Calvini, R. Seeber, Prediction of parameters related to grape ripening by multivariate calibration of voltammetric signals acquired by an electronic tongue, *Talanta* 178 (2018) 178–187.
- [17] Z. Pothen, S.T. Nuske, Automated assessment and mapping of grape quality through image based color analysis, *IFAC-PapersOnLine.* 49(16) (2016) 72–78.
- [18] F.J. Rodríguez-Pulido, L. Gómez-Robledo, M. Melgosa, B. Gordillo, M.L. González-Miret, F.J. Heredia, Ripeness estimation of grape berries and seeds by image analysis, *Comput. Electron. Agric.* 82 (2012) 128–133.
- [19] F. Avila, M. Mora, C. Fedres, A method to estimate grape phenolic maturity based on seed images, *Comput. Electron. Agric.* 101 (2014) 76–83.

- 1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65
- [20] F.J. Rodríguez-Pulido, R. Ferrer-Gallego, M.L. González-Miret, J.C. Rivas-Gonzalo, M.T. Escribano-Bailón, F.J. Heredia, Preliminary study to determine the phenolic maturity stage of grape seeds by computer vision, *Anal. Chim. Acta* 732 (2012) 78–82.
- [21] A. Antonelli, M. Cocchi, P. Fava, G. Foca, G.C. Franchini, D. Manzini, Automated evaluation of food colour by means of multivariate image analysis coupled to a wavelet-based classification algorithm, *Anal. Chim. Acta* 515 (2004) 3–13.
- [22] G. Orlandi, R. Calvini, G. Foca, A. Ulrici, Automated quantification of defective maize kernels by means of multivariate image analysis, *Food Control* 85 (2018) 259-268.
- [23] A. Ulrici, G. Foca, M.C. Ielo, L.A. Volpelli, D.P. Lo Fiego, Automated identification and visualization of food defects using RGB imaging: Application to the detection of red skin defect of raw hams, *Innov. Food Sci. Emerg. Tech.* 16 (2012) 417–426.
- [24] A. Borin, M. F. Ferrão, C. Mello, L. Cordi, L. C. Pataca, N. Durán, R. J. Poppi, Quantification of *Lactobacillus* in fermented milk by multivariate image analysis with least-squares support-vector machines. *Anal. Bioanal. Chem.*, 387(3) (2007) 1105-1112.
- [25] G. Foca, F. Masino, A. Antonelli, A. Ulrici, Prediction of compositional and sensory characteristics using RGB digital images and multivariate calibration techniques, *Anal. Chim. Acta* 706 (2011) 238–245.
- [26] L. Norgaard, A. Saudland, J. Wagner, J.P. Nielsen, L. Munck, S.B. Engelsen, Interval partial least-squares regression (iPLS): a comparative chemometric study with an example from near-infrared spectroscopy, *Appl. Spectrosc.* 54(3) (2000) 413–419.
- [27] R. Di Stefano, M.C. Cravero, N. Gentilini, Metodi per lo studio dei polifenoli dei vini L'Enotecnico XXV (5) (1989) 83–89.
- [28] R. Di Stefano, M.C. Cravero, Metodi per lo studio dei polifenoli dell'uva Rivista di Viticoltura e di Enologia 2 (1991) 37–45.
- [29] Y. Glories, La couleur des vins rouges. Mesure, origine et interprétation. Partie I, *Connaiss. Vigne Vin.* 18 (1984) 195–217.
- [30] P. Ribéreau-Gayon, Y. Glories, A. Maujean, D. Dubourdieu, *Handbook of Enology Volume 2: The Chemistry of Wine Stabilization and Treatments*, John Wiley & Sons, New York, 2006, p. 178.
- [31] F. Chinnici, F. Sonni, N. Natali, S. Galassi, C. Riponi, Colour features and pigment composition of Italian carbonic macerated red wines, *Food Chem.* 113(2) (2009) 651–657.
- [32] G. Vasile Simone, G. Montevicchi, F. Masino, V. Matrella, S.A. Imazio, A. Antonelli, C. Bignami, Ampelographic and chemical characterization of Reggio Emilia and Modena (northern Italy) grapes for two traditional seasonings: 'saba' and 'agresto', *J. Sci. Food Agric.* 93 (2013) 3502–3511.
- [33] A. Ulrici, S. Serranti, C. Ferrari, D. Cesare, G. Foca, G. Bonifazi, Efficient chemometric strategies for PET-PLA discrimination in recycling plants using hyperspectral imaging, *Chemom. Intell. Lab. Syst.* 122 (2013) 31–39.
- [34] E. Petruzza, E. Braidot, M. Zancani, C. Peresson, A. Bertolini, S. Patui, A. Vianello, Plant flavonoids – biosynthesis, transport and involvement in stress responses, *Int. J. Mol. Sci.* 14(7) (2013) 14950–14973.
- [35] G. Foca, D. Salvo, A. Cino, C. Ferrari, D.P. Lo Fiego, G. Minelli, A. Ulrici, Classification of pig fat samples from different subcutaneous layers by means of fast and non-destructive analytical techniques, *Food Res. Int.* 52 (2013) 185–197.



## Captions to Tables and Figures

**Table 1** – Results of the best iPLS calibration models obtained for the colour-related parameters.

**Figure 1** – Acquisition scheme of the RGB images and key steps followed for the image elaboration.

**Figure 2** – Sample index vs. PC1 score values (a); representative RGB images of Ancellotta, Lambrusco and Malbo Gentile must spots collected at the five harvest times (b).

**Figure 3** – Actual amount of total anthocyanins (TAnt, expressed as mg of oenin chloride/L) vs. predicted amount, for the test set samples. Results obtained considering each single spot (a) and the averages over the eight spots (b).

**Figure 4** –VIP scores plot resulting from the calibration models of TAnt, CI and Mv-3-glc. The zoom corresponds to the SC3RAW variables.

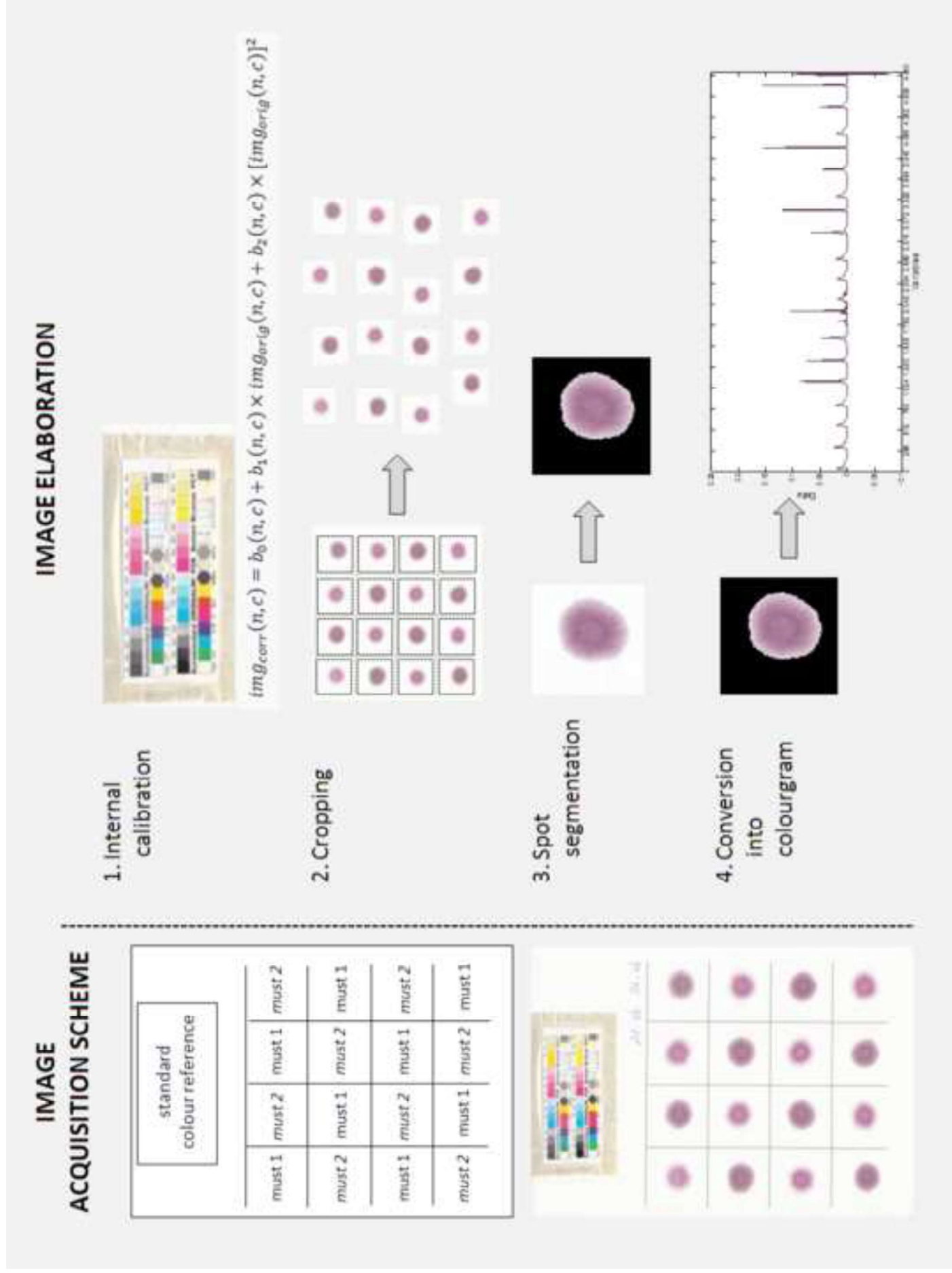
**Figure 5** – Segmented RGB images and corresponding reconstructed images of five must spots at different maturity levels, for each grape variety.

Table1

Y variable	Interv. width	# of vars	# of LVs	RMSEC	RMSECV	RMSEP	aRMSEP	R <sup>2</sup> <sub>Cal</sub>	R <sup>2</sup> <sub>CV</sub>	R <sup>2</sup> <sub>Pred</sub>	aR <sup>2</sup> <sub>Pred</sub>
<b>TF *</b>	8	232	2	18.69	18.91	25.35	25.02	0.86	0.85	0.67	0.68
<b>TAnt *</b>	16	416	10	11.51	15.03	21.78	20.00	0.98	0.97	0.94	0.95
<b>Ton</b>	8	240	6	0.07	0.08	0.12	0.12	0.97	0.96	0.88	0.88
<b>CI</b>	32	576	8	0.57	0.72	1.05	0.99	0.97	0.95	0.91	0.92
<b>OD420%</b>	8	224	9	2.02	2.46	3.41	3.24	0.97	0.96	0.91	0.93
<b>OD520%</b>	8	184	8	3.12	3.26	4.62	4.55	0.95	0.94	0.86	0.86
<b>OD620%</b>	8	192	9	1.29	1.47	1.97	1.88	0.66	0.56	-1.50	-1.29
<b>Df-3-glc *</b>	8	200	8	2.24	2.29	8.69	8.65	0.91	0.90	0.59	0.59
<b>Cn-3-glc *</b>	8	272	8	1.33	1.48	6.86	6.83	0.85	0.81	0.09	0.10
<b>Pt-3-glc *</b>	8	296	8	3.76	3.54	5.92	5.84	0.91	0.92	0.84	0.84
<b>Pn-3-glc *</b>	8	184	9	5.08	5.93	8.37	8.05	0.81	0.74	0.73	0.75
<b>Mv-3-glc *</b>	8	248	8	16.37	19.10	29.04	27.88	0.96	0.95	0.85	0.86

\* values in mg/L

Figure1  
[Click here to download high resolution image](#)



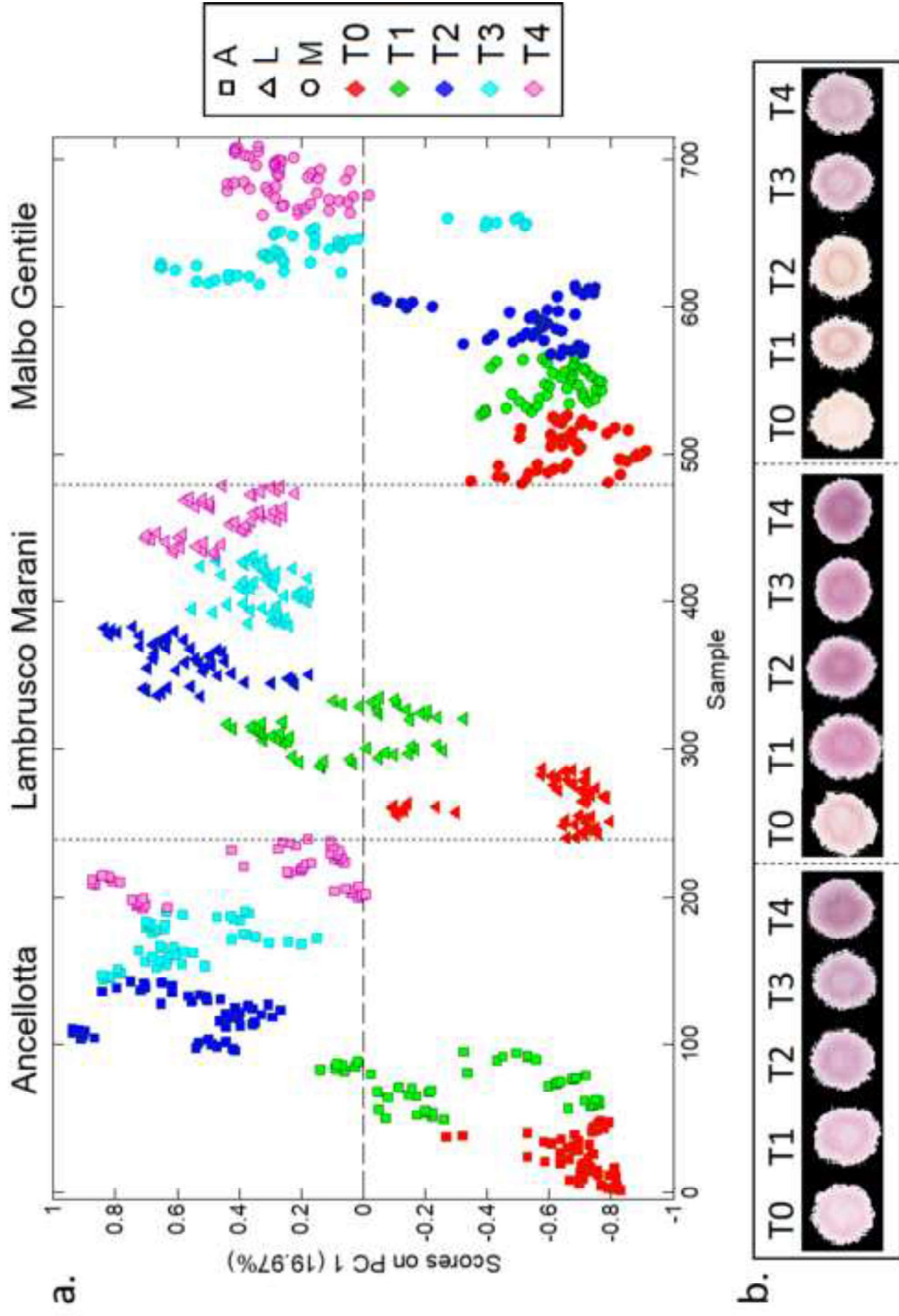


Figure3  
[Click here to download high resolution image](#)

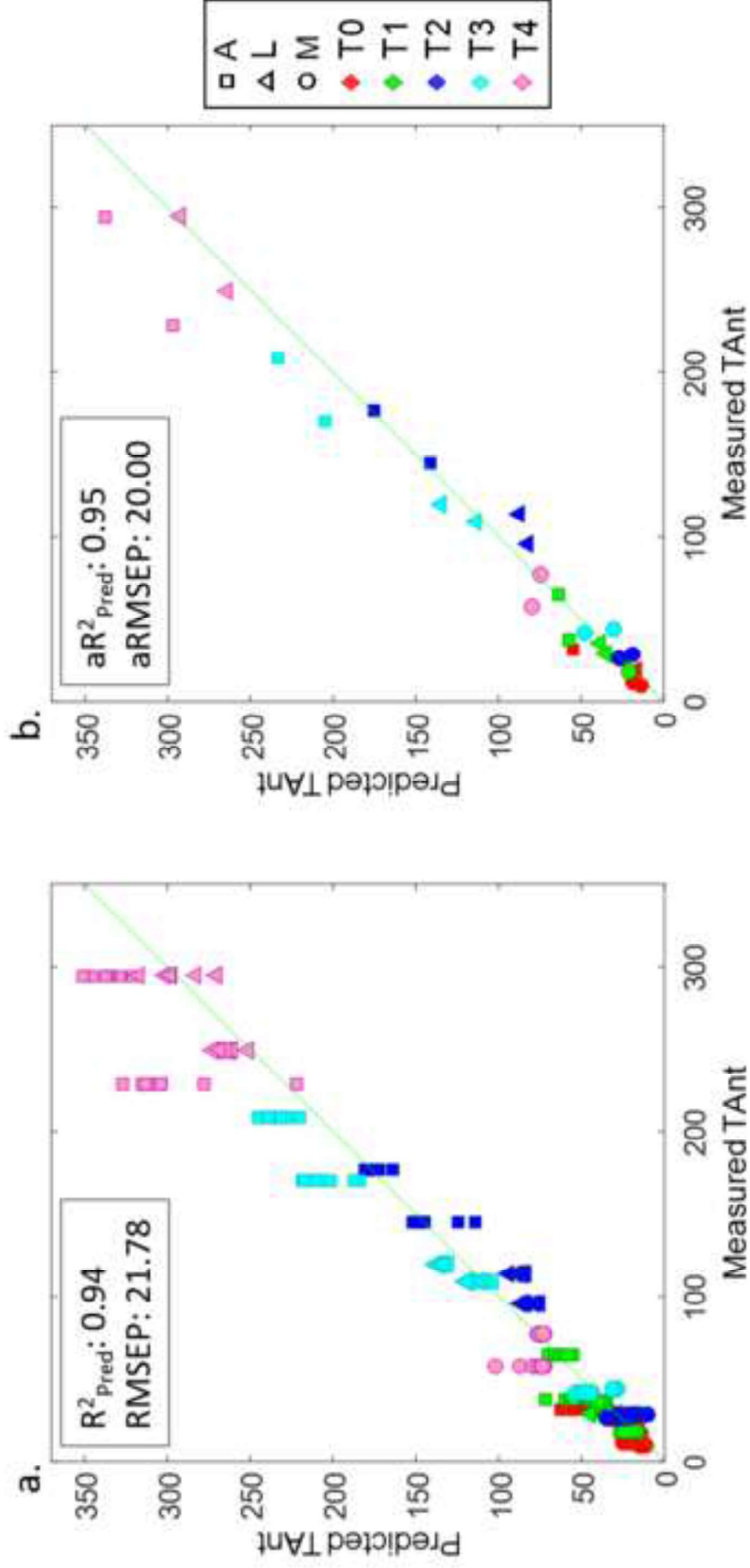


Figure4  
[Click here to download high resolution image](#)

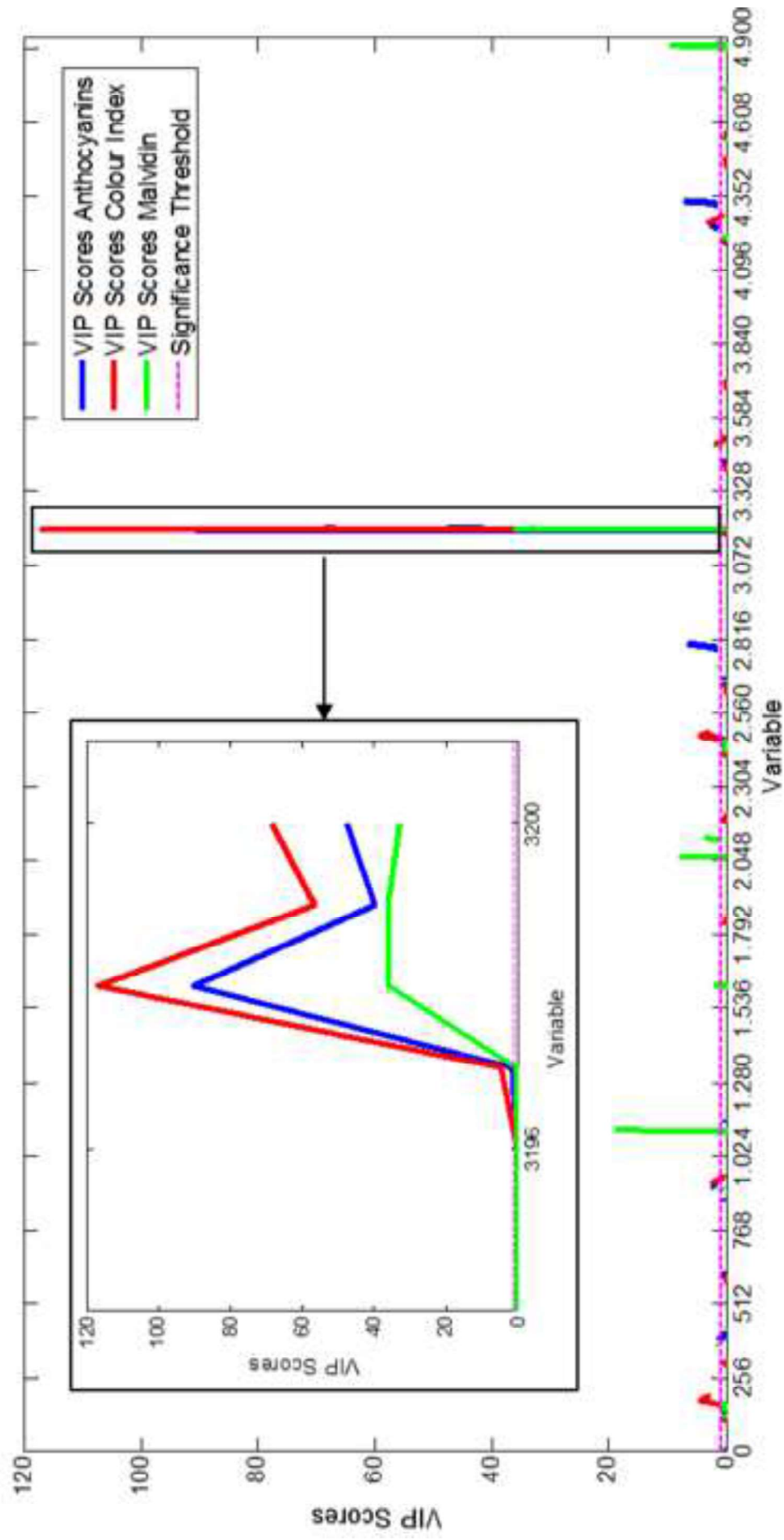


Figure5  
[Click here to download high resolution image](#)

