CD4 cell count response to first-line combination ART in HIV-2+ patients compared with HIV-1+ patients: a multinational, multicohort European study

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© The Author 2017. Published by Oxford University Press on behalf of the British Society for Antimicrobial Chemotherapy. All rights reserved. For Permissions, please email: journals.permissions@oup.com. **Background:** CD4 cell recovery following first-line combination ART (cART) is poorer in HIV-2+ than in HIV-1+ patients. Only large comparisons may allow adjustments for demographic and pretreatment plasma viral load (pVL).

Methods: ART-naive HIV+ adults from two European multicohort collaborations, COHERE (HIV-1 alone) and ACHIeV2e (HIV-2 alone), were included, if they started first-line cART (without NNRTIS or fusion inhibitors) between 1997 and 2011. Patients without at least one CD4 cell count before start of cART, without a pretreatment pVL and with missing *a priori*-defined covariables were excluded. Evolution of CD4 cell count was studied using adjusted linear mixed models.

Results: We included 185 HIV-2+ and 30321 HIV-1+ patients with median age of 46 years (IQR 36-52) and 37 years (IQR 31-44), respectively. Median observed pretreatment CD4 cell counts/mm³ were 203 (95% CI 100-290) in HIV-2+ patients and 223 (95% CI 100-353) in HIV-1+ patients. Mean observed CD4 cell count changes from start of cART to 12 months were +105 (95% CI 77-134) in HIV-2+ patients and +202 (95% CI 199-205) in HIV-1+ patients, an observed difference of 97 cells/mm³ in 1 year. In adjusted analysis, the mean CD4 cell increase was overall 25 CD4 cells/mm³/year lower (95% CI 5-44; P = 0.0127) in HIV-2+ patients compared with HIV-1+ patients.

Conclusions: A poorer CD4 cell increase during first-line cART was observed in HIV-2+ patients, even after adjusting for pretreatment pVL and other potential confounders. Our results underline the need to identify more potent therapeutic regimens or strategies against HIV-2.

Introduction

Methods

ve Data collection

HIV-2 is less prevalent than HIV-1 and HIV-2+ individuals live mainly in West Africa, followed by Angola, Mozambique and Europe (primarily Portugal and France).¹ HIV-2 infection is characterized by a lower plasma viral load (pVL) and a slower clinical progression.²⁻⁷ ART options for HIV-2 are restricted due to its natural resistance to NNRTIs and fusion inhibitors, and because some PIs have shown lower efficacy.⁸⁻¹¹

The 2013 WHO guidelines for treatment of HIV-2 infection recommended either triple NRTIs or two NRTIs combined with a ritonavir-boosted PI (PI/r) as first-line combination ART (cART), with a lopinavir-containing regimen as preferred option.^{12,13} In Europe, the recommended first-line cART regimen for HIV-2 infection consisted of two NRTIs and one PI or one PI/r since 2010.^{14,15} Of note, performing a randomized clinical trial designed to inform about optimal cART strategies for HIV-2 therapy is not feasible in Europe due to the low prevalence of HIV-2 infection, but is ongoing in West Africa.

CD4 cell recovery in HIV-2+ patients receiving first-line cART has been reported to be lower and slower than in HIV-1+ patients.¹⁶⁻¹⁸ Two observational studies have reported a better immunological response in HIV-2+ patients with a PI/r-based regimen compared with a triple NRTI regimen.^{19,20}

Comparative studies between HIV-1 and HIV-2 are often hampered by the small number of patients infected with HIV-2 under similar standardized follow-up.^{3,21} Contradictory results regarding CD4 cell recovery have been reported when pretreatment pVL was taken into account.^{3,21,22} The lower replication rate of HIV-2 has been hypothesized as one possible explanation, leading to a poorer CD4 response to therapy. We compared immunological outcome in HIV-2+ and HIV-1+ patients under standard follow-up in Europe, by adjusting for pretreatment pVL levels at initiation of first cART, and other potential confounders.

COHERE (HIV-1+ patients) and ACHIeV2e (HIV-2+ patients) are prospective, multinational, observational cohort collaborations. Data were pooled in the COHERE in EuroCoord 2011 and ACHIeV2e 2011 data merger. COHERE is a collaboration of 40 cohorts from across Europe²³ and is part of the EuroCoord network (www.EuroCoord.net). The 23 cohorts participating in the present study through the COHERE network and the 9 cohortsparticipating through the ACHIeV2e network^{24,25} (listed in the Acknowledgements section) submitted a defined dataset (patient demographics, current cART, CD4 counts and HIV RNA values, clinical status and events) to their network-specific Coordination Centre, using the HIV Cohort Data Exchange Protocol (HICDEP).²⁶ The final data set was merged at the Bordeaux Regional Coordinating Centre for COHERE and ACHIeV2e, adhering to strict quality-assurance guidelines and performing data quality checks.

Study population

Adult patients aged >18 years infected with either HIV-2 or HIV-1 (dualseropositive and dual-infected patients were excluded) who started a firstline cART regimen from 1997 to 2011 were included in the analysis. HIV-2+ and HIV-1+ patients receiving an NNRTI- or fusion inhibitor-containing regimen were not included because of the natural resistance of HIV-2 to these drug classes.¹¹ Observations were excluded if patients presented without one pretreatment CD4 cell count in a window of 6 months before start of cART²⁷ and without a pretreatment pVL based on quantification methods with a detection limit of \leq 500 copies/mL. We used a cut-off of 500 copies/ mL (2.7 log₁₀ copies/mL) to define undetectable pVL, as a consensus shared by the majority of contributing centres in the dataset.^{24,25} Only patients with complete data for potential confounders (listed below) were included in all analyses (Figure S1, available as Supplementary data at JAC Online). Criteria used to initiate therapy of HIV-2 infection were those fitting the national treatment guidelines of each contributing centre effective at the time of cART start. The choice of antiretroviral combination was at the physician's discretion based on these treatment guidelines.

Follow-up began at initiation of the first cART regimen (baseline). Followup was censored when the ART combination was modified for whatever reason, at death or at the last available CD4 cell counts, whichever occurred first.

Virological data and CD4 cell count

All serological results, pVL values and CD4 cell counts were obtained from laboratories of participating centres. HIV-1 and HIV-2 infections were diagnosed by ELISA tests confirmed by western blot. Quantification of HIV-2 viral load was assessed by in-house methods;²⁵ the quality control assessments of quantification assays used by the ACHIeV2e network showed heterogeneity, which improved from 2006 to 2011.^{24,25} Pretreatment pVL and pretreatment CD4 cell counts were defined as the closest measurement in a window of 6 months before cART start.

Statistical analysis

We studied the effect of the HIV type on CD4 cell count at initiation of first cART (intercept) and on CD4 cell count change (cells/mm³/year) (slope) using linear mixed effect models with a random intercept and a random slope. The correlation between individual baseline CD4 value(s) and the subsequent CD4 slope(s) was handled through an unstructured covariance matrix of random effects. We checked the underlying model assumptions (normality and homoscedasticity of residuals).²⁸

We studied the effect of the following *a priori*-defined covariables on CD4 cell count at cART initiation and on CD4 cell count change by introducing an interaction term with the slope: HIV type (HIV-2 versus HIV-1, the main exposure variable), pVL as a continuous covariable (log₁₀ copies/mL with imputation of the limit of detection for undetectable pVL), age (per 10 year increase), sex, geographical origin (Europe, Africa, Asia, other/unknown), HIV transmission route (heterosexual, homosexual, drug use, other/unknown), prior AIDS diagnosis, cART regimen [two NRTIs plus one ritonavir-boosted PI (other than lopinavir/ritonavir and darunavir/ritonavir), two NRTIs plus lopinavir/ritonavir or darunavir/ritonavir, three NRTIs, other ART combinations (mainly two or three NRTIs plus an unboosted PI or combinations with integrase inhibitors)], period of cART initiation (1998–99, 2000–01, 2002–03, 2004–05, 2006–07, 2008–09, 2010–11) and time between HIV diagnosis and cART start. The slope was additionally adjusted for pretreatment CD4 cell count (per 100 cells/mm³ increase).

For all stratified or subgroup analyses described below we used the linear mixed regression models adjusted for the same covariables described above. Our main hypothesis was related to the potential for differences in pVL that might explain the differences in CD4 cell count evolution between HIV-2+ and HIV-1+ patients. To explore the stability of estimations, we further restricted the analysis to patients with a baseline pVL measured by a test with a limit of \leq 100 copies/mL. We explored whether the effect of the virus type (HIV-2 or HIV-1) on CD4 cell response was modified by the level of baseline pVL by testing homogeneity of the association between HIV type and CD4 cell count evolution across baseline pVL strata (pVL <500 and \geq 500 copies/mL) by integrating an interaction term in the linear mixed model.

We stratified the analyses of immunological response according to the type of cART received, i.e. three NRTIs, two NRTIs combined with a ritonavirboosted PI (other than lopinavir/ritonavir and darunavir/ritonavir), and two NRTIs combined with either lopinavir/ritonavir or darunavir/ritonavir, to assess whether the cART regimen had an effect on the association between HIV type and CD4 cell response by testing homogeneity of the association of HIV type and CD4 cell count evolution across baseline cART regimen. The three categories of cART were chosen because these cART regimens were those recommended by WHO during the study period.¹³

Results are presented for 12 months of follow-up after cART initiation but analyses are based on all available CD4 cell counts after cART initiation up to 115 and 150 months in HIV-2+ and HIV-1+ patients. We described categorical variables with frequencies (%) and continuous variables with medians (IQR). CD4 cell count changes were described with means and 95% CI after having assessed the normality assumption. Categorical variables were compared between HIV-2+ and HIV-1+ (included and excluded) patients using χ^2 tests or Fisher's exact test as appropriate. Quantitative variables were compared between groups using the Wilcoxon-Mann-Whitney test.

All analyses were performed with SAS 9.2 (SAS Institute, Cary, NC, USA).

Results

Characteristics at cART start

In the ACHIeV2e dataset 243 HIV-2+ patients fulfilled inclusion criteria; 58 of these were excluded, giving a total of 185 HIV-2+ patients in the analyses (Figure S1). We observed no significant difference between included and excluded HIV-2+ patients (Table S1). In the COHERE dataset 66483 HIV-1+ patients fulfilled inclusion criteria; 36162 of these were excluded, giving a total of 30231 HIV-1+ patients in the analyses (Figure S1). Included HIV-1+ patients were less often of European origin, were less often injection drug users, and were more often treated with two NRTIs combined with either lopinavir/ritonavir or darunavir/ritonavir compared with non-included patients (Table S1).

Baseline characteristics are presented in Table 1. At start of firstline cART, HIV-2+ patients were significantly older [median age 46 years (IQR 36–52)] compared with HIV-1+ patients [median age 37 years (32–44)] (P < 0.0001). The proportion of HIV-2+ patients with a pVL <500 copies/mL was significantly higher compared with patients infected with HIV-1 (60% versus 12%; P < 0.0001). Median pVL (IQR) was 3.2 loq₁₀ copies/mL (2.2–4.2) and 4.8 loq₁₀ copies/mL (4.0–5.4) in HIV-2+ and HIV-1+ patients, respectively. Median time between HIV diagnosis and cART start was significantly longer in HIV-2+ compared with HIV-1+ patients (1.0 versus 0.6 years; P = 0.0390). Two NRTIs plus lopinavir/ritonavir or darunavir/ritonavir was the most frequent cART regimen in HIV-2+ and HIV-1+ patients (43% and 36.5%, respectively) followed by two NRTIs plus other boosted PIs (23.8% versus 22.6%). Three NRTIs were prescribed in 13% and 7.2% of HIV-2+ and HIV-1+ patients, respectively. Median pretreatment CD4 cell count was similar in HIV-2+ versus HIV-1+ patients [203 cells/mm³ (IQR 100-290) versus 223 cells/mm³ (IQR 100–353); P = 0.1480]. After adjustment for a priori-defined confounding covariables a significant difference in CD4 cell count at cART initiation remained between HIV-2+ and HIV-1+ patients. HIV-2+ patients had 56 CD4 cells/mm³ less at cART initiation compared with HIV-1+ patients (P < 0.0001).

Follow-up data

Median follow-up from first-line cART start until treatment modification, death or last available CD4 cell count was 10 months (IQR 1–27) and 8 months (IQR 2–21) in HIV-2+ and HIV-1+ patients, respectively (P = 0.0412). A median of 3 (IQR 2–8) and 4 (IQR 2–8) CD4 cell counts per HIV-2+ and HIV-1+ patient were available, respectively (P = 0.1555). Among HIV-2+ and HIV-1+ patients, 4.3% and 5.4% were lost to follow-up (P = 0.5017) and 1.1% and 1.7% died (P = 0.5344), respectively.

CD4 cell count evolution

In patients still followed at 12 months, median observed CD4 cell counts at 12 months were 265 cells/mm^3 (IQR 182-420) in

Table 1. Characteristics of patients at initiation of first cART

Characteristic	HIV-1 (<i>N</i> = 30231)	HIV-2 (<i>N</i> = 185)	Р
Age (years), median (IQR)	37 (31–44)	46 (36–52)	< 0.0001
Female, n (%)	8179 (27.1)	88 (47.6)	< 0.0001
Region of origin, n (%)			< 0.0001
Europe	16517 (54.6)	47 (25.4)	
Africa	2495 (8.3)	130 (70.3)	
Asia	413 (1.4)	2 (1.1)	
unknown/other ^a	10806 (35.7)	6 (3.2)	
Transmission risk group, n (%)			< 0.0001
heterosexual	10783 (35.7)	154 (83.2)	
homo/bisexual male	11825 (39.1)	7 (3.8)	
injecting drug user	4887 (16.2)	2 (1.1)	
mother-to-child	26 (0.1)		
unknown/other	2710 (9.0)	22 (11.9)	
Prior AIDS diagnosis, n (%)	7169 (23.7)	42 (22.7)	0.7471
First-line cART regimen, n (%)			0.0002
2 NRTIs + 1 PI/RTV (not LPV or DRV)	6823 (22.6)	44 (23.8)	
2 NRTIS + LPV/RTV or DRV/RTV	11039 (36.5)	79 (42.7)	
3 NRTIs	2180 (7.2)	24 (13)	
other regimens ^b	10189 (33.7)	38 (20.5)	
Period of treatment initiation, <i>n</i> (%)			< 0.0001
1998–99	6292 (20.8)	13 (7.0)	
2000-01	3708 (12.3)	19 (10.3)	
2002-03	3810 (12.6)	32 (173)	
2004–05	4613 (15.3)	34 (18.4)	
2006–07	5557 (18.4)	44 (23.8)	
2008-09	5102 (16.9)	30 (16.2)	
2010-11	1149 (3.8)	13 (7.0)	
Pretreatment HIV RNA viral load			
<500 copies/mL, n (%)	3719 (12.3)	110 (59.5)	< 0.0001
log ₁₀ copies/mL, median (IQR) ^c	4.8 (4.0-5.4)	3.2 (2.2-4.2)	< 0.0001
Pretreatment CD4 cell count (cells/mm³), median (IQR)	223 (100-353)	203 (100-290)	0.1480
Delay between first HIV seropositivity and cART start (years), median (IQR)	0.61 (0.11-3.79)	0.97 (0.21-5.25)	0.0390

DRV, darunavir; LPV, lopinavir; RTV, ritonavir (boost).

^aHIV-1: for 9462 patients the geographical origin was reported as unknown and 1344 were from other regions (Oceania-not Australia, Australia and New Zealand, America, North America, Central and South America and Middle East). HIV-2: for two patients the geographical origin was reported as unknown, two patients were from America and two patients were from Central and South America.

^bOther regimens largely consisted of two or three NRTIs plus an unboosted PI or of combinations with integrase inhibitors [347 (1.1%) HIV-1-infected and 7 HIV-2-infected (3.8%) patients were treated with an integrase inhibitor].

^cFor patients with a viral load below the detection limit of the test, the detection limit has been imputed for the calculation.

HIV-2+ and 404 cells/mm³ (IQR 262–571) in HIV-1+ patients (Figure 1). The mean observed change in CD4 cell count from start of cART to month 12 was +105 (95% CI 77–134) in HIV-2+ and +202 (95% CI 199–205) in HIV-1+ patients, which is an observed difference of 97 CD4 cells/mm³ in CD4 cell increase in 1 year.

After adjusting for pretreatment pVL only, the mean CD4 cell count increase remained significantly lower in HIV-2+ compared with HIV-1+ patients [difference of -41 CD4 cells/mm³/year (95% CI -20 to -61); P < 0.0001]. This difference persisted [difference of -25 CD4 cells/mm³/year (95% CI -44 to -5); P = 0.0127] after adjustment for pretreatment pVL and the other potential confounders (Table 2). All *a priori*-defined covariables were significantly and independently associated with CD4 cell count change. Of note, irrespective of the HIV type and all other variables included

in the model, patients receiving three NRTIs had on average a significantly lower CD4 cell increase when compared with patients receiving a boosted PI-based cART regimen [difference in slope of -34 cells/mm³/year (95% CI -40 to -26); P < 0.0001; Table 2].

All subgroup and stratified analyses showed stable results. When considering only patients with a baseline pVL measured by an assay with a detection limit of \leq 100 copies/mL (HIV-1 n = 27594; HIV-2 n = 129), CD4 cell increase was lower in HIV-2+ patients in adjusted analysis [difference of -29 CD4 cells/mm³/year (95% CI -4 to -53); P = 0.0227]. The effect of HIV type on CD4 cell count response was not modified by baseline pVL; thus, the effect of HIV on CD4 cell count response did not differ in those with a baseline pVL of \geq 500 copies/mL (HIV-1 n = 26602; HIV-2 n = 75) and those with a baseline pVL <500 copies/mL (HIV-1 n = 3719; HIV-2 n = 110)

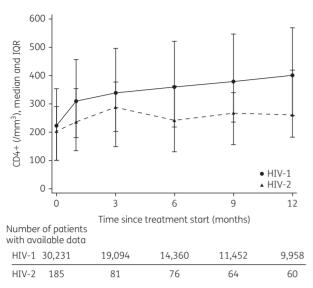


Figure 1. Median observed CD4 cell counts after first-line cART initiation in HIV-2- and HIV-1-infected patients up to 12 months of follow-up. Vertical bars represent the IQR.

(P = 0.1711). Differences in CD4 cell increase between HIV-2+ and HIV-1+ patients were not modified by the initial cART regimen (interaction test: P = 0.9093).

Discussion

The reasons for the previously reported poorer immunological response to first-line cART in HIV-2+ patients are still poorly understood. Using data from two large European observational cohort collaborations, we found a significantly lower CD4 cell increase after starting first-line cART between HIV-2+ and HIV-1+ patients (i.e. observed difference of 97 CD4 cells/mm³). The difference in CD4 cell count increase between HIV-2+ and HIV-1+ patients remained significant after adjusting for pretreatment pVL, cART regimen and other main confounders (i.e. difference of 25 cells/mm³/year). Furthermore, the difference between HIV-2 and HIV-1 was not modified by the cART regimen.

ART regimens have been mainly developed for and validated in HIV-1+ patients; their potency is likely to be different in HIV-2+ patients. Lower potency of several PIs, such as nelfinavir, fosamprenavir and atazanavir, has been reported in phenotypic studies.⁸ The replication cycle of HIV-2 is clearly different from that of HIV-1. HIV-2 pVL is more often below the threshold of detectability, and lower when quantified, compared with HIV-1 pVL, even at advanced disease stages. Total proviral DNA is very similar in HIV-1 and HIV-2 infection after adjustment for CD4 cell count.^{29–32} This suggests at least a blockade of HIV-2 replication at the postintegration level and may explain the differences between HIV-1 and HIV-2 with regard to the potency of PIs. Of note, the difference in CD4 increase between HIV-2 and HIV-1 was smaller (i.e. difference of 25 cells/mm³/year) after adjusting for pretreatment pVL, cART regimen and other main confounders, but remained significant. Furthermore, our results were robust after adjustment for pVL as a time-dependent covariable.

The time between HIV diagnosis and cART initiation, known to contribute to a poorer immunological response to cART, was longer

in HIV-2+ patients compared with HIV-1+ patients. This longer delay is probably due to the slower CD4 cell decrease and the longer asymptomatic stage distinguishing the natural history of HIV-2 infection.³ Nonetheless, CD4 cell increase remained significantly lower in HIV-2+ patients in our analysis adjusted for this delay between HIV diagnosis and cART initiation. For both HIV-1 and HIV-2 infections initiation of cART was recommended at more advanced stages during the study period than currently is the case. Criteria used to initiate ART in HIV-2+ patients were those fitting the national guidelines of each contributing centre effective at the time of cART start. These guidelines were consistent across Europe and included CDC stages B and C, CD4 lymphopenia and HIV-2 RNA levels above the detection limit of the quantification assay; the level of CD4 lymphopenia defined for cART indication in HIV-2 infection was lower than that defined for HIV-1 infection, and may also explain the difference observed in pretreatment CD4 cell counts. Of note, median CD4 cell count was 203 cells/mm³ in HIV-2+ patients at cART start, lower than in HIV-1+ patients, similar to that reported at ART start in the IeDEA-West Africa HIV-2 cohort study (i.e. 166 cells/mm³).³³ It has since been demonstrated that earlier treatment of HIV-1 infection is the best strategy for successful immune restoration.³⁴ Long-term non-progressors account for 6% of the whole asymptomatic HIV-2 population, applying the same methodology as used to determine their proportion in HIV-1-infected patients.³⁵ Among the remaining 94% there are patients with slightly progressive infection, who might benefit from earlier treatment leading to a better immune restoration.

We found an overall significantly lower CD4 cell increase in patients receiving a three-NRTI regimen compared with those receiving a ritonavir-boosted PI-based regimen. These findings obviously confirm the results of a European observational study that included some of the same HIV-2-infected patients,²⁰ but also those of a recent West African study.¹⁹ Direct comparisons of different treatment regimens in observational studies are challenging and the gold standard design for such analyses and for evidence-based conclusions are randomized clinical trials. Such trials are not realizable in Europe due to the restricted number of potentially eligible HIV-2-infected patients living there. However, we report for the first time a comparison of CD4 cell response to firstline cART in a large population of HIV-2- and HIV-1-infected patients under standardized routine follow-up in Europe, allowing us to adjust the comparison for initial pVL levels, cART regimen, region of origin and other patient characteristics, such as pretreatment CD4 cell count and previous AIDS diagnosis, and for calendar periods of cART initiation.

Chronic immune activation and inflammation markers have been linked to disease progression in HIV-1+ 36,37 and HIV-2+ patients.³⁸⁻⁴⁰ Differences in CD4 cell count response to first cART may be linked to differences in the underlying pathogenicity of these two viruses regarding immune activation. However, chronic immune activation is directly linked to pVL^{39,41} and in our comparison of immune response we controlled for pVL in various ways. Of note, it has been shown that HIV-2 non-progression is associated with better immune response to the virus, less immune activation and broad neutralizing antibody response; nevertheless, we do not know what came first: the particularities of HIV-2 replication or the immune response.

There are several limitations to our analyses. Differences in immunological outcome between HIV-1 and HIV-2 may have

Table 2. Estimated mean CD4 cell count differences at first-line cART start and CD4 cell count changes adjusted for all listed covariables

	CD4 cell count at initiation of first cART (cells/mm ³)			CD4 cell count change (cells/mm ³ /year)		
	mean	95% CI	Р	mean	95% CI	Р
Intercept/slope	533	518-549	< 0.0001	154	142-166	0.0003
HIV type						
HIV-2 versus HIV-1	-56	-86 to -26	< 0.0001	-25	−44 to −5	0.0127
Pretreatment pVL (per additional 1 log ₁₀ copies/mL)	-26	-28 to -24	< 0.0001	9	8-10	< 0.0001
Age at treatment initiation (per additional 10 years)	-22	−25 to −20	< 0.0001	-9	−10 to −7	< 0.0001
Sex						
female versus male	26	19-32	< 0.0001	10	6-15	< 0.0001
Geographical origin			< 0.0001			< 0.0001
Europe (reference category)						
Africa	-31	-40 to -22		-25	-32 to 19	
Asia	-53	−73 to −33		-7	-21 to 6	
unknown/other	-18	−23 to −13		-4	−8 to −1	
Transmission group			< 0.0001			< 0.0001
heterosexual (reference category)						
homo/bisexual male	53	47-59		9	4-13	
injecting drug user	-25	−33 to −18		-16	−21 to −10	
unknown/other	-9	-17 to 0		-12	−18 to −6	
Prior AIDS diagnosis						
yes versus no	-126	−132 to −121	< 0.0001	6	2-10	0.0047
cART regimen			< 0.0001			< 0.0001
2 NRTIs + 1 PI/RTV (reference category)						
2 NRTIS + LPV/RTV or DRV/RTV	-9	−15 to −3		4	-1 to 8	
3 NRTIS	53	42-63		-34	-40 to -26	
other regimens ^a	24	17-32		-5	-11 to 0.5	
Period of cART initiation			< 0.0001	-		< 0.0001
1998–99	-7	-16 to 2		-53	-60 to -46	
2000-01	-26	-36 to -17		-59	-66 to -52	
2002-03	-14	-23 to -6		-56	-62 to -49	
2004-05	-17	-25 to -9		-50	-57 to -44	
2006-07	3	-4 to 11		-38	-44 to -32	
2008–09 (reference category)	5			50	1100 52	
2010-11	-11	-24 to 3		130	108 to 153	
Delay between first HIV seropositivity and cART start (per additional year)	-0.8	-1.4 to -0.3	0.0039	-1.1	-1.5 to -0.7	< 0.0001
Pretreatment CD4 cell count (per additional 100 cells/mm ³) ^b	-	-	-	-2	-3 to -1	< 0.0001

DRV, darunavir; LPV, lopinavir; RTV, ritonavir (boost).

For CD4 cell count changes, a negative value indicates a lower CD4 increase and a positive value indicates a higher CD4 increase. N = 30231 for HIV-1-infected patients and N = 185 for HIV-2-infected patients.

^aOther regimens largely consisted of two or three NRTIs plus an unboosted PI or of combinations with integrase inhibitors.

^bThe reported difference in CD4 cell count change is for a difference of 100 CD4 cells/mm³ at baseline, i.e. in a patient with 400 CD4 cells/mm³ the increase is of 2 cells less than compared with a patient with 300 CD4 cells/mm³.

been observed due to unmeasured confounding factors. We cannot completely rule out a selection bias as we did not analyse pretreatment slopes of CD4 cell decreases in HIV-1 and HIV-2 populations. We were unable to adjust for hepatitis B and/or for hepatitis C co-infection, although we adjusted for geographical origin and transmission risk group, which are closely linked to hepatitis B and C seroprevalences. Furthermore, follow-up was quite short and may hamper the extrapolation of our results to the long term.

Our study has several strengths. This is a large study comparing CD4 cell dynamics between HIV-2+ and HIV-1+ patients after start of first-line cART. The data quality was assured through two large European collaborative networks adhering to strict quality control checks, which allowed us to adjust our comparative analysis for the major confounding variables. Furthermore, we could confirm our main findings in subgroup and stratified analyses showing the robustness of the results and the absence of effect modification by initial pVL and cART regimen.

Differences in CD4 cell dynamics between HIV-2 and HIV-1 were consistent in all analyses, with a poorer CD4 cell increase after start of treatment in HIV-2+ patients, even after adjustment for pVL. Our results underline the need to identify other factors contributing to this lower CD4 cell response, such as more potent drugs against HIV-2, adapted to the particularities of the virus replication when compared with HIV-1, in order to improve case management.

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Supplementary data

Figure S1 and Table S1 are available as Supplementary data at JAC Online.

References

1 Campbell-Yesufu OT, Gandhi RT. Update on human immunodeficiency virus (HIV)-2 infection. *Clin Infect Dis* 2011; **52**: 780–7.

2 Andersson S, Norrgren H, da Silva Z *et al.* Plasma viral load in HIV-1 and HIV-2 singly and dually infected individuals in Guinea-Bissau, West Africa: significantly lower plasma virus set point in HIV-2 infection than in HIV-1 infection. *Arch Intern Med* 2000; **160**: 3286–93.

ΙΔ

3 Drylewicz J, Matheron S, Lazaro E *et al.* Comparison of viro-immunological marker changes between HIV-1 and HIV-2-infected patients in France. *AIDS* 2008; **22**: 457–68.

4 Hansmann A, Schim van der Loeff MF, Kaye S *et al.* Baseline plasma viral load and CD4 cell percentage predict survival in HIV-1- and HIV-2-infected women in a community-based cohort in The Gambia. *J Acquir Immune Defic Syndr* 2005; **38**: 335–41.

5 Marlink R, Kanki P, Thior I *et al*. Reduced rate of disease development after HIV-2 infection as compared to HIV-1. *Science* 1994; **265**: 1587–90.

6 Popper SJ, Sarr AD, Travers KU *et al*. Lower human immunodeficiency virus (HIV) type 2 viral load reflects the difference in pathogenicity of HIV-1 and HIV-2. *J Infect Dis* 1999; **180**: 1116–21.

7 Whittle H, Morris J, Todd J *et al.* HIV-2-infected patients survive longer than HIV-1-infected patients. *AIDS* 1994; **8**: 1617–20.

8 Desbois D, Roquebert B, Peytavin G *et al*. In vitro phenotypic susceptibility of human immunodeficiency virus type 2 clinical isolates to protease inhibitors. *Antimicrob Agents Chemother* 2008; **52**: 1545–8.

9 Poveda E, Rodes B, Toro C *et al*. Are fusion inhibitors active against all HIV variants? *AIDS Res Hum Retroviruses* 2004; **20**: 347–8.

10 Tuaillon E, Gueudin M, Lemee V *et al*. Phenotypic susceptibility to nonnucleoside inhibitors of virion-associated reverse transcriptase from different HIV types and groups. *J Acquir Immune Defic Syndr* 2004; **37**: 1543–9.

11 Witvrouw M, Pannecouque C, Switzer WM *et al*. Susceptibility of HIV-2, SIV and SHIV to various anti-HIV-1 compounds: implications for treatment and postexposure prophylaxis. *Antivir Ther* 2004; **9**: 57–65.

12 Ekouevi DK, Tchounga BK, Coffie PA *et al.* Antiretroviral therapy response among HIV-2 infected patients: a systematic review. *BMC Infect Dis* 2014; **14**: 461.

13 WHO. Consolidated Guidelines on the Use of Antiretroviral Drugs for Treating and Preventing HIV Infection. 2013; 272. Geneva: WHO Press.

14 Prise en charge médicale des personnes vivant avec le VIH— Recommandations du groupe d'experts. Sous la direction du Pr Philippe Morlat et sous l'égide du CNS et de l'ANRS 2017. http://cns.sante.fr/actualites/priseen-charge-du-vih-recommandations-du-groupe-dexperts/.

15 Gilleece Y, Chadwick DR, Breuer J *et al.* British HIV Association guidelines for antiretroviral treatment of HIV-2-positive individuals 2010. *HIV Med* 2010; **11**: 611–9.

16 Adje-Toure CA, Cheingsong R, Garcia-Lerma JG *et al.* Antiretroviral therapy in HIV-2-infected patients: changes in plasma viral load, CD4+ cell counts, and drug resistance profiles of patients treated in Abidjan, Cote d'Ivoire. *AIDS* 2003; **17** Suppl 3: S49–54.

17 Benard A, Damond F, Campa P *et al.* Good response to lopinavir/ritonavircontaining antiretroviral regimens in antiretroviral-naive HIV-2-infected patients. *AIDS* 2009; **23**: 1171–3.

18 Matheron S, Damond F, Benard A *et al.* CD4 cell recovery in treated HIV-2-infected adults is lower than expected: results from the French ANRS CO5 HIV-2 cohort. *AIDS* 2006; **20**: 459–62.

19 Balestre E, Ekouevi DK, Tchounga B *et al.* Immunologic response in treatment-naive HIV-2-infected patients: the IeDEA West Africa cohort. *J Int AIDS Soc* 2016; **19**: 20044.

20 Benard A, van Sighem A, Taieb A *et al.* Immunovirological response to triple nucleotide reverse-transcriptase inhibitors and ritonavir-boosted protease inhibitors in treatment-naive HIV-2-infected patients: the $ACHI_EV_{2E}$ Collaboration Study Group. *Clin Infect Dis* 2011; **52**: 1257–66.

21 Gottlieb GS, Sow PS, Hawes SE *et al*. Equal plasma viral loads predict a similar rate of CD4+ T cell decline in human immunodeficiency virus (HIV)

type 1- and HIV-2-infected individuals from Senegal, West Africa. *J Infect Dis* 2002; **185**: 905–14.

22 Drylewicz J, Eholie S, Maiga M *et al.* First-year lymphocyte T CD4+ response to antiretroviral therapy according to the HIV type in the IeDEA West Africa collaboration. *AIDS* 2010; **24**: 1043–50.

23 Chene G, Phillips A, Costagliola D *et al.* Cohort profile: Collaboration of Observational HIV Epidemiological Research Europe (COHERE) in EuroCoord. *Int J Epidemiol* 2016; doi:10.1093/ije/dyw211.

 ${\bf 24}\,$ Damond F, Benard A, Balotta C et al. An international collaboration to standardize HIV-2 viral load assays: results from the 2009 ACHI_EV_{2E} quality control study. J Clin Microbiol 2011; ${\bf 49}$: 3491–7.

25 Damond F, Benard A, Ruelle J *et al.* Quality control assessment of human immunodeficiency virus type 2 (HIV-2) viral load quantification assays: results from an international collaboration on HIV-2 infection in 2006. *J Clin Microbiol* 2008; **46**: 2088–91.

26 Kjaer J, Ledergerber B. HIV cohort collaborations: proposal for harmonization of data exchange. *Antivir Ther* 2004; **9**: 631–3.

27 Thiebaut R, Walker S. When it is better to estimate a slope with only one point. *QJM* 2008; **101**: 821-4.

28 Jacqmin-Gadda H, Sibillot S, Proust C *et al*. Robustness of the linear mixed model to misspecified error distribution. *Comput Stat Data Anal* 2007; **51**: 5142–54.

29 Foxall RB, Cortesao CS, Albuquerque AS *et al.* Gag-specific CD4+ T-cell frequency is inversely correlated with proviral load and directly correlated with immune activation in infection with human immunodeficiency virus type 2 (HIV-2) but not HIV-1. *J Virol* 2008; **82**: 9795–9.

30 Matheron S, Pueyo S, Damond F *et al*. Factors associated with clinical progression in HIV-2 infected-patients: the French ANRS cohort. *AIDS* 2003; **17**: 2593–601.

31 Popper SJ, Sarr AD, Gueye-Ndiaye A *et al*. Low plasma human immunodeficiency virus type 2 viral load is independent of proviral load: low virus production in vivo. *J Virol* 2000; **74**: 1554–7. **32** Soares RS, Tendeiro R, Foxall RB *et al.* Cell-associated viral burden provides evidence of ongoing viral replication in aviremic HIV-2-infected patients. *J Virol* 2011; **85**: 2429–38.

33 Ekouevi DK, Balestre E, Coffie PA *et al.* Characteristics of HIV-2 and HIV-1/ HIV-2 dually seropositive adults in West Africa presenting for care and antiretroviral therapy: the IeDEA-West Africa HIV-2 Cohort Study. *PLoS One* 2013; **8**: e66135.

34 INSIGHT Strategic Timing of AntiRetroviral Treatment (START) Study Group. Why START? Reflections that led to the conduct of this large longterm strategic HIV trial. *HIV Med* 2015; **16** Suppl 1: 1–9.

35 Thiebaut R, Matheron S, Taieb A *et al*. Long-term nonprogressors and elite controllers in the ANRS CO5 HIV-2 cohort. *AIDS* 2011; **25**: 865–7.

36 Deeks SG, Kitchen CM, Liu L *et al.* Immune activation set point during early HIV infection predicts subsequent CD4+ T-cell changes independent of viral load. *Blood* 2004; **104**: 942–7.

37 Giorgi JV, Hultin LE, McKeating JA *et al.* Shorter survival in advanced human immunodeficiency virus type 1 infection is more closely associated with T lymphocyte activation than with plasma virus burden or virus chemokine coreceptor usage. *J Infect Dis* 1999; **179**: 859–70.

38 Honge BL, Andersen MN, Jespersen S *et al.* Macrophage activation in HIV-2-infected patients is less affected by antiretroviral treatment-sCD163 in HIV-1, HIV-2, and HIV-1/2 dually infected patients. *J Acquir Immune Defic Syndr* 2016; **72**: 254–8.

39 Leligdowicz A, Feldmann J, Jaye A *et al*. Direct relationship between virus load and systemic immune activation in HIV-2 infection. *J Infect Dis* 2010; **201**: 114–22.

40 Thiebaut R, Charpentier C, Damond F *et al.* Association of soluble CD14 and inflammatory biomarkers with HIV-2 disease progression. *Clin Infect Dis* 2012; **55**: 1417–25.

41 Mavigner M, Delobel P, Cazabat M *et al.* HIV-1 residual viremia correlates with persistent T-cell activation in poor immunological responders to combination antiretroviral therapy. *PLoS One* 2009; **4**: e7658.