



Review

The “Hitchhiker’s Guide to the Galaxy” of Endothelial Dysfunction Markers in Human Fertility

Daniele Santi ^{1,2,*} , Giorgia Spaggiari ², Carla Greco ^{1,2}, Clara Lazzaretti ^{1,3}, Elia Paradiso ^{1,3}, Livio Casarini ^{1,4} , Francesco Potì ⁵ , Giulia Brigante ^{1,2} and Manuela Simoni ^{1,2,4}

- ¹ Department of Biomedical, Metabolic and Neural Sciences, University of Modena and Reggio Emilia, 42121 Modena, Italy; carlagreco@unimore.it (C.G.); clara.lazzaretti@unimore.it (C.L.); elia.paradiso@unimore.it (E.P.); livio.casarini@unimore.it (L.C.); giulia.brigante@unimore.it (G.B.); manuela.simoni@unimore.it (M.S.)
- ² Unit of Endocrinology, Department of Medical Specialties, Azienda Ospedaliero-Universitaria of Modena, 41125 Modena, Italy; spaggiari.giorgia@aou.mo.it
- ³ International PhD School in Clinical and Experimental Medicine (CEM), University of Modena and Reggio Emilia, 42121 Modena, Italy
- ⁴ Center for Genomic Research, University of Modena and Reggio Emilia, 42121 Modena, Italy
- ⁵ Department of Medicine and Surgery-Unit of Neurosciences, University of Parma, 43121 Parma, Italy; francesco.poti@unipr.it
- * Correspondence: daniele.santi@unimore.it; Tel.: +39-05-9396-1816



Citation: Santi, D.; Spaggiari, G.; Greco, C.; Lazzaretti, C.; Paradiso, E.; Casarini, L.; Potì, F.; Brigante, G.; Simoni, M. The “Hitchhiker’s Guide to the Galaxy” of Endothelial Dysfunction Markers in Human Fertility. *Int. J. Mol. Sci.* **2021**, *22*, 2584. <https://doi.org/10.3390/ijms22052584>

Academic Editors: Davide Francomano, Gabriele Antonini and Enrique Lledo

Received: 15 February 2021
Accepted: 28 February 2021
Published: 4 March 2021

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Endothelial dysfunction is an early event in the pathogenesis of atherosclerosis and represents the first step in the pathogenesis of cardiovascular diseases. The evaluation of endothelial health is fundamental in clinical practice and several direct and indirect markers have been suggested so far to identify any alterations in endothelial homeostasis. Alongside the known endothelial role on vascular health, several pieces of evidence have demonstrated that proper endothelial functioning plays a key role in human fertility and reproduction. Therefore, this state-of-the-art review updates the endothelial health markers discriminating between those available for clinical practice or for research purposes and their application in human fertility. Moreover, new molecules potentially helpful to clarify the link between endothelial and reproductive health are evaluated herein.

Keywords: endothelium; endothelial dysfunction; male fertility; female fertility; reproduction

1. Sex Differences in Endothelial Function Phenotype

Human fertility is controlled by hormones constituting the hypothalamic–pituitary–gonadal (HPG) axis, aiming to promote gametogenesis in both males and females [1]. The action of sex steroids is strictly connected to the endothelial cell functioning, which is crucial in regulating reproductive system homeostasis. The endothelium is a monolayer of cells forming the inner lining of all blood vessels with protective properties essential to maintain physiological vascular functions upon balancing vasoconstriction and vasodilation stimuli [2]. Dysfunctions of the single-cell layer may contribute to the pathogenesis of vascular disease [3,4] and could lead to detrimental effects for human fertility.

Since the occurrence of cardiovascular diseases (CVD) is higher in men than women [5–7], endothelial dysfunctions could impact fertility in a sex-specific manner and could be indicative of gender-related differences of the hormonal milieu. The risk of stroke, coronary artery and peripheral vascular diseases is very low in young women and increases with age, achieving maximal levels after the menopause [8], similar to those described in men. This gender-related diversity was imputed to estrogens, since both endogenous levels of estradiol and the expression of its receptors differ considerably between sexes [9–12]. In line with this view, during women’s fertile window, the cyclicity of estradiol levels is demonstrated as cardiovascular-protective, inducing total and low-density

lipoprotein (LDL) cholesterol reduction [13], inhibiting LDL oxidation [14], reducing fibrosis, stimulating angiogenesis and vasodilation, and improving mitochondrial functions [15]. Estradiol directly regulates endothelial function and homeostasis inducing vasodilation by activating the transcription of endothelial nitric oxide (NO) synthase (eNOS) and up-regulating NO production [15,16]. Indeed, physiological changes in estradiol serum levels during menstrual cycle are directly related to flow-mediated dilation (FMD) [17]. During menopause, the decline of estrogen levels corresponds to increased CVD risk in females, achieving similar levels in males [18]. Since vascular smooth muscle functionality does not change during menopausal age, these data suggest that increased CVD risk is linked to the loss of sex hormone regulation on endothelial cell functioning [19].

From a molecular point of view, the impact of estrogen on the endothelium could involve three different mechanisms: (i) induction of vasodilator factors such as NO and prostacyclin, (ii) stimulation of endothelial damage repair systems, and (iii) anti-inflammatory and antioxidant properties [20,21] (Figure 1).

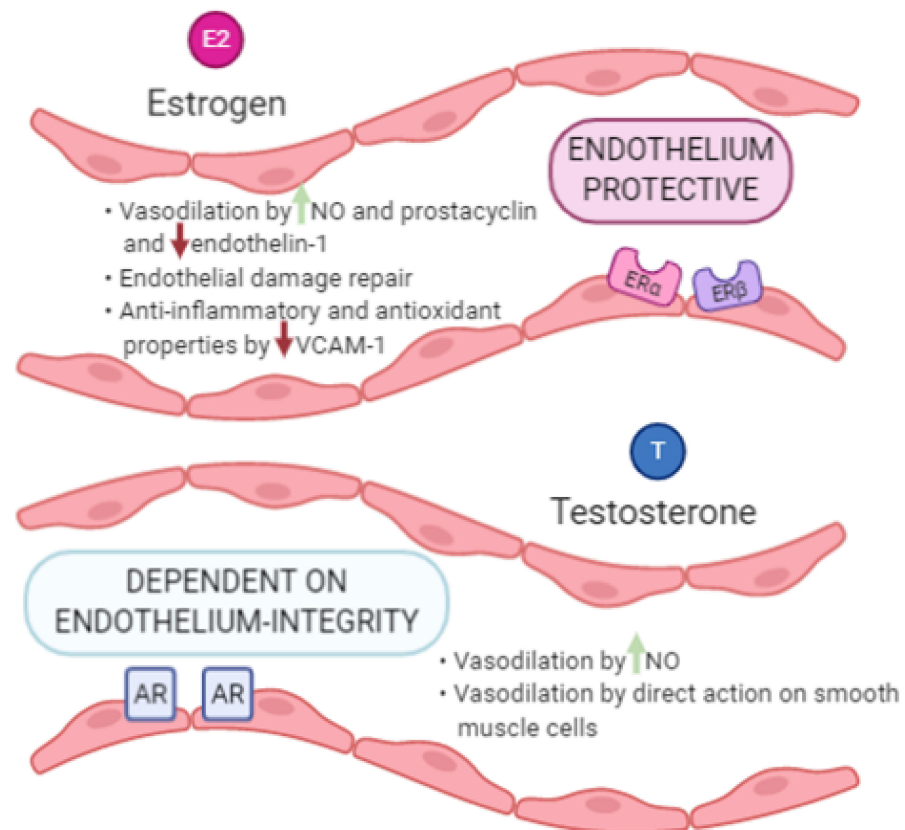


Figure 1. A graphical representation of the synthesis of the estrogens and testosterone actions at endothelial level. Footnote to Figure 1: AR: androgen receptor; ER: estrogen receptor; NO: nitric oxide; VCAM: vascular cell adhesion molecules.

Estrogenic actions are mediated via genomic (long-term) and non-genomic (short-term) pathways depending on the receptor involved [10,21]. Although the estrogen action on endothelial cell is largely studied, the androgens effects are less examined [9]. Testosterone exerts a vasodilator effect via both genomic and non-genomic actions on the endothelial cell [22]. The genomic effect is mediated by the nuclear androgen receptor (AR)-dependent increase in NO production, activating the eNOS [23,24] (Figure 1). The non-genomic effect requires the action of four membrane receptors [25,26], activating the phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT) pathway and eNOS [23,24]. Although the exact molecular mechanisms induced by testosterone on endothelial cells is not completely understood [22], its vascular action is direct and not mediated by estrogens syn-

thesized through testosterone aromatization, as demonstrated *in vitro* [27–31]. These data are strengthened by studies demonstrating the vasodilation effect of dihydrotestosterone (DHT), a non-aromatizable androgen [32,33]. The testosterone-induced vasodilation needs the integrity of the vascular endothelium, while higher concentrations of the hormone are required to induce a dilatory response of dysfunctional endothelium [34]. In summary, sex differences in endothelium functions may be influenced by the action of both estradiol and testosterone, that clearly show sex-specific serum concentrations. Other circulating molecules, such as microRNAs [35–38], contribute to gender-related endothelial functions since they are differently distributed between premenopausal women and age-matched men [39].

This evidence confirms the gender-related difference in endothelial functions/dysfunctions which has to be considered in the context of human fertility. In this review, we summarize evidence available on the interconnection between male and female fertility and endothelial function, with a specific focus on the direct and indirect markers available to measure it.

2. How to Measure Endothelial Dysfunction: Direct and Indirect Parameters of Endothelial Health

2.1. Dynamic Markers

Endothelial dysfunction, expressed as a worsened capacity of the vessels to respond to physical and chemical stimuli, could be evaluated using several direct and indirect parameters aiming at detecting the principal endothelial mediator (i.e., NO). These markers can be measured using both invasive and non-invasive approaches. The angiographic measure of vascular diameter change in response to intracoronary infusions of acetylcholine (ACh) provides the most accurate currently available test [40]. ACh induces the NO release from the endothelial cell, in turn inducing vasodilation. Since the angiographic evaluation is an invasive practice, the non-invasive “ACh test” was developed. This protocol compares the response to an endothelium-independent vasodilator, such as nitroprusside, to that inducible by ACh [41]. Briefly, the venous occlusion plethysmography at forearm is used to compare the blood flow after ACh and sodium nitroprusside infusions [41]. However, since the clinical application of this technique is burdened by high costs and limited availability, further indirect measures have been developed, such as the FMD. This latter consists of the high-frequency ultrasonography measure of the brachial artery dilation after an induced hypoxia [42,43]. Using an arm cuff inflated to supra-systolic blood pressure level, the NO release from endothelial cells mediates relaxation of smooth muscle cells with subsequent vasodilation [42,43]. The FMD is expressed as the difference percentage between arterial diameter measured at baseline and after cuff releasing [42,43]. Since FMD is correlated with the NO concentration [44], measurable during coronary angiography via invasive procedures, FMD assessment remains the most used non-invasive parameter in clinical practice and research [45]. Two recent meta-analyses confirmed the FMD clinical relevance, predicting peripheral arterial disease [46], CVD [47,48], and all-cause mortality [47,49–52].

Alongside vascular elasticity, arterial stiffness is largely demonstrated to be related with CVD and mortality. Thus, the assessment of arterial stiffness in several proximal and distal vascular compartments has been suggested so far [53]. To this purpose, the most reliable parameter is the intima-media thickness (IMT), measured applying the linear probe longitudinally on arteries, searching for a double-line density between the intimal-luminal and the medial-adventitial interfaces [54]. Carotid IMT (cIMT) has been demonstrated to be associated with atherosclerosis, CVD [55,56], and erectile dysfunction. The latter is a precocious marker of endothelial dysfunction [57,58]. Indeed, subjects with erectile dysfunction present higher cIMT and lower FMD than controls, and these parameters may be improved upon vasodilation induced by treatment with phosphodiesterase 5 inhibitors [59–62]. Beyond cIMT, the aortic IMT (aIMT) is considered a potential marker of early atherosclerosis in children, adolescents, and young adults [63]. Interestingly, the aIMT has been found associated to impaired Sertoli cell markers, such as anti-Müllerian hormone and inhibin B, in obese adolescent men with insulin resistance [64]. Together

with cIMT and aIMT, the arterial stiffness could also be measured in brachial (bIMT) and common arteries (caIMT) (Table 1) [65].

Table 1. Markers of endothelial dysfunction applied to clinical practice.

Marker	Target	Test Type	Clinical Application
Ach intracoronary infusions	eNO production: vasodilation	Invasive dynamic	Limited
Non-invasive Ach test	eNO production: vasodilation	Non-invasive dynamic	Limited
FMD	Vasodilation	Non-invasive dynamic	Yes
Carotid IMT	Arterial stiffness	Non-invasive dynamic	Yes
Aortic IMT	Arterial stiffness	Non-invasive dynamic	Yes
Brachial IMT	Arterial stiffness	Non-invasive dynamic	Yes
Common arteries IMT	Arterial stiffness	Non-invasive dynamic	Yes
Pulse wave velocity	Arterial stiffness	Non-invasive dynamic	Limited
Retinal vessels vasodilation assessment	eNO production: vasodilation	Non-invasive dynamic	No
BOLD MRI	Capillary oxygen content	Non-invasive dynamic	Limited
ASL MRI	Tissue perfusion	Non-invasive dynamic	Limited
PC MRI	Blood flow velocity and arterial stiffness	Non-invasive dynamic	Limited
FMD MRI	Vasodilation	Non-invasive dynamic	Limited
Oximetry MRI	Hemoglobin oxygen saturation	Non-invasive dynamic	Limited
Renal resistive index	Arterial elasticity	Non-invasive dynamic	Limited
Dynamic renal resistive index	Arterial elasticity	Non-invasive dynamic	No
LDL	Atherosclerotic plaque development	Biochemical marker	Yes
Oxidative LDL	Atherosclerotic plaque development	Biochemical marker	Yes
Triglycerides	Atherosclerotic plaque development	Biochemical marker	Yes
HDL	Atherosclerotic plaque development	Biochemical marker	Yes
Hs-CRP	Inflammation	Biochemical marker	Yes
Homocysteine	Inflammation	Biochemical marker	Yes
Lipoprotein-associated phospholipase A2	Inflammation	Biochemical marker	Limited

Ach: acetylcholine; ASL: arterial spin labelling; BOLD: blood oxygen level-dependent; eNO: endothelial nitric oxide; FMD: flow-mediated dilation; HDL: high-density lipoprotein; hs-CRP: high sensitivity C-reactive protein; IMT: intima-media thickness; LDL: low-density lipoprotein; MRI: magnetic resonance imaging; PC: phase contrast.

Since the endothelial dysfunction is characterized by reduced endothelial NO production, resulting in increased peripheral arterial resistance [66], pulse wave velocity (PWV) is a further vessels stiffness marker. Carotid-femoral PWV (cfPWV) indicates the velocity of propagation of the arterial blood pressure wave along the vascular wall [67] and represents the gold standard for arterial stiffness assessment [68]. This parameter has been demonstrated to predict organ damage [69] and CVD mortality [70], although it is still poorly applied in clinical practice.

Even magnetic resonance imaging (MRI) has been increasingly used for assessing endothelial functioning [71]. Quantitative, dynamic, non-contrast MRI studies have been

developed, including the blood oxygen level-dependent (BOLD) imaging, the arterial spin labelling (ASL), the phase contrast (PC), the MRI-measured PWV, the luminal FMD, and the dynamic MRI oximetry [71]. These innovative approaches provided the opportunity to measure endothelial dysfunctions in human sites not easily accessible. For instance, MRI may be used for evaluating the NO-mediated retinal vessels vasodilation, as an indirect marker of the vascular elasticity in the microcirculation [72]. The dynamic MRI oximetry is applied to assess the flicker light-induced retinal vasodilatation after pupil dilatation induced by 1% topical tropicamide. The vasodilation is indicated as the percentage change in retinal vessel diameter from the baseline [73]. Moreover, the arterial elasticity could be measured in the kidney by the duplex ultrasound-derived parameter “renal resistive index” (RI) [74] and the dynamic renal RI [75]. The latter provides the RI measure after sublingual nitrate administration and is correlated to endothelial function and arterial stiffness [75].

2.2. Biochemical Markers

Impairment of endothelial function correlates with all cardiovascular risk factors, in particular, dyslipidemia. Oxidized low-density lipoprotein (ox-LDL) would stimulate the endothelial secretion of adhesion molecules and chemoattractant proteins, enhancing the atherosclerosis plaque development [76,77]. Similarly, LDL and triglycerides have been associated to endothelial dysfunction in both males and females [78]. Accordingly, cholesterol-lowering therapies have been demonstrated to improve endothelial vasomotor function of brachial arteries analyzed by FMD [79–81]. On the other hand, high-density lipoprotein (HDL) is known to exert endothelial protective roles through the apolipoprotein A-I receptor [82], mediating the cholesterol and phospholipid efflux and reducing the atherogenic stimulus [83,84]. Moreover, HDL stimulates vasodilation via NO release, which underlies the activation of eNOS by the scavenger receptor B type I [85], and the transport of bioactive lysophospholipids regulating vascular tone via sphingosine-1-phosphate 3 receptor [86–89]. Thus, the measurement of these various components of lipid homeostasis could serve as an indirect marker of endothelial dysfunction.

C-reactive protein (CRP), a hepatic acute-phase protein, is considered a prognostic factor for major CVD [90,91]. Moreover, increased CRP serum levels have been associated with the Ach test impairment [92]. Indeed, CRP decreases eNOS expression and bioactivity in human aortic endothelial and smooth muscle cells [93,94]. Accordingly, the CRP reduction correlates with improvement of the endothelial function [92]. In addition, high-sensitivity CRP (hs-CRP), a sensitive and more precise biomarker of inflammation, provides results associated with a higher risk of atherosclerosis and all-cause and CVD mortalities [95].

Homocysteine is a sulfur-containing, non-proteinogenic amino acid biosynthesized during the metabolism of the essential amino acid methionine [96]. Hyperhomocysteinemia is an independent risk factor for atherosclerosis, stroke, CVD [96,97], and endothelial dysfunction [98]. Accordingly, several studies have reported the correlation between total homocysteine concentration and FMD [99], cIMT [100], and cFPWV [101]. Indeed, high homocysteine plasma levels presumably affect NO bioavailability and impair endothelial-dependent vasodilation [102,103]. Thus, the panel of biochemical indirect markers of endothelial dysfunction available in clinical practice should combine the lipid asset with the measurements of hs-CRP and homocysteine (Table 1).

A recently proposed marker of endothelial dysfunction is the lipoprotein-associated phospholipase A2 (Lp-PLA2), an inflammatory mediator produced by macrophages in atherosclerotic plaques. Physiologically, about 80% of Lp-PLA2 circulates bound to LDL, whereas the remaining 20% to HDL. Lp-PLA2 levels are elevated in patients with endothelial dysfunction [104], suggesting they might have a pro-atherogenic role [105]. Lp-PLA2 hydrolyses ox-LDL to LDL, generating pro-inflammatory and pro-atherogenic molecules promoting vascular inflammation and atherosclerotic plaque development [105]. These data were corroborated by a clinical study enrolling a group of 172 subjects with no significant coronary artery disease [106]. In this study, high concentrations of Lp-PLA2 were

demonstrated as a strong predictor of endothelial dysfunction assessed by the Ach test. However, discordant results have been obtained using FMD [107] and Ach test after the administration of a Lp-PLA2 inhibitor [108]. Thus, these contrasting results, together with the difficult to measure Lp-PLA2, limit the reliability and use of this parameter in clinical practice.

Several other biochemical markers of endothelial dysfunction are currently used exclusively for research purposes, but not in clinical practice. These parameters could be divided according to their pro- or anti-inflammatory properties, as well as vasoconstriction or vasodilation actions, and location in specific tissues, or solubilized in the plasma (Table 2).

Table 2. Markers of endothelial dysfunction commonly used in research setting.

	Soluble Markers	Tissue Markers
Markers of Inflammation	ADAMs (disintegrin and metalloproteinase)	
	ADAMs with thrombospondin motifs (ADAMTS)	
	-13	
	Asymmetric dimethyl arginine (ADMA)	
	Bone morphogenetic protein 4 (BMP4)	Angiotensin converting enzyme (ACE)
	Endothelial progenitor cells (EPCs)	E-selectin
	Glutathione	Intercellular adhesion molecule-1 (ICAM-1)
	Interleukin (IL)-1 β , -6, -7, -8, -12, -15, -18	Metalloproteinases (MMPs)
	Interferon- γ (IFN γ)	P-selectin
	Long non-coding RNA (LncRNAs)	Platelet endothelial cell adhesion molecule (PECAM-1)
	MicroRNAs (miRNAs)	Tissue plasminogen activator (t-PA)
	N-terminal fragment of proatrial natriuretic peptide (NTpro-ANP)	Vascular cell adhesion molecule-1 (VCAM-1)
	Plasminogen activator inhibitor-1 (PAI-1)	Vascular Endothelium (VE)-cadherin
	Reactive oxygen species (ROS)	
	Sphingosine 1-phosphate	
	Soluble ICAM-1 and VCAM-1	
	Soluble P-selectin	
Soluble E-selectin		
Tumor necrosis factor- α (TNF- α)		
Von Willebrand factor (VWF)		
Markers of Vasoconstrictions		ADAMs
		ADAMTS-1, -4, -7, -13, -18
		Advanced glycation end-products (AGEs)
		E-selectin, P-selectin
		Fatty acid binding proteins
		ICAM-1, -2
		Neuropeptide Y receptor
		Thromboplastin (coagulation factor III)
		Tumor endothelial marker 8
		Tissue inhibitor of metalloproteinases (TIMPs)
		Tissue nonspecific alkaline phosphatase
		Tumor necrosis factor receptor 2
		VCAM-1
	VEGF receptor-1	

Table 2. Cont.

	Soluble Markers	Tissue Markers
Markers of Vasodilation	Acetylcholine (Ach)	α 1-subunit of the soluble guanylyl cyclase
	Atrial natriuretic factor (ANF)	Adenosine
	Bradykinin	Aminopeptidase N
	C-terminal pro-arginine vasopressin	Angiotensin receptor (AGTR) 1
	C-terminal endothelin-1 precursor fragment	Endothelin receptor type B
	Endothelial nitric oxide synthase	Endothelial cell-selective adhesion molecule
	Nitric oxide (NO)	IL-1 receptor, IL-13 receptor α 1
	Prostaglandins	Integrin β 1
	Serotonin	Notch
	Soluble VE-cadherin	Platelet endothelial cell adhesion molecule
	Transforming growth factor beta (TGF- β)	Prostacyclin receptor
	Tumor necrosis factor α (TNF α)	VEGF, VEGF receptor 2
		VE-cadherin
		Vasoactive intestinal peptide (VIP) receptor 1,2

Inflammation markers may be informative of the activation state of the vascular endothelium, which, in turn, influences the adhesion and infiltration of leukocytes and the initiation of the coagulation cascade [109]. Moreover, other molecules are used to evaluate the response to oxidative stress and of the endothelium recovery in response to harmful stimuli [109,110]. Vasoconstriction molecules are mainly involved in the renin–angiotensin–aldosterone system, regulating blood pressure, plasma volume, and arterial muscle tone [111,112]. On the contrary, vasodilation molecules are generally related to the molecular paracrine action of NO, bradykinin, and potassium ion channels [113].

3. Endothelial Function and Male Fertility: Endothelial Gene-Related Polymorphisms

The male reproductive system requires the interaction between neurons, blood vessels, and cells of the immune system, which act synergistically with the main actors, such as germ cells, stromal testicular cells, and hormones [114]. In this refined mechanism, a potential endothelium role is suggested. Indeed, a bidirectional relationship occurs between the endothelial function and those hormones involved in reproduction. From one side, the endothelium's role in balancing vasodilation and vasoconstriction is fundamental for reproductive functions. On the other hand, the endothelium is demonstrated to directly secrete or indirectly mediate the production of several molecules able to influence testicular functions [114]. However, the complex relationship between endothelial function and the male reproductive system is still not completely clarified and only few endothelium-related health markers described above have been evaluated in the context of male fertility/infertility. A step forward may be provided by the study of single-nucleotide polymorphisms (SNPs) of genes encoding key factors for endothelial physiology. They might elucidate the potential role of these genes in the male infertility pathogenesis and address some cases of infertility currently defined as “idiopathic”. Table 3 and Figure 2 describe the role and the effect of these potential markers of male infertility.

Table 3. Potential markers of male infertility. The level of evidence is determined through studies available in the literature.

Marker	Category	Effect on Male Fertility	Level of Evidence
c.T786C (rs2070744)	SNP on promoter region of <i>eNOS</i>	Increased NO production, inducing direct oxidative damage	Strong
c.G894T (rs1799983)	SNP on exon 7 of <i>eNOS</i>	Increased NO production, inducing direct oxidative damage	Mild
Variable number of tandem 4a4b repeats (rs61722009)	CNV in intron 4 of <i>eNOS</i>	Increased NO production, inducing direct oxidative damage	Mild
p.Val16Ala (rs4880)	SNP on <i>SOD2</i>	Increased seminal levels of H ₂ O ₂ , resulting noxious for sperm quality	Mild
c.G362A (rs2536512)	SNP on <i>SOD3</i>	Reduce SOD activity	No evidence
c.C262T (rs1001179)	SNP on <i>CAT</i>	Doubled risk of male infertility	No evidence
p.Pro200Leu (rs1050450)	SNP on <i>GXP1</i>	Doubled risk of male infertility	No evidence
IL-8	Pro-inflammatory cytokine	Directly related to leucocytospermia and inversely to ejaculatory volume only in patients with male accessory gland infections	No evidence
IL-7	Pro-inflammatory cytokine	Reduced semen quality	Mild
IL-6	Pro-inflammatory cytokine	Increase of pro-inflammatory mediators in the seminal plasma	Contradictory results
IL-1	Pro-inflammatory cytokine	Increase of pro-inflammatory mediators in the seminal plasma	Contradictory results
IFN- γ	Macrophage-activating cytokine	Reduced semen quality	Mild
TNF- α	Pro-inflammatory cytokine	Increase of pro-inflammatory mediators in the seminal plasma	Contradictory results
ICAM-1 and ICAM-2	Adhesion molecule	The balance between the ICAM-1/soluble-ICAM-1 opposite actions influences the permeability of the blood–testis barrier	Mild
VCAM-1	Adhesion molecule	VCAM-1 facilitates the leukocyte adhesion to the vascular endothelium	Mild
ET-1	Vasoconstriction molecule	ET-1 dysfunction could impair sperm progression through the genital tract	Evidence only in animal models
VEGF-A	Gene with pro- and anti-angiogenic properties	VEGF-A isoforms determines a sperm number reduction	Evidence only in animal models

CAT: catalase; CNV: copy number variants; eNOS: endothelial nitric oxide synthase; ET-1: endothelin-1; GXP1: glutathione peroxidase 1; ICAM: intracellular cell adhesion molecules; IFN: interferon; IL: interleukin; SOD: superoxide dismutase; SNP: single-nucleotide polymorphism; TNF- α : tumor necrosis factor-alpha; VCAM: vascular cell adhesion molecules; VEGF-a: vascular endothelial growth factor-A.

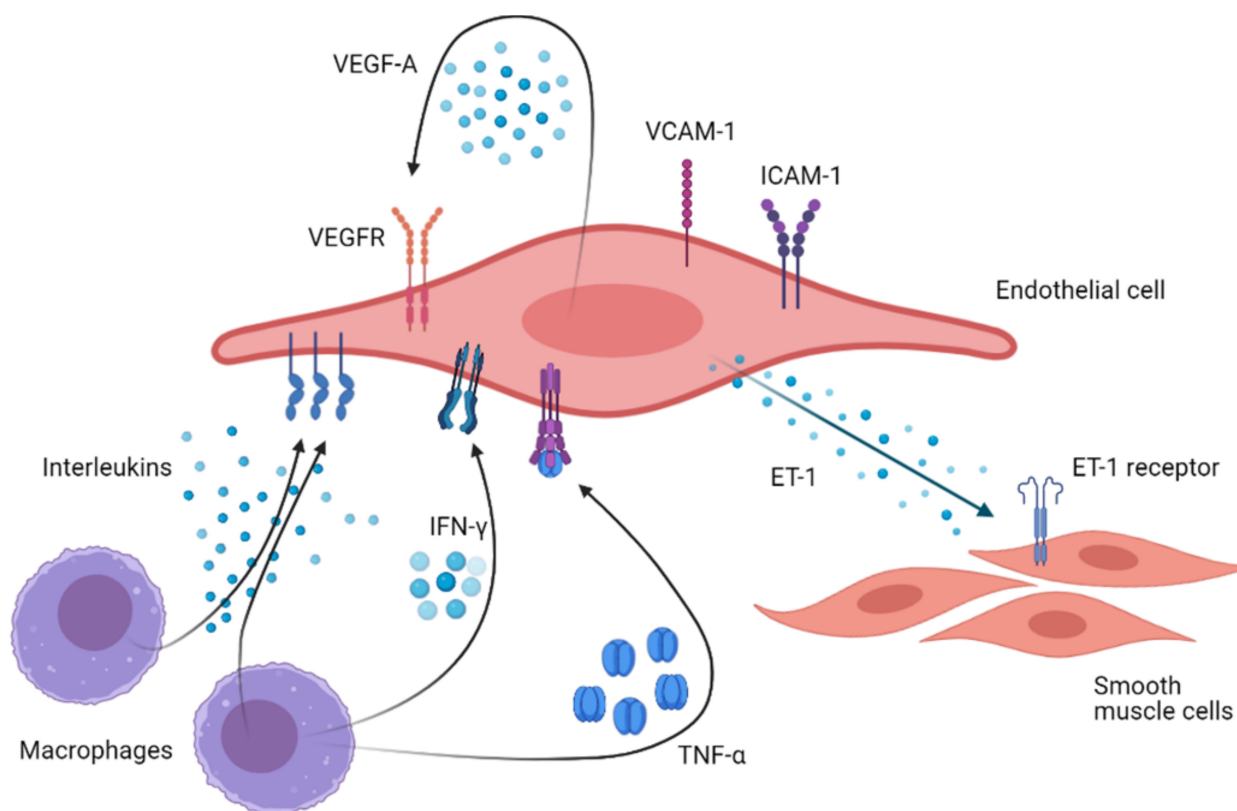


Figure 2. A graphical representation of the paths of endothelial markers involved in male infertility. Footnote to Figure 2: ET-1: endothelin-1; ICAM: intracellular cell adhesion molecules; IFN: interferon; TNF- α : tumor necrosis factor-alpha; VCAM: vascular cell adhesion molecules; VEGF-a: vascular endothelial growth factor-A.

3.1. *eNOS* Gene Polymorphism and Male Infertility

NO is a molecular mediator relevant to many physiological systems, including reproduction [114]. Its production is regulated by NOS [115], available in three different isoforms: eNOS, neural NOS (nNOS), and inducible macrophage NOS (iNOS), depending on the originator cell population [116]. Among the three isoforms, eNOS is the most expressed in male reproductive organs, since it is synthesized in endothelial testicular cells, vas deferens, epididymis, and both Sertoli and Leydig cells [117]. In testicular tissues, eNOS regulates the NO local production, balancing the reactive oxygen species (ROS)/antioxidants ratio [118,119]. This fine mechanism is crucial for maintaining proper testicular functionality, since germ cells are very susceptible to elevated ROS levels [118]. NO is supposed to be inversely related to sperm motility, to attend sperm acrosome reaction and capacitation process, and to regulate the Sertoli/germ cells ratio in the seminiferous epithelium, allowing the production of mature and functional sperms [118,120–122]. It is widely demonstrated that SNPs falling within the *eNOS* gene, located on the chromosome 7q36, modulate the expression of the enzyme and, in turn, NO production [123]. Some of these SNPs, such as the c.T786C (rs2070744), falling in the promoter region, the c.G894T (rs1799983) on exon 7, and a variable number of intron 4 tandem “4a4b repeats” (rs61722009) [124], have been associated with fertility and sperm morphology.

In vitro and epidemiological studies evaluated the association between *eNOS* SNPs and infertility, obtaining inconclusive results [117,124–130]. It is largely demonstrated that the excess in NO production leads to a sperm number reduction, by inducing germ cells apoptosis [117], and a sperm motility impairment, via activation of the s-Guanylyl Cyclase (sGC)-cGMP-protein kinase G (PKG) pathway [118]. This molecular mechanism is supposed to impact RNA stability [131], although the increased NO production is directly

linked to oxidative damage of lipids, proteins, and DNA of sperm cells [132]. The *eNOS* SNPs c.T786C and 4a4b would modulate the activation of this signalling pathway and were identified as risk factors for male infertility, due to their association with higher NO production [117]. However, another study found that a specific c.T786C and c.G894T allelic variant was significantly associated with impaired sperm number, motility, morphology, and seminal glutathione peroxidase [128]. A recent meta-analysis detected only c.T786C *eNOS* SNP as a predictor of semen alterations, i.e., sperm concentration and motility, in a population of 3507 men [123,132]. Despite the existence of pathophysiological rationale and experimental evidence, rigorous association studies and functional demonstrations of the association between *eNOS* SNPs and infertility are still lacking. Therefore, these missing points prevent the clear comprehension of the *eNOS* variants' role in spermatogenesis and their clinical implications.

3.2. Antioxidant Systems Gene Polymorphisms and Male Infertility

ROS are free radicals physiologically required in seminal plasma to promote sperm capacitation, acrosome reaction, and sperm–oocyte fusion. These effects are strictly dose-dependent, since relatively high ROS concentrations in seminal plasma induce detrimental consequences, such as sperm DNA damage, lipid peroxidation, membrane fluidity reduction and sperm motility impairment [133,134]. Effects of seminal ROS may be neutralized by the homeostatic antioxidant systems, to which belong manganese superoxide dismutase (SOD2), catalase (CAT), glutathione peroxidase 1 (GXP1), and glutathione S transferase (GST) [135]. This complex enzymatic kit was found also in endothelial cells [136]. In particular, SOD2 catalyzes the detoxification of superoxide radicals in H₂O and O₂ in mitochondria [136,137]. Although the gene sequences encoding these enzymes are phylogenetically conserved, SNPs could modulate their expression levels [136] and, consequently, interfere with the ROS/antioxidants balance, influencing male fertility [136,137].

Several studies have found an association between *SOD2* gene SNPs and male infertility. The *SOD2* promoter SNP -262C > T “T” allele (rs4880) was recently demonstrated to be more frequent in infertile men than in the control group [135]. Moreover, the incidence of the Val16Ala “Val” homozygous allele is higher in infertile than fertile men, suggesting that the “Ala” carriers have higher seminal H₂O₂ levels and lower sperm quality [138]. Similar associations were achieved by the evaluation of the *SOD2* SNPs p.Val16Ala [138] and p.Ile58Thr [139]. However, the clear link between these genotype variants and phenotypical translations is still absent. For instance, a different distribution of the *SOD2* p.Ile58Thr SNP in fertile vs. infertile men was not detected by a previous study [135]. Finally, the *SOD3* c.G362A SNP (rs2536512) was recently evaluated in 111 infertile men compared to 104 fertile controls [140], not detecting an association between genotypes and infertility, although a significant reduction in SOD activity was found in samples from the infertile group [140]. Thus, these three SNPs are promising candidates for explaining the role of SOD2 and SOD3 enzymes in male fertility, although further studies are needed to clearly address this issue.

Considering other antioxidant systems involved in ROS balance, other SNPs were studied, such as *CAT* c.C262T (rs1001179), *GXP1* p.Pro200Leu (rs1050450), and two human GST isozymes presenting a null allele as a result of a deletion (*GSTT* and *GSTM*) [135]. However, the current literature is scant to establish the role of these SNPs in male infertility. The *CAT* gene is located on chromosome 11p3, encoding for an enzyme involved in the detoxification of H₂O₂ to H₂O [141]. Only one study suggested that the allelic variant *CAT* C-262T CT identified a favorable fertility phenotype, while the CC genotype was associated with a doubled risk of male infertility [135]. Accordingly, higher H₂O₂ concentration and a reduced *CAT* activity were detected in the seminal plasma of infertile than fertile men [135]. However, this result needs to be further confirmed by independent clinical studies evaluating the contribution of other SNPs in the pathogenesis of male infertility.

Overall, these studies, although not entirely conclusive, suggest that the allelic variant of genes implicated in oxidative stress and expressed at the endothelial level could impact parameters of male fertility.

4. Endothelial Function and Male Fertility: Inflammation Markers

Soluble adhesion molecules and pro-inflammatory cytokines are markers of systemic inflammatory status [142,143] and are involved in several pathophysiological processes [142–144]. Indeed, a link between the immune system and fertility parameter was proposed decades ago [145] and was recently confirmed by experimental data [146]. Endothelial cells are primary targets for several pro-inflammatory cytokines and mediate the interaction with circulating immune cells, regulating tissue infiltration. Although these mechanisms may be activated at the systemic level, a focus on their implications in the male reproductive system is provided in the following sections.

4.1. Cytokines and Semen Quality

The presence of leucocytospermia reduces semen quality and increases the sperm DNA fragmentation index [147,148]. This relationship is not completely understood, since a beneficial effect of moderate leucocytospermia in obtaining pregnancy has been suggested [149,150]. In any case, while the presence of leucocytospermia is generally considered a poor sensitive factor [151], cytokine seminal levels are considered a more accurate parameter to discriminate inflammatory and non-inflammatory semen [147]. Interleukin (IL)-8 is a cytokine produced by several cells, including blood and endothelial cells [152]. IL-8 acts synergistically with IL-1 to chemo-attract leucocytes in the inflammation site and to promote phagocytosis [153]. IL-8 concentration within the seminal plasma has been widely investigated as a possible marker of spermatogenesis [152,154]. However, its application as a predictive factor of the fertility status in clinical practice is still limited, since seminal IL-8 appeared directly related to leucocytospermia, while being inversely related to ejaculatory volume only in patients with male accessory gland infections [152]. In addition, conflicting evidence is available about the link between seminal IL-8 and semen parameters [155–159], which likely resulted as a possible indicator of alterations in the post-testicular male genital tract, rather than a marker of spermatogenesis.

Increased levels of interferon (IFN)- γ and IL-7 were found in the seminal plasma of men with chronic prostatitis and chronic pelvic pain syndrome [154]. In these patients, both IFN- γ and IL-7 could be related to reduced semen quality [154]. Moreover, other pro-inflammatory cytokines, such as IL-6, IL-1 β , and tumor necrosis factor-alpha (TNF- α), were largely investigated both in blood and in semen samples of infertile males [160], providing contradictory results. Indeed, several studies described the connection between seminal impairment and increased levels of these molecules [160–167], while others did not [160,163,168,169]. Taken together, these data suggest that the activation of the inflammatory/immune response documented by the increase of pro-inflammatory mediators in the seminal plasma are mainly the mirror of an inflammatory status of the genital male tract, rather than an expression of a testicular inflammatory state. The link between inflammation and the measured male genital product, i.e., seminal fluid, remains still far from being elucidated.

4.2. Adhesion Molecules and Sperm Maturation

In the testicular seminiferous tubules, the physiological micro-environment required for sperm development is preserved through dynamic reworking of cell junctions, constituting the blood–testis barrier [170]. Intercellular junctions are mainly formed by cadherins/catenins cell adhesion protein complexes and integrins, which connect proteins of the extracellular matrix to the intracellular cytoskeleton [171]. The main role of this barrier is the control of those molecules that could reach the intraluminal site and, in turn, the spermatogenic cells, limiting interferences in sperm maturation [170,172]. Indeed, there is evidence supporting that several chemotherapy agents induce spermatogonia differentia-

tion arrest and the blockade of spermatocyte meiosis. These events lead to azoospermia, damaging the blood–testis barrier and inhibiting cell adhesion molecules (CAMs) and cadherin expression [170]. CAMs are integral membrane proteins typically constituted by three domains, i.e., extracellular, transmembrane, and intra-cytoplasmic, involved in inducing proper modifications of cell adhesion in response to physiological functional needs [171]. In particular, the CAMs extracellular domain has a Ig-like region able to mediate homophilic or heterophilic dimerization with integrins and cadherins [173]. The crosstalk between these cell adhesion systems supports the junction renovation in human testis [171]. Several members of CAMs family acting within the testis have been identified and studied in knock-out animal models, providing specific fertility phenotypes [171]. Among these CAMs, ICAM-1, its soluble form (sICAM-1), and VCAM-1 are the most investigated molecules in the context of male fertility.

ICAM-1 is expressed in Sertoli cells and germ cells in a stage-specific manner modulating blood–testis barrier properties. In particular, the over-expression of the integral membrane form reinforces the barrier function, while the soluble form (sICAM-1) down-regulates several blood–testis barrier protein components [171,174]. The balance between ICAM-1 and sICAM-1 opposite action impacts the permeability of the blood–testis barrier [171]. Moreover, ICAM-1 recruits VCAM-1 at the endothelial cell level, mediating the cell adhesion signaling [175]. Indeed, VCAM-1 facilitates the leukocyte adhesion to the vascular endothelium [176,177], while its active soluble form (sVCAM-1) mediates immune and inflammatory reactions at the peripheral level [178]. The complexity of this scenario is increased by the up-regulatory role exerted by several cytokines, such as TNF- α , IL-6 and IFN- γ , on ICAM-1 and VCAM-1 [179–181].

Another actor playing a potential role in the physiological sperm migration into seminiferous tubules seems to be ICAM-2 [171]. This molecule is constitutively expressed in epithelial and endothelial cells at the testicular level, where it mediates the conformational “stabilization” of actin molecules, inducing spermiation [180].

Vascular endothelial cadherin (VEC) is a single-pass transmembrane protein playing a crucial role in stabilizing the intracellular cytoskeleton and mediating cell–cell contacts [182]. Alongside its adhesive function, VEC is involved in maintaining the homeostasis of the seminiferous epithelial cycle, showing a stage-specific expression pattern at the testicular level [183]. More precisely, VEC may be involved in the acquisition of spermatid polarity within the seminiferous epithelium [182]. In male mice, the loss of VEC expression occurring together with the progression of sperm maturation stages leads to impairment of germ-Sertoli cell contacts, intra-lumen transition of immature spermatids and, subsequently, to low sperm count in the ejaculate [183].

Taken together, our knowledge of adhesion molecule functioning in testicular tissues was provided mainly by animal models. However, its role in human spermatogenesis needs further clarifications. On the other hand, it is undeniable that this issue is hardly surmountable, considering that in vivo studies investigating the role of these molecules in human tissues are hardly doable.

4.3. Role of Endothelin-1 and Sperm Function

In the male genital tract, endothelin-1 (ET-1) is detectable at testicular, epididymal and prostatic levels both in human and in rat males [184,185]. Within the testis, ET receptors were mainly found in Leydig cells, in which they promote testosterone secretion [186]. In Sertoli cells, ET receptors are involved in micro-tubule contractility [187]. However, the most known ET-1 role was observed in the epididymis, where it induces smooth muscle contractions required for supporting the transit of immotile sperms through the excretory ducts [188]. This concept is reflected by the positive correlation between ET-1 levels and seminal fluid volume [189]. Although these data demonstrated that ET-1 is involved in the regulation of human fertility, no evidence is currently available suggesting a physiological direct relationship between ET-1 and parameters indicative of sperm functions [190]. A genetically modified mouse model with ET-1 overexpression had reduced testicular blood

flow, but neither increased inflammatory status, nor impaired cell proliferation [184]. Hypothetically, ET-1 dysfunction could impair sperm progression through the genital tract, negatively impacting the viability of gametes in the ejaculate. To date, properly designed clinical studies are needed to clarify this mechanism in humans.

4.4. Vascular Endothelial Growth Factor-A and Testicular Function

Vascular endothelial growth factor-A-encoding (*VEGF-A*) gene transcription leads to multiple alternative splicing isoforms resulting in either anti- or pro-angiogenic properties GF [191,192]. In humans, *VEGF-A* is physiologically expressed in testis, seminal vesicles, prostate, measurable in the seminal fluid, and involved in the control of male reproductive functions [193]. *VEGF-A* stimulates angiogenetic processes and modulates the testicular blood vessels permeability via balancing of pro-angiogenic and anti-angiogenic isoforms [191,194]. In addition, this factor is proposed to be a regulator of the spermatogonia stem cell pool within the testis, balancing at the same time its self-renewal throughout premature differentiation and cell death [191]. Accordingly, loss of *VEGF-A* isoforms in mouse models determines the reduction of sperm number, due to the impairment of those mechanisms required for long-term maintenance of undifferentiated spermatogonia [195]. In vitro experiments suggested that the action of *VEGF-A* may be beneficial for some sperm motility parameters, in a concentration-dependent manner. Moreover, mouse models revealed that *VEGF-A* induces germ cells proliferation, promotes testicular regeneration by enhancement of the vascularization [196] and supports Leydig cell steroidogenesis [197]. Since both steroidogenesis and gametogenesis require an intact testicular microvascular compartment [193,197], it is not surprising that *VEGF-A* paracrine effects are required to maintain physiological testicular functions [197].

5. Endothelial Function and Female Fertility

While the link between endothelial dysfunction and female sex hormones is described in the literature as reported above, the impact of endothelial dysfunction on female fertility in humans is unclear. Mouse models suggested that estradiol exerts a number of vasoactive properties, beyond its physiological action on the reproductive axis [198]. Therefore, it could be hypothesized that estrogen may be involved in the regulation of endothelial functions supporting female reproduction.

5.1. Endothelial Dysfunction Markers in Premature Ovarian Insufficiency

Premature ovarian insufficiency (POI) is defined as ovarian function failure occurring before 40 years. Clinically, it is characterized by amenorrhea, sex steroid deficiency, and high gonadotropin serum levels [199]. The rapid decline in circulating estrogen levels exposes women to a number of complications, such as the increased risk for cardiovascular morbidity and mortality [200,201]. Endothelial functions could be related to the pathogenesis of POI, as suggested by the lower FMD, IMT, and endothelial progenitor cells (EPCs) in women affected by the disease compared to healthy controls [202,203]. The FMD improvement after oral, six months' estrogen/progestogen cyclic treatment supports the hormone-dependent etiology of endothelial dysfunction in POI [202]. These results were confirmed by a clinical study demonstrating that POI women treated with estrogen replacement therapy had similar endothelial function compared to controls [204].

5.2. Endothelial Dysfunction Markers in Polycystic Ovary Syndrome

Polycystic ovary syndrome (PCOS) is the most frequent endocrine disorder in women worldwide [205]. The disease is clinically characterized by a combination of hyperandrogenism, anovulation, and metabolic disorders [206]. The latter could display both endothelial dysfunctions [207] and early signs of atherosclerosis, such as increased IMT and coronary calcium deposit [208]. Endothelial dysfunction may be related to tissue resistance to the insulin-induced vasodilation, as demonstrated in vivo [207]. Indeed, the role of insulin in modulating endothelial function is confirmed by the beneficial effect of

metformin treatment. This drug increases the peripheral insulin sensitivity, improving the reactive hyperemia-peripheral arterial tonometry in subjects with abnormal endothelial functions [209]. The same effect is demonstrated in PCOS rat models, in which the increase of insulin sensitivity by treatment with metformin and the anti-androgen flutamide improves endothelial functions [210]. A systematic review confirmed the relationship between endothelial dysfunction and insulin resistance [211]. However, since not all women with both insulin resistance and PCOS experienced endothelial dysfunction, other pathogenic contributing mechanisms should be considered.

Alongside insulin resistance, the estrogens/androgens imbalance has been proposed as a co-causative factor of endothelial dysfunction in PCOS [207]. Serum androgen levels may be elevated in women with PCOS [212]. Androgens could lead to endothelial dysfunction in both lean and obese women with PCOS, as recently evaluated by the elevation of ET-B (an ET-1 receptor) in the same cohort of patients [213]. Moreover, PCOS women have reduced EPCs number and function, likely due to increased ROS levels and impaired insulin signaling [214]. Finally, other biomarkers of endothelial dysfunction are increased in women with PCOS, such as visfatin, VEGF, matrix metalloproteinase 9 (MMP9), high-mobility group box 1, pentraxin 3, and soluble lectin-like oxidized low-density lipoprotein receptor-1 [215].

5.3. Endothelial Dysfunction Markers in Endometriosis: The Role of Cytokines

Endometriosis is a frequent benign gynecological disorder characterized by the presence of endometrial tissue outside the uterine cavity, caused by pathogenic mechanisms not fully understood yet. Endometrial cell survival outside the uterus is potentially supported by the formation of new vessels. Thus, angiogenic growth factors and circulating EPCs could serve as biomarkers for the diagnosis and classification of endometriosis [216]. Indeed, women affected by endometriosis have higher peritoneal levels of several angiogenic cytokines, such as VEGF, IL-1, IL-6 and IL-8 [217]. This cytokine-rich peritoneal fluid may increase the mitotic activity of endothelial cells, indicating the role of these molecules in promoting angiogenesis, implantation, and growth of endometrial transplants [218].

Interestingly, women with endometriosis have high oxidative stress and systemic inflammation, together with a pro-atherogenic lipid profile, which could increase the risk of developing atherosclerosis. Accordingly, some studies demonstrated the presence of subclinical atherosclerosis in women affected by endometriosis, with regression after surgical removal of ectopic masses [219]. When endothelial function was tested, women with endometriosis showed significantly lower FMD compared to controls, although no differences in IMT were found [220]. Thus, endometriosis seems to be related only to a subclinical endothelial dysfunction and atherosclerosis.

5.4. Tumor Necrosis Factor- α in Female Fertility

TNF- α has been widely studied in relation to female fertility, both in animals and in humans. In TNF- α knockout mouse models, increased proliferation of granulosa cells and decreased oocytes apoptosis were described compared to wild type, confirming that the lack of TNF- α activity increases fertility [221]. In humans, TNF- α has been associated with inflammatory mechanisms related to implantation, placentation, and pregnancy outcome. Although the evidence is still limited, the reduction of TNF- α could be the target of specific therapies in women with obstetric complications, such as recurrent pregnancy loss, early and severe pre-eclampsia, and recurrent implantation failure syndrome [222].

6. Endothelial Function and Assisted Reproductive Technology

In light of the increasing evidence describing the connection between endothelial health and reproductive function, several endothelial markers have been evaluated as potential predictors of assisted reproductive technology (ART) outcome. For instance, among these, VEGF-A was identified as a negative predictor of in vitro fertilization (IVF) success, measured in both female blood [223] and embryo culture medium [224]. Moreover,

several cytokines have been investigated in an ART setting. TNF- α detected in follicular fluid on the day of oocyte retrieval has been associated with reduced number of fertilized oocytes while serum TNF- α levels were negatively correlated with fertilization rate [225]. Other pro-inflammation cytokines, such as IFN- γ , IL-12, IL-15, IL-18, and MMPs, were demonstrated to be detrimental to reproductive success in women with repeated IVF-embryo transfer failure [226–228]. Accordingly, the ART outcome is potentially influenced by cytokine levels detected in the seminal plasma of the male counterpart, suggesting that ART negative outcomes are related to the increased inflammatory status in at least one of the two partners. Contrasting results were provided by studies not detecting different follicular fluid and blood serum concentrations of TNF- α , IL-1 β , IL-6, IL-8, IL-12, and NO between women becoming pregnant after ART and those who do not [229,230]. Therefore, agreed opinions on the link between pro-inflammatory cytokines and ART outcomes is not achieved so far. Hitherto, considering the overall evidence, neither serum nor intrafollicular concentrations of these molecules can be considered predictive of the ART outcome.

The involvement of *VEGF* gene SNPs in reproductive functions was previously evaluated [193]. Several *VEGF* SNPs were associated with recurrent spontaneous miscarriage, preeclampsia, and premature birth [231,232]. In addition, the frequency of C allele genotype in *VEGF-A* c.936C > T (rs7664413) and in c.405G > C (rs2010963) were higher in non-pregnant women after IVF than in those pregnant [224,233]. Similarly, the *TNF* c.-308A > G (rs1800629) A allele, associated to a reduced transcriptional activity [233], was linked to higher implantation and pregnancy rates in women undergoing ART [233,234]. Recently, other genes were investigated, trying to obtain an a priori prediction of ART outcomes [235]. As observed in others multifactorial conditions, a combination of SNPs belonging to different genes likely contributes in determining the success of the clinical treatment [233]. In the ART context, we are currently far from the identification of reliable markers.

7. Conclusions

Endothelial dysfunction is largely investigated both in clinic and research settings to predict cardiovascular risk. Several molecular markers of endothelial dysfunction are known nowadays and here we described their roles in human fertility [236,237]. However, a real future use of these molecules as reliable markers in the clinical work-up of infertile couples is still far to be achieved. Indeed, previous research attempting to find a correlation between fertility and infertility parameters and endothelial dysfunction detected possible candidates unsuitable or weakly exploitable in clinical practice. The polygenic nature of regulatory mechanisms underlying reproduction prevents to find unique, clear-cut markers of fertility and infertility, which is rather the result of several contributing factors. In conclusion, although a link between endothelial health and reproductive parameters was established, no clear and suitable markers of fertility and infertility are available so far.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Authors are grateful to the Italian Ministry of University and Research for supporting the Department of Biomedical, Metabolic, and Neural Sciences (University of Modena and Reggio Emilia, Italy) in the context of the Departments of Excellence Programme. The authors also thank “Progetti di Rilevante Interesse Nazionale” (PRIN) 2017.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Green, M.P.; Harvey, A.J.; Finger, B.J.; Tarulli, G.A. Endocrine disrupting chemicals: Impacts on human fertility and fecundity during the peri-conception period. *Environ. Res.* **2021**, *194*, 110694. [[CrossRef](#)]
2. Cahill, P.A.; Redmond, E.M. Vascular endothelium—Gatekeeper of vessel health. *Atherosclerosis* **2016**, *248*, 97–109. [[CrossRef](#)]
3. Libby, P.; Ridker, P.M.; Hansson, G.K. Progress and challenges in translating the biology of atherosclerosis. *Nat. Cell Biol.* **2011**, *473*, 317–325. [[CrossRef](#)]
4. Poredos, P.; Poredos, A.V.; Gregoric, I. Endothelial Dysfunction and Its Clinical Implications. *Angiology* **2021**, 3319720987752. [[CrossRef](#)]
5. Peters, S.A.; Colantonio, L.D.; Dai, Y.; Zhao, H.; Bittner, V.A.; Farkouh, M.E.; Dlugniewski, P.; Poudel, B.; Muntner, P.; Woodward, M. Trends in Recurrent Coronary Heart Disease After Myocardial Infarction Among US Women and Men Between 2008 and 2017. *Circulation* **2021**, *143*, 650–660. [[CrossRef](#)]
6. Sullivan, S.; Young, A.; Hammadah, M.; Lima, B.B.; Levantsevych, O.; Ko, Y.-A.; Pearce, B.D.; Shah, A.J.; Kim, J.H.; Moazzami, K.; et al. Sex differences in the inflammatory response to stress and risk of adverse cardiovascular outcomes among patients with coronary heart disease. *Brain Behav. Immun.* **2020**, *90*, 294–302. [[CrossRef](#)]
7. Benjamin, E.J.; Muntner, P.; Alonso, A.; Bittencourt, M.S.; Callaway, C.W.; Carson, A.P.; Chamberlain, A.M.; Chang, A.R.; Cheng, S.; Das, S.R.; et al. Heart disease and stroke statistics—2019 update: A report from the American heart association. *Circulation* **2019**, *139*, e56–e528. [[CrossRef](#)]
8. Colditz, G.A.; Willett, W.C.; Stampfer, M.J.; Rosner, B.; Speizer, F.E.; Hennekens, C.H. Menopause and the Risk of Coronary Heart Disease in Women. *N. Engl. J. Med.* **1987**, *316*, 1105–1110. [[CrossRef](#)] [[PubMed](#)]
9. Boese, A.C.; Kim, S.C.; Yin, K.-J.; Lee, J.-P.; Hamblin, M.H. Sex differences in vascular physiology and pathophysiology: Estrogen and androgen signaling in health and disease. *Am. J. Physiol. Circ. Physiol.* **2017**, *313*, H524–H545. [[CrossRef](#)] [[PubMed](#)]
10. Green, D.J.; Hopkins, N.D.; Jones, H.; Thijssen, D.H.J.; Eijssvogels, T.M.H.; Yeap, B.B. Sex differences in vascular endothelial function and health in humans: Impacts of exercise. *Exp. Physiol.* **2016**, *101*, 230–242. [[CrossRef](#)] [[PubMed](#)]
11. Stanhewicz, A.E.; Wenner, M.M.; Stachenfeld, N.S. Sex differences in endothelial function important to vascular health and overall cardiovascular disease risk across the lifespan. *Am. J. Physiol. Circ. Physiol.* **2018**, *315*, H1569–H1588. [[CrossRef](#)]
12. Sapp, R.M.; Landers-Ramos, R.Q.; Shill, D.D.; Springer, C.B.; Hagberg, J.M. Sex-specific alterations in blood-borne factors in physically inactive individuals are detrimental to endothelial cell functions. *J. Appl. Physiol.* **2020**, *129*, 664–674. [[CrossRef](#)] [[PubMed](#)]
13. Walsh, B.W.; Schiff, I.; Rosner, B.; Greenberg, L.; Ravnkar, V.; Sacks, F.M. Effects of Postmenopausal Estrogen Replacement on the Concentrations and Metabolism of Plasma Lipoproteins. *N. Engl. J. Med.* **1991**, *325*, 1196–1204. [[CrossRef](#)] [[PubMed](#)]
14. Sack, M.; Rader, D.J.; Cannon, R. Oestrogen and inhibition of oxidation of low-density lipoproteins in postmenopausal women. *Lancet* **1994**, *343*, 269–270. [[CrossRef](#)]
15. Iorga, A.; Cunningham, C.M.; Moazeni, S.; Ruffenach, G.; Umar, S.; Eghbali, M. The protective role of estrogen and estrogen receptors in cardiovascular disease and the controversial use of estrogen therapy. *Biol. Sex Differ.* **2017**, *8*, 1–16. [[CrossRef](#)] [[PubMed](#)]
16. Da Silva, J.S.; Montagnoli, T.L.; Rocha, B.S.; Tacco, M.L.C.A.; Marinho, S.C.P.; Zapata-Sudo, G. Estrogen Receptors: Therapeutic Perspectives for the Treatment of Cardiac Dysfunction after Myocardial Infarction. *Int. J. Mol. Sci.* **2021**, *22*, 525. [[CrossRef](#)] [[PubMed](#)]
17. Kannenkeril, D.; Bosch, A.; Kolwelter, J.; Jung, S.; Striepe, K.; Ott, C.; Delles, C.; Schmieder, R.E. Dependency of flow mediated vasodilatation from basal nitric oxide activity. *Clin. Physiol. Funct. Imaging* **2021**. [[CrossRef](#)]
18. Pérez-López, F.R.; Chedraui, P.; Gilbert, J.J.; Perez-Roncero, G. Cardiovascular risk in menopausal women and prevalent related co-morbid conditions: Facing the post-Women’s Health Initiative era. *Fertil. Steril.* **2009**, *92*, 1171–1186. [[CrossRef](#)] [[PubMed](#)]
19. Celermajer, D.S.; Sorensen, K.E.; Spiegelhalter, D.J.; Georgakopoulos, D.; Robinson, J.; Deanfield, J.E. Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. *J. Am. Coll. Cardiol.* **1994**, *24*, 471–476. [[CrossRef](#)]
20. Knowlton, A.A.; Lee, A.R. Estrogen and the cardiovascular system. *Pharmacol. Ther.* **2012**, *135*, 54–70. [[CrossRef](#)]
21. Chakrabarti, S.; Morton, J.S.; Davidge, S.T. Mechanisms of Estrogen Effects on the Endothelium: An Overview. *Can. J. Cardiol.* **2014**, *30*, 705–712. [[CrossRef](#)] [[PubMed](#)]
22. Lorigo, M.; Mariana, M.; Lemos, M.C.; Cairrao, E. Vascular mechanisms of testosterone: The non-genomic point of view. *J. Steroid Biochem. Mol. Biol.* **2020**, *196*, 105496. [[CrossRef](#)] [[PubMed](#)]
23. Yu, J.; Akishita, M.; Eto, M.; Ogawa, S.; Son, B.-K.; Kato, S.; Ouchi, Y.; Okabe, T. Androgen Receptor-Dependent Activation of Endothelial Nitric Oxide Synthase in Vascular Endothelial Cells: Role of Phosphatidylinositol 3-Kinase/Akt Pathway. *Endocrinology* **2010**, *151*, 1822–1828. [[CrossRef](#)]
24. Yu, J.; Akishita, M.; Eto, M.; Koizumi, H.; Hashimoto, R.; Ogawa, S.; Tanaka, K.; Ouchi, Y.; Okabe, T. Src kinase mediates androgen receptor-dependent non-genomic activation of signaling cascade leading to endothelial nitric oxide synthase. *Biochem. Biophys. Res. Commun.* **2012**, *424*, 538–543. [[CrossRef](#)] [[PubMed](#)]
25. Thomas, P. Membrane Androgen Receptors Unrelated to Nuclear Steroid Receptors. *Endocrinology* **2019**, *160*, 772–781. [[CrossRef](#)]
26. Thomas, P.; Converse, A.; Berg, H.A. ZIP9, a novel membrane androgen receptor and zinc transporter protein. *Gen. Comp. Endocrinol.* **2018**, *257*, 130–136. [[CrossRef](#)]

27. Yue, P.; Chatterjee, K.; Beale, C.; Poole-Wilson, P.A.; Collins, P. Testosterone Relaxes Rabbit Coronary Arteries and Aorta. *Circulation* **1995**, *91*, 1154–1160. [[CrossRef](#)] [[PubMed](#)]
28. Tep-Areenan, P.; Kendall, D.A.; Randall, M.D. Testosterone-induced vasorelaxation in the rat mesenteric arterial bed is mediated predominantly via potassium channels. *Br. J. Pharmacol.* **2002**, *135*, 735–740. [[CrossRef](#)] [[PubMed](#)]
29. Chou, T.M.; Sudhir, K.; Hutchison, S.J.; Ko, E.; Amidon, T.M.; Collins, P.; Chatterjee, K. Testosterone Induces Dilation of Canine Coronary Conductance and Resistance Arteries In Vivo. *Circulation* **1996**, *94*, 2614–2619. [[CrossRef](#)]
30. Yildiz, O.; Seyrek, M.; Gul, H.; Un, I.; Yildirim, V.; Ozal, E.; Uzun, M.; Bolu, E. Testosterone Relaxes Human Internal Mammary Artery In Vitro. *J. Cardiovasc. Pharmacol.* **2005**, *45*, 580–585. [[CrossRef](#)] [[PubMed](#)]
31. Tep-Areenan, P.; Kendall, D.A.; Randall, M.D. Mechanisms of vasorelaxation to testosterone in the rat aorta. *Eur. J. Pharmacol.* **2003**, *465*, 125–132. [[CrossRef](#)]
32. Montañó, L.M.; Calixto, E.; Figueroa, A.; Flores-Soto, E.; Carbajal, V.; Perusquía, M. Relaxation of Androgens on Rat Thoracic Aorta: Testosterone Concentration Dependent Agonist/Antagonist I-Type Ca²⁺ Channel Activity, and 5 β -Dihydrotestosterone Restricted to I-Type Ca²⁺ Channel Blockade. *Endocrinology* **2008**, *149*, 2517–2526. [[CrossRef](#)]
33. Isidoro, L.; Ferrer, M.; Perusquía, M. Vasoactive androgens: Vasorelaxing effects and their potential regulation of blood pressure. *Endocr. Res.* **2018**, *43*, 166–175. [[CrossRef](#)]
34. Rowell, K.O.; Hall, J.; Pugh, P.J.; Jones, T.H.; Channer, K.S.; Jones, R.D. Testosterone acts as an efficacious vasodilator in isolated human pulmonary arteries and veins: Evidence for a biphasic effect at physiological and supra-physiological concentrations. *J. Endocrinol. Investig.* **2009**, *32*, 718–723. [[CrossRef](#)] [[PubMed](#)]
35. Bartel, D.P. Metazoan MicroRNAs. *Cell* **2018**, *173*, 20–51. [[CrossRef](#)] [[PubMed](#)]
36. Bammert, T.D.; Hijmans, J.G.; Kavlich, P.J.; Lincenberg, G.M.; Reiakvam, W.R.; Fay, R.T.; Greiner, J.J.; Stauffer, B.L.; DeSouza, C.A. Influence of sex on circulating endothelial microparticle number and micro RNA expression in middle-aged adults. *Exp. Physiol.* **2017**, *102*, 894–900. [[CrossRef](#)] [[PubMed](#)]
37. Bye, A.; Røsjø, H.; Aspenes, S.T.; Condorelli, G.; Omland, T.; Wisløff, U. Circulating MicroRNAs and Aerobic Fitness – The HUNT-Study. *PLOS ONE* **2013**, *8*, e57496. [[CrossRef](#)]
38. Regitz-Zagrosek, V.; Kararigas, G. Mechanistic Pathways of Sex Differences in Cardiovascular Disease. *Physiol. Rev.* **2017**, *97*, 1–37. [[CrossRef](#)] [[PubMed](#)]
39. Lau, E.S.; Paniagua, S.M.; Guseh, J.S.; Bhambhani, V.; Zanni, M.V.; Courchesne, P.; Lyass, A.; Larson, M.G.; Levy, D.; Ho, J.E. Sex Differences in Circulating Biomarkers of Cardiovascular Disease. *J. Am. Coll. Cardiol.* **2019**, *74*, 1543–1553. [[CrossRef](#)]
40. Ludmer, P.L.; Selwyn, A.P.; Shook, T.L.; Wayne, R.R.; Mudge, G.H.; Alexander, R.W.; Ganz, P. Paradoxical Vasoconstriction Induced by Acetylcholine in Atherosclerotic Coronary Arteries. *N. Engl. J. Med.* **1986**, *315*, 1046–1051. [[CrossRef](#)]
41. Chowienczyk, P.J.; Cockcroft, J.R.; Ritter, J.M. Blood Flow Responses to Intra-Arterial Acetylcholine in Man: Effects of Basal Flow and Conduit Vessel Length. *Clin. Sci.* **1994**, *87*, 45–51. [[CrossRef](#)]
42. Corretti, M.C.; Anderson, T.J.; Benjamin, E.J.; Celermajer, D.; Charbonneau, F.; Creager, M.A.; Deanfield, J.; Drexler, H.; Gerhard-Herman, M.; Herrington, D.; et al. Guidelines for the ultrasound assessment of endothelial-dependent flow-mediated vasodilation of the brachial artery. *J. Am. Coll. Cardiol.* **2002**, *39*, 257–265. [[CrossRef](#)]
43. Deanfield, J.E.; Halcox, J.P.; Rabelink, T.J. Endothelial Function and Dysfunction. *Circulation* **2007**, *115*, 1285–1295. [[CrossRef](#)]
44. Anderson, T.J.; Uehata, A.; Gerhard, M.D.; Meredith, I.T.; Knab, S.; Delagrangé, D.; Lieberman, E.H.; Ganz, P.; Creager, M.A.; Yeung, A.C.; et al. Close relation of endothelial function in the human coronary and peripheral circulations. *J. Am. Coll. Cardiol.* **1995**, *26*, 1235–1241. [[CrossRef](#)]
45. Verma, S.; Buchanan, M.R.; Anderson, T.J. Endothelial Function Testing as a Biomarker of Vascular Disease. *Circulation* **2003**, *108*, 2054–2059. [[CrossRef](#)]
46. Brevetti, G.; Silvestro, A.; Schiano, V.; Chiariello, M. Endothelial Dysfunction and Cardiovascular Risk Prediction in Peripheral Arterial Disease. *Circulation* **2003**, *108*, 2093–2098. [[CrossRef](#)] [[PubMed](#)]
47. Xu, Y.; Arora, R.C.; Hiebert, B.M.; Lerner, B.; Sz wajczer, A.; McDonald, K.; Rigatto, C.; Komenda, P.; Sood, M.M.; Tangri, N. Non-invasive endothelial function testing and the risk of adverse outcomes: A systematic review and meta-analysis. *Eur. Heart J. - Cardiovasc. Imaging* **2014**, *15*, 736–746. [[CrossRef](#)]
48. Matsuzawa, Y.; Kwon, T.; Lennon, R.J.; Lerman, L.O.; Lerman, A. Prognostic Value of Flow-Mediated Vasodilation in Brachial Artery and Fingertip Artery for Cardiovascular Events: A Systematic Review and Meta-Analysis. *J. Am. Heart Assoc.* **2015**, *4*, e002270. [[CrossRef](#)] [[PubMed](#)]
49. Neunteufl, T.; Heher, S.; Katzenschlager, R.; Wöfl, G.; Kostner, K.; Maurer, G.; Weidinger, F. Late prognostic value of flow-mediated dilation in the brachial artery of patients with chest pain. *Am. J. Cardiol.* **2000**, *86*, 207–210. [[CrossRef](#)]
50. Gokce, N.; Keaney, J.F.; Hunter, L.M.; Watkins, M.T.; Menzoian, J.O.; Vita, J.A. Risk Stratification for Postoperative Cardiovascular Events via Noninvasive Assessment of Endothelial Function. *Circulation* **2002**, *105*, 1567–1572. [[CrossRef](#)]
51. Gokce, N.; Keaney, J.F.; Hunter, L.M.; Watkins, M.T.; Nedeljkovic, Z.S.; Menzoian, J.O.; Vita, J.A. Predictive value of noninvasively determined endothelial dysfunction for long-term cardiovascular events in patients with peripheral vascular disease. *J. Am. Coll. Cardiol.* **2003**, *41*, 1769–1775. [[CrossRef](#)]
52. Patti, G.; Pasceri, V.; Melfi, R.; Goffredo, C.; Chello, M.; D’Ambrosio, A.; Montesanti, R.; Di Sciascio, G. Impaired Flow-Mediated Dilation and Risk of Restenosis in Patients Undergoing Coronary Stent Implantation. *Circulation* **2005**, *111*, 70–75. [[CrossRef](#)]

53. Piollet, M.; Sturza, A.; Chadet, S.; Gabillard-Lefort, C.; Benoist, L.; Muntean, D.-M.; Aburel, O.-M.; Angoulvant, D.; Ivanes, F. P2Y11 Agonism Prevents Hypoxia/Reoxygenation- and Angiotensin II-Induced Vascular Dysfunction and Intimal Hyperplasia Development. *Int. J. Mol. Sci.* **2021**, *22*, 855. [[CrossRef](#)] [[PubMed](#)]
54. Naqvi, T.Z.; Lee, M.-S. Carotid Intima-Media Thickness and Plaque in Cardiovascular Risk Assessment. *JACC Cardiovasc. Imaging* **2014**, *7*, 1025–1038. [[CrossRef](#)]
55. Cobble, M.; Bale, B. Carotid Intima-Media Thickness: Knowledge and Application to Everyday Practice. *Postgrad. Med.* **2010**, *122*, 10–18. [[CrossRef](#)] [[PubMed](#)]
56. Polak, J.F.; O'Leary, D.H. Carotid Intima-Media Thickness as Surrogate for and Predictor of CVD. *Glob. Heart* **2016**, *11*, 295–312.e3. [[CrossRef](#)]
57. Osondu, C.U.; Vo, B.; Oni, E.T.; Blaha, M.J.; Veledar, E.; Feldman, T.; Agatston, A.S.; Nasir, K.; Aneni, E.C. The relationship of erectile dysfunction and subclinical cardiovascular disease: A systematic review and meta-analysis. *Vasc. Med.* **2018**, *23*, 9–20. [[CrossRef](#)] [[PubMed](#)]
58. Salvio, G.; Ciarloni, A.; Cutini, M.; Balercia, G. Hyperhomocysteinemia: Focus on Endothelial Damage as a Cause of Erectile Dysfunction. *Int. J. Mol. Sci.* **2021**, *22*, 418. [[CrossRef](#)]
59. Pelit, E.S.; Dokumaci, D.Ş.; Kati, B.; Yağmur, I.; Arslan, E.; Tunçtekin, A.; Kırteke, A.; Çiftçi, H.; Yeni, E. Carotid artery intima-media thickness can predict the response of patients with erectile dysfunction to phosphodiesterase 5 inhibitors. *Int. J. Impot. Res.* **2019**, *31*, 139–144. [[CrossRef](#)] [[PubMed](#)]
60. Santi, D.; Granata, A.R.; Pignatti, E.; Trenti, T.; Roli, L.; Bozic, R.; Zaza, S.; Pacchioni, C.; Rochira, V.; Carani, C.; et al. Effects of chronic administration of the phosphodiesterase inhibitor vardenafil on serum levels of adrenal and testicular steroids in men with type 2 diabetes mellitus. *Endocrine* **2016**, *56*, 426–437. [[CrossRef](#)] [[PubMed](#)]
61. Santi, D.; Giannetta, E.; Isidori, A.M.; Vitale, C.; Aversa, A.; Simoni, M. THERAPY OF ENDOCRINE DISEASE: Effects of chronic use of phosphodiesterase inhibitors on endothelial markers in type 2 diabetes mellitus: A meta-analysis. *Eur. J. Endocrinol.* **2015**, *172*, R103–R114. [[CrossRef](#)] [[PubMed](#)]
62. Santi, D.; Locaso, M.; Granata, A.R.; Trenti, T.; Roli, L.; Pacchioni, C.; Rochira, V.; Carani, C.; Simoni, M. Could chronic Vardenafil administration influence the cardiovascular risk in men with type 2 diabetes mellitus? *PLOS ONE* **2018**, *13*, e0199299. [[CrossRef](#)] [[PubMed](#)]
63. Skilton, M.R.; Celermajer, D.S.; Cosmi, E.; Crispi, F.; Gidding, S.S.; Raitakari, O.T.; Urbina, E.M. Natural History of Atherosclerosis and Abdominal Aortic Intima-Media Thickness: Rationale, Evidence, and Best Practice for Detection of Atherosclerosis in the Young. *J. Clin. Med.* **2019**, *8*, 1201. [[CrossRef](#)]
64. Buyukinan, M.; Atar, M.; Pirgon, Ö.; Kurku, H.; Erdem, S.S.; Deniz, I. Anti-Mullerian Hormone and Inhibin B Levels in Obese Boys; Relations with Cardiovascular Risk Factors. *Exp. Clin. Endocrinol. Diabetes* **2018**, *126*, 528–533. [[CrossRef](#)]
65. Maurice, R.L.; Vaujois, L.; Dahdah, N.; Nuyt, A.-M.; Bigras, J.-L. Comparing Carotid and Brachial Artery Stiffness: A First Step Toward Mechanical Mapping of the Arterial Tree. *Ultrasound Med. Biol.* **2015**, *41*, 1808–1813. [[CrossRef](#)] [[PubMed](#)]
66. Namba, T.; Masaki, N.; Takase, B.; Adachi, T. Arterial Stiffness Assessed by Cardio-Ankle Vascular Index. *Int. J. Mol. Sci.* **2019**, *20*, 3664. [[CrossRef](#)] [[PubMed](#)]
67. Saugel, B.; Kouz, K.; Scheeren, T.W.; Greiwe, G.; Hoppe, P.; Romagnoli, S.; De Backer, D. Cardiac output estimation using pulse wave analysis—physiology, algorithms, and technologies: A narrative review. *Br. J. Anaesth.* **2021**, *126*, 67–76. [[CrossRef](#)] [[PubMed](#)]
68. Laurent, S.; Cockcroft, J.; Van Bortel, L.; Boutouyrie, P.; Giannattasio, C.; Hayoz, D.; Pannier, B.; Vlachopoulos, C.; Wilkinson, I.; Struijker-Boudier, H.; et al. Expert consensus document on arterial stiffness: Methodological issues and clinical applications. *Eur. Heart J.* **2006**, *27*, 2588–2605. [[CrossRef](#)]
69. Mancia, G.; De Backer, G.; Dominiczak, A.; Cifkova, R.; Fagard, R.; Germano, G.; Grassi, G.; Heagerty, A.M.; Kjeldsen, S.E.; Laurent, S.; et al. Guidelines for the Management of Arterial Hypertension. *J. Hypertens.* **2007**, *25*, 1105–1187. [[CrossRef](#)]
70. Sequí-Domínguez, I.; Caverro-Redondo, I.; Álvarez-Bueno, C.; Pozuelo-Carrascosa, D.P.; De Arenas-Arroyo, S.N.; Martínez-Vizcaíno, V. Accuracy of Pulse Wave Velocity Predicting Cardiovascular and All-Cause Mortality. A Systematic Review and Meta-Analysis. *J. Clin. Med.* **2020**, *9*, 2080. [[CrossRef](#)]
71. Englund, E.K.; Langham, M.C. Quantitative and Dynamic MRI Measures of Peripheral Vascular Function. *Front. Physiol.* **2020**, *11*, 120. [[CrossRef](#)]
72. Dorner, G.T.; Garhofer, G.; Kiss, B.; Polska, E.; Polak, K.; Riva, C.E.; Schmetterer, L. Nitric oxide regulates retinal vascular tone in humans. *Am. J. Physiol. Circ. Physiol.* **2003**, *285*, H631–H636. [[CrossRef](#)]
73. Theuerle, J.D.; Al-Fiadh, A.H.; Islam, F.M.A.; Patel, S.K.; Burrell, L.M.; Wong, T.Y.; Farouque, O. Impaired retinal microvascular function predicts long-term adverse events in patients with cardiovascular disease. *Cardiovasc. Res.* **2020**. [[CrossRef](#)]
74. Ikee, R.; Kobayashi, S.; Hemmi, N.; Imakiire, T.; Kikuchi, Y.; Moriya, H.; Suzuki, S.; Miura, S. Correlation Between the Resistive Index by Doppler Ultrasound and Kidney Function and Histology. *Am. J. Kidney Dis.* **2005**, *46*, 603–609. [[CrossRef](#)]
75. Bruno, R.M.; Salvati, A.; Barzacchi, M.; Raimo, K.; Taddei, S.; Ghiadoni, L.; Solini, A. Predictive value of dynamic renal resistive index (drin) for renal outcome in type 2 diabetes and essential hypertension: A prospective study. *Cardiovasc. Diabetol.* **2015**, *14*, 1–9. [[CrossRef](#)]
76. Chen, M.; Masaki, T.; Sawamura, T. LOX-1, the receptor for oxidized low-density lipoprotein identified from endothelial cells: Implications in endothelial dysfunction and atherosclerosis. *Pharmacol. Ther.* **2002**, *95*, 89–100. [[CrossRef](#)]

77. Locatelli, L.; Fedele, G.; Castiglioni, S.; Maier, J.A. Magnesium Deficiency Induces Lipid Accumulation in Vascular Endothelial Cells via Oxidative Stress—The Potential Contribution of EDF-1 and PPAR γ . *Int. J. Mol. Sci.* **2021**, *22*, 1050. [[CrossRef](#)]
78. Wierzbicki, A.S.; Chowienczyk, P.J.; Cockcroft, J.R.; Brett, S.E.; Watts, G.F.; Jenkins, B.S.; Ritter, J.M. Cardiovascular risk factors and endothelial dysfunction. *Clin. Sci.* **2004**, *107*, 609–615. [[CrossRef](#)]
79. Nakamura, T.; Hirano, M.; Kitta, Y.; Fujioka, D.; Saito, Y.; Kawabata, K.-I.; Obata, J.-E.; Watanabe, Y.; Watanabe, K.; Kugiyama, K. A comparison of the efficacy of combined ezetimibe and statin therapy with doubling of statin dose in patients with remnant lipoproteinemia on previous statin therapy. *J. Cardiol.* **2012**, *60*, 12–17. [[CrossRef](#)]
80. Lupattelli, G.; Marchesi, S.; Roscini, A.R.; Siepi, D.; Gemelli, F.; Pirro, M.; Sinzinger, H.; Schillaci, G.; Mannarino, E. Direct association between high-density lipoprotein cholesterol and endothelial function in hyperlipemia. *Am. J. Cardiol.* **2002**, *90*, 648–650. [[CrossRef](#)]
81. Kuvin, J.T.; Patel, A.; Sidhu, M.; Rand, W.M.; Sliney, K.A.; Pandian, N.G.; Karas, R.H. Relation between high-Density lipoprotein cholesterol and peripheral vasomotor function. *Am. J. Cardiol.* **2003**, *92*, 275–279. [[CrossRef](#)]
82. Tran-Dinh, A.; Diallo, D.; Delbosc, S.; Varela-Perez, L.M.; Dang, Q.B.; Lapergue, B.; Burillo, E.; Michel, J.B.; Levoye, A.; Martin-Ventura, J.L.; et al. HDL and endothelial protection. *Br. J. Pharmacol.* **2013**, *169*, 493–511. [[CrossRef](#)]
83. Choi, H.Y.; Rahmani, M.; Wong, B.W.; Allahverdian, S.; McManus, B.M.; Pickering, J.G.; Chan, T.; Francis, G.A. ATP-Binding Cassette Transporter A1 Expression and Apolipoprotein A-I Binding Are Impaired in Intima-Type Arterial Smooth Muscle Cells. *Circulation* **2009**, *119*, 3223–3231. [[CrossRef](#)]
84. Fielding, P.E.; Nagao, K.; Hakamata, H.; Chimini, G.; Fielding, C.J. A two-step mechanism for free cholesterol and phospholipid efflux from human vascular cells to apolipoprotein A-1. *Biochemistry* **2000**, *39*, 14113–14120. [[CrossRef](#)] [[PubMed](#)]
85. Yuhanna, I.S.; Zhu, Y.; Cox, B.E.; Hahner, L.D.; Osborne-Lawrence, S.; Lu, P.; Marcel, Y.L.; Anderson, R.G.; Mendelsohn, M.E.; Hobbs, H.H.; et al. High-density lipoprotein binding to scavenger receptor-BI activates endothelial nitric oxide synthase. *Nat. Med.* **2001**, *7*, 853–857. [[CrossRef](#)] [[PubMed](#)]
86. Nofer, J.-R.; Van Der Giet, M.; Tolle, M.; Wolinska, I.; Lipinski, K.V.W.; Baba, H.A.; Tietge, U.J.; Gödecke, A.; Ishii, I.; Kleuser, B.; et al. HDL induces NO-dependent vasorelaxation via the lysophospholipid receptor S1P3. *J. Clin. Investig.* **2004**, *113*, 569–581. [[CrossRef](#)] [[PubMed](#)]
87. Potì, F.; Ceglarek, U.; Burkhardt, R.; Simoni, M.; Nofer, J.-R. SKI-II – a sphingosine kinase 1 inhibitor – exacerbates atherosclerosis in low-density lipoprotein receptor-deficient (LDL-R $-/-$) mice on high cholesterol diet. *Atherosclerosis* **2015**, *240*, 212–215. [[CrossRef](#)]
88. Potì, F.; Gualtieri, F.; Sacchi, S.; Weißen-Plenz, G.; Varga, G.; Brodde, M.; Weber, C.; Simoni, M.; Nofer, J.-R. KRP-203, Sphingosine 1-Phosphate Receptor Type 1 Agonist, Ameliorates Atherosclerosis in LDL-R $-/-$ Mice. *Arter. Thromb. Vasc. Biol.* **2013**, *33*, 1505–1512. [[CrossRef](#)]
89. Potì, F.; Simoni, M.; Nofer, J.-R. Atheroprotective role of high-density lipoprotein (HDL)-associated sphingosine-1-phosphate (S1P). *Cardiovasc. Res.* **2014**, *103*, 395–404. [[CrossRef](#)]
90. Brezis, M. C-reactive protein in the prediction of cardiovascular events. *N. Engl. J. Med.* **2003**, *348*, 1059.
91. Ridker, P.M.; Buring, J.E.; Cook, N.R.; Rifai, N. C-Reactive Protein, the Metabolic Syndrome, and Risk of Incident Cardiovascular Events. *Circulation* **2003**, *107*, 391–397. [[CrossRef](#)]
92. Fichtlscherer, S.; Rosenberger, G.; Walter, D.H.; Breuer, S.; Dimmeler, S.; Zeiher, A.M. Elevated C-reactive protein levels and impaired endothelial vasoreactivity in patients with coronary artery disease. *Circulation* **2000**, *102*, 1000–1006. [[CrossRef](#)] [[PubMed](#)]
93. Venugopal, S.K.; Devaraj, S.; Yuhanna, I.; Shaul, P.; Jialal, I. Demonstration That C-Reactive Protein Decreases eNOS Expression and Bioactivity in Human Aortic Endothelial Cells. *Circulation* **2002**, *106*, 1439–1441. [[CrossRef](#)]
94. Ikeda, U.; Takahashi, M.; Shimada, K. C-Reactive Protein Directly Inhibits Nitric Oxide Production by Cytokine-stimulated Vascular Smooth Muscle Cells. *J. Cardiovasc. Pharmacol.* **2003**, *42*, 607–611. [[CrossRef](#)] [[PubMed](#)]
95. Li, Y.; Zhong, X.; Cheng, G.; Zhao, C.; Zhang, L.; Hong, Y.; Wan, Q.; He, R.; Wang, Z. Hs-CRP and all-cause, cardiovascular, and cancer mortality risk: A meta-analysis. *Atherosclerosis* **2017**, *259*, 75–82. [[CrossRef](#)]
96. Homocysteine Studies Collaboration Homocysteine and Risk of Ischemic Heart Disease and Stroke. *JAMA* **2002**, *288*, 2015–2022. [[CrossRef](#)]
97. Shenoy, V.; Mehendale, V.; Prabhu, K.; Shetty, R.; Rao, P. Correlation of Serum Homocysteine Levels with the Severity of Coronary Artery Disease. *Indian J. Clin. Biochem.* **2013**, *29*, 339–344. [[CrossRef](#)]
98. Woo, K.; Chook, P.; Lolin, Y.; Cheung, A.; Chan, L.; Sun, Y.; Sanderson, J.; Metreweli, C.; Celermajer, D. Hyperhomocyst(e)inemia Is a Risk Factor for Arterial Endothelial Dysfunction in Humans. *Circulation* **1997**, *96*, 2542–2544. [[CrossRef](#)] [[PubMed](#)]
99. Wilson, J.B.; Welsch, M.; Allen, J.; Thomson, J.; Tulley, R.; Lefevre, M. The association of homocysteine and related factors to brachial artery diameter and flow-mediated dilation. *Metabolism* **2007**, *56*, 641–648. [[CrossRef](#)]
100. Basu, A.; Jenkins, A.J.; Stoner, J.A.; Thorpe, S.R.; Klein, R.L.; Lopes-Virella, M.F.; Garvey, W.T.; Lyons, T.J. Plasma total homocysteine and carotid intima-media thickness in type 1 diabetes: A prospective study. *Atherosclerosis* **2014**, *236*, 188–195. [[CrossRef](#)] [[PubMed](#)]
101. Zhang, S.; Bai, Y.-Y.; Luo, L.-M.; Xiao, W.-K.; Wu, H.-M.; Ye, P. Association between serum homocysteine and arterial stiffness in elderly: A community-based study. *J. Geriatr. Cardiol.* **2014**, *11*, 32–38. [[PubMed](#)]
102. Lai, W.K.C.; Kan, M.Y. Homocysteine-Induced Endothelial Dysfunction. *Ann. Nutr. Metab.* **2015**, *67*, 1–12. [[CrossRef](#)] [[PubMed](#)]

103. Joseph, J.; Joseph, L. Hyperhomocysteinemia and Cardiovascular Disease: New Mechanisms Beyond Atherosclerosis. *Metab. Syndr. Relat. Disord.* **2003**, *1*, 97–104. [[CrossRef](#)] [[PubMed](#)]
104. Lavi, S.; McConnell, J.P.; Rihal, C.S.; Prasad, A.; Mathew, V.; Lerman, L.O.; Lerman, A. Local Production of Lipoprotein-Associated Phospholipase A 2 and Lysophosphatidylcholine in the Coronary Circulation. *Circulation* **2007**, *115*, 2715–2721. [[CrossRef](#)] [[PubMed](#)]
105. Macphee, C.; Benson, G.M.; Shi, Y.; Zalewski, A. Lipoprotein-associated phospholipase A2: A novel marker of cardiovascular risk and potential therapeutic target. *Expert Opin. Investig. Drugs* **2005**, *14*, 671–679. [[CrossRef](#)] [[PubMed](#)]
106. Yang, E.H.; McConnell, J.P.; Lennon, R.J.; Barsness, G.W.; Pumper, G.; Hartman, S.J.; Rihal, C.S.; Lerman, L.O.; Lerman, A. Lipoprotein-Associated Phospholipase A2Is an Independent Marker for Coronary Endothelial Dysfunction in Humans. *Arter. Thromb. Vasc. Biol.* **2006**, *26*, 106–111. [[CrossRef](#)] [[PubMed](#)]
107. Garg, P.K.; McClelland, R.L.; Jenny, N.S.; Criqui, M.; Liu, K.; Polak, J.F.; Jorgensen, N.W.; Cushman, M. Association of lipoprotein-associated phospholipase A2 and endothelial function in the Multi-Ethnic Study of Atherosclerosis (MESA). *Vasc. Med.* **2011**, *16*, 247–252. [[CrossRef](#)] [[PubMed](#)]
108. Prasad, M.; Lennon, R.; Barsness, G.W.; Prasad, A.; Gulati, R.; Lerman, L.O.; Lerman, A. Chronic inhibition of lipoprotein-associated phospholipase A2 does not improve coronary endothelial function: A prospective, randomized-controlled trial. *Int. J. Cardiol.* **2018**, *253*, 7–13. [[CrossRef](#)]
109. Scioli, M.G.; Storti, G.; D’Amico, F.; Guzmán, R.R.; Centofanti, F.; Doldo, E.; Miranda, E.M.C.; Orlandi, A. Oxidative Stress and New Pathogenetic Mechanisms in Endothelial Dysfunction: Potential Diagnostic Biomarkers and Therapeutic Targets. *J. Clin. Med.* **2020**, *9*, 1995. [[CrossRef](#)] [[PubMed](#)]
110. Ronda, N.; Potì, F.; Palmisano, A.; Gatti, R.; Orlandini, G.; Maggiore, U.; Cabassi, A.; Regolisti, G.; Fiaccadori, E. Effects of the radiocontrast agent iodixanol on endothelial cell morphology and function. *Vasc. Pharmacol.* **2013**, *58*, 39–47. [[CrossRef](#)] [[PubMed](#)]
111. Colafella, K.M.M.; Bovée, D.M.; Danser, A.J. The renin-angiotensin-aldosterone system and its therapeutic targets. *Exp. Eye Res.* **2019**, *186*, 107680. [[CrossRef](#)]
112. Nouredine, F.Y.; Altara, R.; Fan, F.; Yabluchanskiy, A.; Booz, G.W.; Zouein, F.A. Impact of the Renin–Angiotensin System on the Endothelium in Vascular Dementia: Unresolved Issues and Future Perspectives. *Int. J. Mol. Sci.* **2020**, *21*, 4268. [[CrossRef](#)]
113. Charkoudian, N. Mechanisms and modifiers of reflex induced cutaneous vasodilation and vasoconstriction in humans. *J. Appl. Physiol.* **2010**, *109*, 1221–1228. [[CrossRef](#)] [[PubMed](#)]
114. Rosselli, M.; Keller, P.J.; Dubey, R.K. Role of nitric oxide in the biology, physiology and pathophysiology of reproduction. *Hum. Reprod. Update* **1998**, *4*, 3–24. [[CrossRef](#)] [[PubMed](#)]
115. Yang, Y.; Yu, T.; Lian, Y.-J.; Ma, R.; Yang, S.; Cho, J.Y. Nitric oxide synthase inhibitors: A review of patents from 2011 to the present. *Expert Opin. Ther. Pat.* **2014**, *25*, 49–68. [[CrossRef](#)] [[PubMed](#)]
116. Epstein, F.H.; Moncada, S.; Higgs, A. The L-Arginine-Nitric Oxide Pathway. *N. Engl. J. Med.* **1993**, *329*, 2002–2012. [[CrossRef](#)]
117. Song, P.; Zou, S.; Chen, T.; Chen, J.; Wang, Y.; Yang, J.; Song, Z.; Jiang, H.; Shi, H.; Huang, Y.; et al. Endothelial Nitric Oxide Synthase (eNOS) T-786C, 4a4b, and G894T Polymorphisms and Male Infertility: Study for Idiopathic Asthenozoospermia and Meta-Analysis1. *Biol. Reprod.* **2015**, *92*, 38. [[CrossRef](#)]
118. Agarwal, A.; Tvrda, E.; Sharma, R. Relationship amongst teratozoospermia, seminal oxidative stress and male infertility. *Reprod. Biol. Endocrinol.* **2014**, *12*, 45. [[CrossRef](#)] [[PubMed](#)]
119. Daiber, A.; Steven, S.; Vujacic-Mirski, K.; Kalinovic, S.; Oelze, M.; Di Lisa, F.; Münzel, T. Regulation of Vascular Function and Inflammation via Cross Talk of Reactive Oxygen and Nitrogen Species from Mitochondria or NADPH Oxidase—Implications for Diabetes Progression. *Int. J. Mol. Sci.* **2020**, *21*, 3405. [[CrossRef](#)] [[PubMed](#)]
120. Herrero, E.D.L.A.C.G.M.B.; De Lamirande, E.; Gagnon, C. Nitric Oxide is a Signaling Molecule in Spermatozoa. *Curr. Pharm. Des.* **2003**, *9*, 419–425. [[CrossRef](#)] [[PubMed](#)]
121. Buzadzic, B.; Vucetic, M.; Jankovic, A.; Stancic, A.; Korac, A.; Korac, B.; Otasevic, V. New insights into male (in)fertility: The importance of NO. *Br. J. Pharmacol.* **2015**, *172*, 1455–1467. [[CrossRef](#)]
122. Du Plessis, S.S.; McAllister, D.A.; Luu, A.; Savia, J.; Agarwal, A.; Lampiao, F. Effects of H2O2 exposure on human sperm motility parameters, reactive oxygen species levels and nitric oxide levels. *Andrologia* **2010**, *42*, 206–210. [[CrossRef](#)] [[PubMed](#)]
123. Chang, J.; Pan, F.; Tang, Q.; Wu, W.; Chen, M.; Lu, C.; Ding, H.; Hu, L.; Chen, D.; Xia, Y.; et al. eNOS gene T786C, G894T and 4a4b polymorphisms and male infertility susceptibility: A meta-analysis. *Andrologia* **2016**, *49*, e12646. [[CrossRef](#)]
124. Yun, Y.-J.; Park, J.-H.; Song, S.-H.; Lee, S. The association of 4a4b polymorphism of endothelial nitric oxide synthase (eNOS) gene with the sperm morphology in Korean infertile men. *Fertil. Steril.* **2008**, *90*, 1126–1131. [[CrossRef](#)] [[PubMed](#)]
125. Bianco, B.; Ghirelli-Filho, M.; Cavalheiro, C.M.; Cavalcanti, V.; Peluso, C.; Gava, M.M.; Glina, S.; Christofolini, D.M.; Barbosa, C.P. Variants in endothelial nitric oxide synthase (eNOS) gene in idiopathic infertile Brazilian men. *Gene* **2013**, *519*, 13–17. [[CrossRef](#)] [[PubMed](#)]
126. Buldreghini, E.; Mahfouz, R.Z.; Vignini, A.; Mazzanti, L.; Ricciardo-Lamonica, G.; Lenzi, A.; Agarwal, A.; Balercia, G. Single Nucleotide Polymorphism (SNP) of the Endothelial Nitric Oxide Synthase (eNOS) Gene (Glu298Asp Variant) in Infertile Men With Asthenozoospermia. *J. Androl.* **2010**, *31*, 482–488. [[CrossRef](#)] [[PubMed](#)]
127. Faure, C.; Leveille, P.; Dupont, C.; Julia, C.; Chavatte-Palmer, P.; Sutton, A.; Levy, R.; Alifert Group. Are Superoxide Dismutase 2 and Nitric Oxide Synthase Polymorphisms Associated with Idiopathic Infertility? *Antioxid. Redox Signal.* **2014**, *21*, 565–569. [[CrossRef](#)]

128. Mostafa, T.; Rashed, L.A.; Nabil, N.; Fouad, H.; Sabry, D.; El-Saied, D.M.; Bostamy, H. Endothelial Nitric Oxide Synthase Gene Polymorphism Relationship With Semen Parameters and Oxidative Stress in Infertile Oligoasthenoteratozoospermic Men. *Urology* **2015**, *85*, 1058–1061. [[CrossRef](#)]
129. Safarinejad, M.R.; Shafiei, N.; Safarinejad, S. The role of endothelial nitric oxide synthase (eNOS) T-786C, G894T, and 4a/b gene polymorphisms in the risk of idiopathic male infertility. *Mol. Reprod. Dev.* **2010**, *77*, 720–727. [[CrossRef](#)]
130. Ying, H.-Q.; Pu, X.-Y.; Liu, S.-R.; A, Z.-C. Genetic variants of eNOS gene may modify the susceptibility to idiopathic male infertility. *Biomarkers* **2013**, *18*, 412–417. [[CrossRef](#)]
131. Gruber, A.R.; Lorenz, R.; Bernhart, S.H.F.; Neuböck, R.; Hofacker, I.L. The Vienna RNA Websuite. *Nucleic Acids Res.* **2008**, *36*, W70–W74. [[CrossRef](#)] [[PubMed](#)]
132. Mousavi-Nasab, F.S.; Colagar, A.H. Investigation of the association of endothelial nitric oxide synthase (eNOS)-T786C gene polymorphism with the risk of male infertility in an Iranian population. *Environ. Sci. Pollut. Res.* **2020**, *27*, 22434–22440. [[CrossRef](#)] [[PubMed](#)]
133. Aitken, R.J.; Gibb, Z.; Baker, M.A.; Drevet, J.R.; Gharagozloo, P. Causes and consequences of oxidative stress in spermatozoa. *Reprod. Fertil. Dev.* **2016**, *28*, 1–10. [[CrossRef](#)]
134. Aitken, R.J.; Smith, T.B.; Jobling, M.S.; Baker, M.A.; De Iulius, G.N. Oxidative stress and male reproductive health. *Asian J. Androl.* **2014**, *16*, 31–38. [[CrossRef](#)] [[PubMed](#)]
135. Rodríguez, A.G.; De La Casa, M.; Johnston, S.; Gosálvez, J.; Roy, R. Association of polymorphisms in genes coding for antioxidant enzymes and human male infertility. *Ann. Hum. Genet.* **2019**, *83*, 63–72. [[CrossRef](#)] [[PubMed](#)]
136. Li, G.; Wang, X.; Yang, H.; Zhang, P.; Wu, F.; Li, Y.; Zhou, Y.; Zhang, X.; Ma, H.; Zhang, W.; et al. α -Linolenic acid but not linolenic acid protects against hypertension: Critical role of SIRT3 and autophagic flux. *Cell Death Dis.* **2020**, *11*, 83. [[CrossRef](#)]
137. Ruiz-Sanz, J.I.; Aurrekoetxea, I.; Matorras, R.; Ruiz-Larrea, M.B. Ala16Val SOD2 polymorphism is associated with higher pregnancy rates in in vitro fertilization cycles. *Fertil. Steril.* **2011**, *95*, 1601–1605. [[CrossRef](#)]
138. Yan, L.; Liu, J.; Wu, S.; Zhang, S.; Ji, G.; Gu, A. Seminal superoxide dismutase activity and its relationship with semen quality and SOD gene polymorphism. *J. Assist. Reprod. Genet.* **2014**, *31*, 549–554. [[CrossRef](#)] [[PubMed](#)]
139. Tefik, T.; Kucukgergin, C.; Sanli, O.; Oktar, T.; Seckin, S.; Ozsoy, C. Manganese superoxide dismutase Ile58Thr, catalase C-262T and myeloperoxidase G-463A gene polymorphisms in patients with prostate cancer: Relation to advanced and metastatic disease. *BJU Int.* **2013**, *112*, E406–E414. [[CrossRef](#)] [[PubMed](#)]
140. Bousnane, N.E.H.; Sadiq, M.; Bousnane, Z.; Labed, H.; Abu Alhaja, A.A.; Ayache, H.; Yahia, M. Study of the association between SOD3G362A polymorphism, seminal SOD activity and risk for idiopathic male infertility in Algeria. *Rev. Int. Androl.* **2020**. [[CrossRef](#)]
141. Morielli, T.; O’Flaherty, C. Oxidative stress impairs function and increases redox protein modifications in human spermatozoa. *Reproduction* **2015**, *149*, 113–123. [[CrossRef](#)]
142. Van Der Wal, A.C.; Das, P.K.; Tigges, A.J.; Becker, A.E. Adhesion molecules on the endothelium and mononuclear cells in human atherosclerotic lesions. *Am. J. Pathol.* **1992**, *141*, 1427–1433.
143. Tzoulaki, I.; Murray, G.D.; Lee, A.J.; Rumley, A.; Lowe, G.D.; Fowkes, F.G.R. C-Reactive Protein, Interleukin-6, and Soluble Adhesion Molecules as Predictors of Progressive Peripheral Atherosclerosis in the General Population. *Circulation* **2005**, *112*, 976–983. [[CrossRef](#)]
144. Souter, I.; Huang, A.; Martínez-Maza, O.; Breen, E.C.; DeCherney, A.H.; Chaudhuri, G.; Nathan, L. Serum levels of soluble vascular cell adhesion molecule-1, tumor necrosis factor- α , and interleukin-6 in in vitro fertilization cycles. *Fertil. Steril.* **2009**, *91*, 2012–2019. [[CrossRef](#)] [[PubMed](#)]
145. Jones, W.R. Immunology of infertility. *Clin. Obstet. Gynaecol.* **1981**, *8*, 611–639.
146. Santi, D.; Spaggiari, G.; Casonati, A.; Casarini, L.; Grassi, R.; Vecchi, B.; Roli, L.; De Santis, M.C.; Orlando, G.; Gravotta, E.; et al. Multilevel approach to male fertility by machine learning highlights a hidden link between haematological and spermatogenic cells. *Andrology* **2020**, *8*, 1021–1029. [[CrossRef](#)]
147. Aghazarian, A.; Stancik, I.; Huf, W.; Pflüger, H. Evaluation of Leukocyte Threshold Values in Semen to Detect Inflammation Involving Seminal Interleukin-6 and Interleukin-8. *Urology* **2015**, *86*, 52–56. [[CrossRef](#)]
148. Domes, T.; Lo, K.C.; Grober, E.D.; Mullen, J.B.M.; Mazzulli, T.; Jarvi, K. The incidence and effect of bacteriospermia and elevated seminal leukocytes on semen parameters. *Fertil. Steril.* **2012**, *97*, 1050–1055. [[CrossRef](#)]
149. Barraud-Lange, V.; Pont, J.-C.; Ziyyat, A.; Pocate, K.; Sifer, C.; Cedrin-Durnerin, I.; Fechtali, B.; Ducot, B.; Wolf, J.P. Seminal leukocytes are Good Samaritans for spermatozoa. *Fertil. Steril.* **2011**, *96*, 1315–1319. [[CrossRef](#)] [[PubMed](#)]
150. Barraud-Lange, V.; Pont, J.-C.; Pocate, K.; Kunstmann, J.M.; Chalas-Boissonas, C.; Ducot, B.; Wolf, J.P. Seminal leukocytes and clinical outcomes with donor sperm insemination. *Fertil. Steril.* **2011**, *96*, 1320–1324.e1. [[CrossRef](#)] [[PubMed](#)]
151. Lackner, J.; Schatzl, G.; Horvath, S.; Kratzik, C.; Marberger, M. Value of Counting White Blood Cells (WBC) in Semen Samples to Predict the Presence of Bacteria. *Eur. Urol.* **2006**, *49*, 148–153. [[CrossRef](#)] [[PubMed](#)]
152. Lotti, F.; Corona, G.; Mancini, M.; Filimberti, E.; Degli Innocenti, S.; Colpi, G.M.; Baldi, E.; Noci, I.; Forti, G.; Adorini, L.; et al. Ultrasonographic and clinical correlates of seminal plasma interleukin-8 levels in patients attending an andrology clinic for infertility. *Int. J. Androl.* **2011**, *34*, 600–613. [[CrossRef](#)]

153. Jedrzejczak, P.; Fraczek, M.; Szumala-Kakol, A.; Taszarek-Hauke, G.; Pawelczyk, L.; Kurpisz, M. Consequences of semen inflammation and lipid peroxidation on fertilization capacity of spermatozoa in in vitro conditions. *Int. J. Androl.* **2005**, *28*, 275–283. [[CrossRef](#)]
154. Motrich, R.D.; Bresler, M.L.; Molina, R.I.; Tissera, A.; Olmedo, J.J.; Rivero, V.E. Chronic Prostatitis/Chronic Pelvic Pain Syndrome patients show Th1 and Th17 self-reactive immune responses specific to prostate and seminal antigens and decreased semen quality. *BJU Int.* **2020**, *126*, 379–387. [[CrossRef](#)] [[PubMed](#)]
155. Seshadri, S.; Bates, M.; Vince, G.; Jones, D.I.L. ORIGINAL ARTICLE: The Role of Cytokine Expression in Different Subgroups of Subfertile Men. *Am. J. Reprod. Immunol.* **2009**, *62*, 275–282. [[CrossRef](#)]
156. Maegawa, M.; Kamada, M.; Irahara, M.; Yamamoto, S.; Yoshikawa, S.; Kasai, Y.; Ohmoto, Y.; Gima, H.; Thaler, C.J.; Aono, T. A repertoire of cytokines in human seminal plasma. *J. Reprod. Immunol.* **2002**, *54*, 33–42. [[CrossRef](#)]
157. Furuya, Y.; Akashi, T.; Fuse, H. Soluble Fas and interleukin-6 and interleukin-8 levels in seminal plasma of infertile men. *Arch. Androl.* **2003**, *49*, 449–452. [[CrossRef](#)]
158. Friebe, K.; Bohring, C.; Skrzypek, J.; Krause, W. Levels of interleukin-6 and interleukin-8 in seminal fluid of men attending an andrological clinic. *Andrologia* **2003**, *35*, 126–129. [[CrossRef](#)]
159. Koumantakis, E.; Matalliotakis, I.; Kyriakou, D.; Fragouli, Y.; Relakis, K. Increased levels of interleukin-8 in human seminal plasma. *Andrologia* **2009**, *30*, 339–343. [[CrossRef](#)] [[PubMed](#)]
160. Hærvig, K.K.; Kierkegaard, L.; Lund, R.; Bruunsgaard, H.; Osler, M.; Schmidt, L. Is male factor infertility associated with midlife low-grade inflammation? A population based study. *Hum. Fertil.* **2017**, *21*, 146–154. [[CrossRef](#)]
161. Camejo, M.I.; Segnini, A.; Proverbio, F. Interleukin-6 (IL-6) in seminal plasma of infertile men, and lipid peroxidation of their sperm. *Arch. Androl.* **2001**, *47*, 97–101. [[CrossRef](#)]
162. Moretti, E.; Collodel, G.; Mazzi, L.; Campagna, M.; Iacoponi, F.; Figura, N. Resistin, interleukin-6, tumor necrosis factor- α , and human semen parameters in the presence of leukocytospermia, smoking habit, and varicocele. *Fertil. Steril.* **2014**, *102*, 354–360. [[CrossRef](#)] [[PubMed](#)]
163. Bobjer, J.; Katrinaki, M.; Tsatsanis, C.; Giwercman, Y.L.; Giwercman, A. Negative Association between Testosterone Concentration and Inflammatory Markers in Young Men: A Nested Cross-Sectional Study. *PLoS ONE* **2013**, *8*, e61466. [[CrossRef](#)]
164. Agarwal, A.; Selvam, M.K.P.; Baskaran, S. Proteomic analysis of seminal plasma from bilateral varicocele patients indicates an oxidative state and increased inflammatory response. *Asian J. Androl.* **2019**, *21*, 544–550. [[CrossRef](#)] [[PubMed](#)]
165. Eldamhoury, E.M.; Elatrash, G.A.; Rashwan, H.M.; El-Sakka, A.I. Association between leukocytospermia and semen interleukin-6 and tumor necrosis factor- α in infertile men. *Andrology* **2018**, *6*, 775–780. [[CrossRef](#)] [[PubMed](#)]
166. Kurkowska, W.; Bogacz, A.; Janiszewska, M.; Gabryś, E.; Tiszler, M.; Bellanti, F.; Kasperczyk, S.; Machoń-Grecka, A.; Dobrakowski, M.; Kasperczyk, A. Oxidative Stress is Associated with Reduced Sperm Motility in Normal Semen. *Am. J. Men's Health* **2020**, *14*, 1557988320939731. [[CrossRef](#)]
167. Leisegang, K.; Bouic, P.J.D.; Henkel, R.R. Metabolic syndrome is associated with increased seminal inflammatory cytokines and reproductive dysfunction in a case-controlled male cohort. *Am. J. Reprod. Immunol.* **2016**, *76*, 155–163. [[CrossRef](#)]
168. Eggert-Kruse, W.; Kiefer, I.; Beck, C.; Demirakca, T.; Strowitzki, T. Role for tumor necrosis factor α (TNF- α) and interleukin 1- β (IL-1 β) determination in seminal plasma during infertility investigation. *Fertil. Steril.* **2007**, *87*, 810–823. [[CrossRef](#)]
169. Moretti, E.; Cosci, I.; Spreafico, A.; Serchi, T.; Cuppone, A.M.; Collodel, G. Semen characteristics and inflammatory mediators in infertile men with different clinical diagnoses. *Int. J. Androl.* **2009**, *32*, 637–646. [[CrossRef](#)]
170. Cai, Y.; Xiong, C.; Shen, S.; Rao, J.; Liu, T.; Qiu, F. Mesenchymal stem cell-secreted factors delayed spermatogenesis injuries induced by busulfan involving intercellular adhesion molecule regulation. *Andrologia* **2019**, *51*, e13285. [[CrossRef](#)] [[PubMed](#)]
171. Xiao, X.; Mruk, D.D.; Cheng, C.Y. Intercellular adhesion molecules (ICAMs) and spermatogenesis. *Hum. Reprod. Update* **2013**, *19*, 167–186. [[CrossRef](#)]
172. Piprek, R.P.; Kloc, M.; Mizia, P.; Kubiak, J.Z. The Central Role of Cadherins in Gonad Development, Reproduction, and Fertility. *Int. J. Mol. Sci.* **2020**, *21*, 8264. [[CrossRef](#)] [[PubMed](#)]
173. Cavallaro, U.; Dejana, E. Adhesion molecule signalling: Not always a sticky business. *Nat. Rev. Mol. Cell Biol.* **2011**, *12*, 189–197. [[CrossRef](#)]
174. Xiao, X.; Cheng, C.Y.; Mruk, L.D. Intercellular adhesion molecule-1 is a regulator of blood-testis barrier function. *J. Cell Sci.* **2012**, *125*, 5677–5689. [[CrossRef](#)]
175. Van Buul, J.D.; Van Rijssel, J.; Van Alphen, F.P.J.; Van Stalborch, A.-M.; Mul, E.P.J.; Hordijk, P.L. ICAM-1 Clustering on Endothelial Cells Recruits VCAM-1. *J. Biomed. Biotechnol.* **2010**, *2010*, 1–9. [[CrossRef](#)]
176. Cybulsky, M.I.; Gimbrone, M.A. Endothelial expression of a mononuclear leukocyte adhesion molecule during atherogenesis. *Science* **1991**, *251*, 788–791. [[CrossRef](#)] [[PubMed](#)]
177. Li, H.; Cybulsky, M.I.; Gimbrone, M.A.; Libby, P. Inducible expression of vascular cell adhesion molecule-1 by vascular smooth muscle cells in vitro and within rabbit atheroma. *Am. J. Pathol.* **1993**, *143*, 1551–1559. [[PubMed](#)]
178. Pigott, R.; Dillon, L.; Hemingway, I.; Gearing, A. Soluble forms of E-selectin, ICAM-1 and VCAM-1 are present in the supernatants of cytokine activated cultured endothelial cells. *Biochem. Biophys. Res. Commun.* **1992**, *187*, 584–589. [[CrossRef](#)]
179. Brady, H.R. Leukocyte adhesion molecules and kidney diseases. *Kidney Int.* **1994**, *45*, 1285–1300. [[CrossRef](#)] [[PubMed](#)]
180. Von Der Thüsen, J.H.; Kuiper, J.; Van Berkel, T.J.C.; Biessen, E.A.L. Interleukins in Atherosclerosis: Molecular Pathways and Therapeutic Potential. *Pharmacol. Rev.* **2003**, *55*, 133–166. [[CrossRef](#)] [[PubMed](#)]

181. Riccioli, A.; Filippini, A.; De Cesaris, P.; Barbacci, E.; Stefanini, M.; Starace, G.; Ziparo, E. Inflammatory mediators increase surface expression of integrin ligands, adhesion to lymphocytes, and secretion of interleukin 6 in mouse Sertoli cells. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 5808–5812. [[CrossRef](#)] [[PubMed](#)]
182. Berruti, G.; Ceriani, M.; Martegani, E. Dynamic of VE-cadherin-mediated spermatid–Sertoli cell contacts in the mouse seminiferous epithelium. *Histochem. Cell Biol.* **2018**, *150*, 173–185. [[CrossRef](#)]
183. Aivatiadou, E.; Mattei, E.; Ceriani, M.; Tilia, L.; Berruti, G. Impaired Fertility and Spermiogenetic Disorders with Loss of Cell Adhesion in Male Mice Expressing an Interfering Rap1 Mutant. *Mol. Biol. Cell* **2007**, *18*, 1530–1542. [[CrossRef](#)]
184. Lo, A.C.; Fung, M.K.; Au, C.; Chan, T.S.; Sauer, B.; Chung, S.S.; Chung, S.K. Transgenic mice over-expressing endothelin-1 in testis transactivated by a Cre/loxP system showed decreased testicular capillary blood flow. *Transgenic Res.* **2004**, *13*, 119–134. [[CrossRef](#)] [[PubMed](#)]
185. Ergün, S.; Luttmmer, W.; Fiedler, W.; Holstein, A.F. Functional expression and localization of vascular endothelial growth factor and its receptors in the human epididymis. *Biol. Reprod.* **1998**, *58*, 160–168. [[CrossRef](#)]
186. Romanelli, F.; Fillo, S.; Isidori, A.; Conte, D. Stimulatory action of endothelin-1 on rat Leydig cells: Involvement of endothelin-A subtype receptor and phospholipase A2-arachidonate metabolism system. *Life Sci.* **1997**, *61*, 557–566. [[CrossRef](#)]
187. Tripiciano, A.; Filippini, A.; Giustiniani, Q.; Palombi, F. Direct Visualization of Rat Peritubular Myoid Cell Contraction in Response to Endothelin1. *Biol. Reprod.* **1996**, *55*, 25–31. [[CrossRef](#)] [[PubMed](#)]
188. Peri, A.; Fantoni, G.; Granchi, S.; Vannelli, G.B.; Barni, T.; Amerini, S.; Pupilli, C.; Barbagli, G.; Forti, G.; Serio, M.; et al. Gene Expression of Endothelin-1, Endothelin-Converting Enzyme-1, and Endothelin Receptors in Human Epididymis 1. *J. Clin. Endocrinol. Metab.* **1997**, *82*, 3797–3806. [[CrossRef](#)]
189. Hammami, M.M.; Haq, A.; AlSedairy, S. The level of endothelin-like immunoreactivity in seminal fluid correlates positively with semen volume and negatively with plasma gonadotrophin levels. *Clin. Endocrinol.* **2010**, *40*, 361–366. [[CrossRef](#)]
190. Kamada, S.; Oehninger, S.; Mahony, M.C.; Blackmore, P.F.; Lanzendorf, S.E.; Hodgen, G.D. Does Endothelin-1 Affect Human Spermatozoa Function? *Am. J. Reprod. Immunol.* **1994**, *31*, 91–98. [[CrossRef](#)] [[PubMed](#)]
191. Sargent, K.M.; Clopton, D.T.; Lu, N.; Pohlmeier, W.E.; Cupp, A.S. VEGFA splicing: Divergent isoforms regulate spermatogonial stem cell maintenance. *Cell Tissue Res.* **2015**, *363*, 31–45. [[CrossRef](#)] [[PubMed](#)]
192. Giurdanella, G.; Lupo, G.; Gennuso, F.; Conti, F.; Furno, D.L.; Mannino, G.; Anfuso, C.D.; Drago, F.; Salomone, S.; Bucolo, C. Activation of the VEGF-A/ERK/PLA2 Axis Mediates Early Retinal Endothelial Cell Damage Induced by High Glucose: New Insight from an In Vitro Model of Diabetic Retinopathy. *Int. J. Mol. Sci.* **2020**, *21*, 7528. [[CrossRef](#)] [[PubMed](#)]
193. Minutoli, L.; Antonuccio, P.; Squadrito, F.; Bitto, A.; Nicòtina, P.A.; Fazzari, C.; Polito, F.; Marini, H.R.; Bonvissuto, G.; Arena, S.; et al. Effects of polydeoxyribonucleotide on the histological damage and the altered spermatogenesis induced by testicular ischaemia and reperfusion in rats. *Int. J. Androl.* **2011**, *35*, 133–144. [[CrossRef](#)]
194. Hashimoto, H.; Ishikawa, T.; Yamaguchi, K.; Shiotani, M.; Fujisawa, M. Experimental ischaemia-reperfusion injury induces vascular endothelial growth factor expression in the rat testis. *Andrologia* **2009**, *41*, 216–221. [[CrossRef](#)] [[PubMed](#)]
195. Lu, N.; Sargent, K.M.; Clopton, D.T.; Pohlmeier, W.E.; Brauer, V.M.; McFee, R.M.; Weber, J.S.; Ferrara, N.; Silversides, D.W.; Cupp, A.S. Loss of Vascular Endothelial Growth Factor A (VEGFA) Isoforms in the Testes of Male Mice Causes Subfertility, Reduces Sperm Numbers, and Alters Expression of Genes That Regulate Undifferentiated Spermatogonia. *Endocrinology* **2013**, *154*, 4790–4802. [[CrossRef](#)] [[PubMed](#)]
196. Tian, R.; Yang, S.; Zhu, Y.; Zou, S.; Li, P.; Wang, J.; Zhu, Z.; Huang, Y.; He, Z.; Li, Z. VEGF/VEGFR2 Signaling Regulates Germ Cell Proliferation in vitro and Promotes Mouse Testicular Regeneration in vivo. *Cells Tissues Organs* **2016**, *201*, 1–13. [[CrossRef](#)] [[PubMed](#)]
197. Abdel-Aziz, A.M.; Hafez, S.M.N.A. Sitagliptin protects male albino rats with testicular ischaemia/reperfusion damage: Modulation of VCAM-1 and VEGF-A. *Andrologia* **2019**, *52*, e13472. [[CrossRef](#)]
198. Adlanmerini, M.; Fébrissy, C.; Zahreddine, R.; Vessières, E.; Buscato, M.; Solinhac, R.; Favre, J.; Anquetil, T.; Guihot, A.-L.; Boudou, F.; et al. Mutation of Arginine 264 on ER α (Estrogen Receptor Alpha) Selectively Abrogates the Rapid Signaling of Estradiol in the Endothelium Without Altering Fertility. *Arter. Thromb. Vasc. Biol.* **2020**, *40*, 2143–2158. [[CrossRef](#)]
199. Goswami, D.; Conway, G.S. Premature Ovarian Failure. *Horm. Res. Paediatr.* **2007**, *68*, 196–202. [[CrossRef](#)]
200. Jacobsen, B.K.; Knutsen, S.F.; Fraser, G.E. Age at Natural Menopause and Total Mortality and Mortality from Ischemic Heart Disease. *J. Clin. Epidemiol.* **1999**, *52*, 303–307. [[CrossRef](#)]
201. Hu, F.B.; Grodstein, F.; Hennekens, C.H.; Colditz, G.A.; Johnson, M.; Manson, J.E.; Rosner, B.; Stampfer, M.J. Age at Natural Menopause and Risk of Cardiovascular Disease. *Arch. Intern. Med.* **1999**, *159*, 1061–1066. [[CrossRef](#)]
202. Kalantaridou, S.N.; Naka, K.K.; Papanikolaou, E.; Kazakos, N.; Kravariti, M.; Calis, K.A.; Paraskevaidis, E.A.; Sideris, D.A.; Tsatsoulis, A.; Chrousos, G.P.; et al. Impaired Endothelial Function in Young Women with Premature Ovarian Failure: Normalization with Hormone Therapy. *J. Clin. Endocrinol. Metab.* **2004**, *89*, 3907–3913. [[CrossRef](#)]
203. Yorgun, H.; Tokgözoğlu, L.; Canpolat, U.; Gürses, K.M.; Bozdağ, G.; Yapıcı, Z.; Şahiner, L.; Kaya, E.B.; Kabakçı, G.; Oto, A.; et al. The cardiovascular effects of premature ovarian failure. *Int. J. Cardiol.* **2013**, *168*, 506–510. [[CrossRef](#)]
204. Goldmeier, S.; De Angelis, K.; Casali, K.R.; Vilodre, C.; Consolim-Colombo, F.; Klein, A.B.; Plentz, R.; Spritzer, P.; Irigoyen, M.-C. Cardiovascular autonomic dysfunction in primary ovarian insufficiency: Clinical and experimental evidence. *Am. J. Transl. Res.* **2013**, *6*, 91–101.

205. Vos, T.; Flaxman, A.D.; Naghavi, M.; Lozano, R.; Michaud, C.; Ezzati, M.; Shibuya, K.; Salomon, J.A.; Abdalla, S.; Aboyans, V.; et al. Years lived with disability (YLDs) for 1160 sequelae of 289 diseases and injuries 1990–2010: A systematic analysis for the Global Burden of Disease Study 2010. *Lancet* **2012**, *380*, 2163–2196. [[CrossRef](#)]
206. Norman, R.J.; Dewailly, D.; Legro, R.S.; Hickey, T.E. Polycystic ovary syndrome. *Lancet* **2007**, *370*, 685–697. [[CrossRef](#)]
207. Paradisi, G.; Steinberg, H.O.; Hempfling, A.; Cronin, J.; Hook, G.; Shepard, M.K.; Baron, A.D. Polycystic ovary syndrome is associated with endothelial dysfunction. *Circulation* **2001**, *103*, 1410–1415. [[CrossRef](#)] [[PubMed](#)]
208. Carmina, E. Cardiovascular risk and events in polycystic ovary syndrome. *Climacteric* **2009**, *12*, 22–25. [[CrossRef](#)] [[PubMed](#)]
209. Heidari, B.; Lerman, A.; Lalia, A.Z.; Lerman, L.O.; Chang, A.Y. Effect of Metformin on Microvascular Endothelial Function in Polycystic Ovary Syndrome. *Mayo Clin. Proc.* **2019**, *94*, 2455–2466. [[CrossRef](#)]
210. Hurliman, A.; Brown, J.K.; Maille, N.; Mandala, M.; Casson, P.; Osol, G. Hyperandrogenism and Insulin Resistance, Not Changes in Body Weight, Mediate the Development of Endothelial Dysfunction in a Female Rat Model of Polycystic Ovary Syndrome (PCOS). *Endocrinology* **2015**, *156*, 4071–4080. [[CrossRef](#)]
211. Dube, R. Does endothelial dysfunction correlate with endocrinal abnormalities in patients with polycystic ovary syndrome? *Avicenna J. Med.* **2016**, *6*, 91–102. [[CrossRef](#)]
212. Oncul, M.; Albayrak, M.; Sozer, V.; Karakus, B.; Gelisgen, R.; Karatas, S.; Simsek, G.; Uzun, H. Polycystic ovary syndrome and endothelial dysfunction: A potential role for soluble lectin-like oxidized low density lipoprotein receptor-1. *Reprod. Biol.* **2020**, *20*, 396–401. [[CrossRef](#)]
213. Usselman, C.W.; Yarovinsky, T.O.; Steele, F.E.; Leone, C.A.; Taylor, H.S.; Bender, J.R.; Stachenfeld, N.S. Androgens drive microvascular endothelial dysfunction in women with polycystic ovary syndrome: Role of the endothelin B receptor. *J. Physiol.* **2019**, *597*, 2853–2865. [[CrossRef](#)] [[PubMed](#)]
214. Kao, Y.-H.; Chiu, W.-C.; Hsu, M.-I.; Chen, Y.-J. Endothelial Progenitor Cell Dysfunction in Polycystic Ovary Syndrome: Implications for The Genesis of Cardiovascular Diseases. *Int. J. Fertil. Steril.* **2013**, *6*, 208–213.
215. Dambala, K.; Paschou, S.A.; Michopoulos, A.; Siasos, G.; Goulis, D.G.; Vavilis, D.; Tarlatzis, B.C. Biomarkers of Endothelial Dysfunction in Women With Polycystic Ovary Syndrome. *Angiology* **2019**, *70*, 797–801. [[CrossRef](#)]
216. Laschke, M.W.; Menger, M.D. Basic mechanisms of vascularization in endometriosis and their clinical implications. *Hum. Reprod. Update* **2018**, *24*, 207–224. [[CrossRef](#)] [[PubMed](#)]
217. Barcz, E.; Rózewska, E.S.; Kaminski, P.; Demkow, U.; Bobrowska, K.; Marianowski, L. Angiogenic activity and IL-8 concentrations in peritoneal fluid and sera in endometriosis. *Int. J. Gynecol. Obstet.* **2002**, *79*, 229–235. [[CrossRef](#)]
218. Sokolov, D.I.; Solodovnikova, N.G.; Pavlov, O.V.; Niauri, D.A.; Volkov, N.N.; Sel'Kov, S.A. Study of Cytokine Profile and Angiogenic Potential of Peritoneal Fluid in Patients with External Genital Endometriosis. *Bull. Exp. Biol. Med.* **2005**, *140*, 541–544. [[CrossRef](#)]
219. Santoro, L.; D'Onofrio, F.; Flore, R.A.; Gasbarrini, A.; Santoliquido, A. Endometriosis and atherosclerosis: What we already know and what we have yet to discover. *Am. J. Obstet. Gynecol.* **2015**, *213*, 326–331. [[CrossRef](#)]
220. Santoro, L.; D'Onofrio, F.; Campo, S.; Ferraro, P.M.; Tondi, P.; Campo, V.; Flex, A.; Gasbarrini, A.; Santoliquido, A. Endothelial dysfunction but not increased carotid intima-media thickness in young European women with endometriosis. *Hum. Reprod.* **2012**, *27*, 1320–1326. [[CrossRef](#)] [[PubMed](#)]
221. Cui, L.-L.; Yang, G.; Pan, J.; Zhang, C. Tumor necrosis factor α knockout increases fertility of mice. *Theriogenology* **2011**, *75*, 867–876. [[CrossRef](#)]
222. Alijotas-Reig, J.; Esteve-Valverde, E.; Ferrer-Oliveras, R.; Llorba, E.; Gris, J.M. Tumor Necrosis Factor-Alpha and Pregnancy: Focus on Biologics. An Updated and Comprehensive Review. *Clin. Rev. Allergy Immunol.* **2017**, *53*, 40–53. [[CrossRef](#)] [[PubMed](#)]
223. Bansal, R.; Ford, B.; Bhaskaran, S.; Thum, M.; Bansal, A. Elevated Levels of Serum Vascular Endothelial Growth Factor-A Are Not Related to NK Cell Parameters in Recurrent IVF Failure. *J. Reprod. Infertil.* **2017**, *18*, 280–287. [[PubMed](#)]
224. Chen, H.; Center, Y.H.R.M.; Zheng, J.-B.; Wang, N.-M.; Xing, H.; Wang, H. Association between Vascular Endothelial Growth Factor and Clinical Outcomes of IVF-ET/ICSI. *J. Coll. Physicians Surg. Pak.* **2019**, *29*, 19–23. [[CrossRef](#)]
225. Nikolettos, N.; Asimakopoulos, B.; Nicolettos, N.; Efthimiadou, A.; Mourvati, E.; Demirel, C. Evaluation of leptin, interleukin-1beta, tumor necrosis factor-alpha and vascular endothelial growth factor in serum and follicular fluids of women undergoing controlled ovarian hyperstimulation as prognostic markers of ICSI outcome. *In Vivo* **2004**, *18*, 667–673. [[PubMed](#)]
226. Lédée-Bataille, N.; Bonnet-Chea, K.; Hosny, G.; Dubanchet, S.; Frydman, R.; Chaouat, G. Role of the endometrial tripod interleukin-18, -15, and -12 in inadequate uterine receptivity in patients with a history of repeated in vitro fertilization–embryo transfer failure. *Fertil. Steril.* **2005**, *83*, 598–605. [[CrossRef](#)]
227. Inagaki, N.; Stern, C.; McBain, J.; Lopata, A.; Kornman, L.; Wilkinson, D. Analysis of intra-uterine cytokine concentration and matrix-metalloproteinase activity in women with recurrent failed embryo transfer. *Hum. Reprod.* **2003**, *18*, 608–615. [[CrossRef](#)]
228. Liu, H.-Y.; Liu, Z.-K.; Chao, H.; Li, Z.; Song, Z.; Yang, Y.; Peng, J.-P. High-Dose Interferon- γ Promotes Abortion in Mice by Suppressing Treg and Th17 Polarization. *J. Interferon Cytokine Res.* **2014**, *34*, 394–403. [[CrossRef](#)]
229. Fiçıoğlu, C.; Kumbak, B.; Akcin, O.; Attar, R.; Yıldırım, G.; Tecelioğlu, N.; Yeşildağlar, N. Cytokine and nitric oxide concentrations in follicular fluid and blood serum of patients undergoing assisted reproductive treatment: Relationship to outcome. *J. Turk. Gynecol. Assoc.* **2009**, *10*, 132–136.

230. Asimakopoulos, B.; Abu-Hassan, D.; Metzen, E.; Al-Hasani, S.; Diedrich, K.; Nikolettos, N. The levels of steroid hormones and cytokines in individual follicles are not associated with the fertilization outcome after intracytoplasmic sperm injection. *Fertil. Steril.* **2008**, *90*, 60–64. [[CrossRef](#)]
231. Xu, X.; Du, C.; Li, H.; Du, J.; Yan, X.; Peng, L.; Li, G.; Chen, Z.-J. Association of VEGF Genetic Polymorphisms with Recurrent Spontaneous Abortion Risk: A Systematic Review and Meta-Analysis. *PLOS ONE* **2015**, *10*, e0123696. [[CrossRef](#)] [[PubMed](#)]
232. Shim, S.H.; Kim, J.O.; Jeon, Y.J.; An, H.J.; Lee, H.A.; Kim, J.H.; Ahn, E.H.; Lee, W.S.; Kim, N.K. Association between vascular endothelial growth factor promoter polymorphisms and the risk of recurrent implantation failure. *Exp. Ther. Med.* **2017**, *15*, 2109–2119. [[CrossRef](#)]
233. Boudjenah, R.; Molina-Gomes, D.; Torre, A.; Boitrelle, F.; Taieb, S.; Dos Santos, E.; Wainer, R.; De Mazancourt, P.; Selva, J.; Vialard, F. Associations between Individual and Combined Polymorphisms of the TNF and VEGF Genes and the Embryo Implantation Rate in Patients Undergoing In Vitro Fertilization (IVF) Programs. *PLoS ONE* **2014**, *9*, e108287. [[CrossRef](#)]
234. Vialard, F.; El Sirkasi, M.; Tronchon, V.; Boudjenah, R.; Molina-Gomes, D.; Bergere, M.; Mauduit, C.; Wainer, R.; Selva, J.; Benahmed, M. Tumor necrosis factor-308 polymorphism increases the embryo implantation rate in women undergoing in vitro fertilization. *Hum. Reprod.* **2013**, *28*, 2774–2783. [[CrossRef](#)]
235. Turienzo, A.; Lledó, B.; Ortiz, J.A.; Morales, R.; Sanz, J.; Llácer, J.; Bernabeu, R. Prevalence of candidate single nucleotide polymorphisms on p53, IL-11, IL-10, VEGF and APOE in patients with repeated implantation failure (RIF) and pregnancy loss (RPL). *Hum. Fertil.* **2018**, *23*, 117–122. [[CrossRef](#)] [[PubMed](#)]
236. Gagliano-Jucá, T.; Basaria, S. Testosterone replacement therapy and cardiovascular risk. *Nat. Rev. Cardiol.* **2019**, *16*, 555–574. [[CrossRef](#)]
237. Moradi, M.-T.; Rahimi, Z.; Vaisi-Raygani, A. New insight into the role of long non-coding RNAs in the pathogenesis of preeclampsia. *Hypertens. Pregnancy* **2019**, *38*, 41–51. [[CrossRef](#)] [[PubMed](#)]