

Molecular Characterization of *Klebsiella Pneumonia* in Clinical Isolates with High Resistance toward Carbapenemases

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Carbapenem-resistant *Enterobacteriaceae* (CRE) is spreading in worldwide and has become endemic in several countries, including Italy [1,2]. The global dissemination of carbapenem-resistant *Enterobacteriaceae* is mostly due to strains of *Klebsiella pneumoniae* and less frequently of other species producing various types of acquired carbapenemases [3]. However, antimicrobials (such as carbapenems, cephalosporin) which are widely used in the treatment of *K. pneumoniae* infection have been proven to be a major factor to mortality and morbidity in settings with limited diagnostic facilities and treatment options [4], especially in increasing emergent multidrug resistant (MDR) *K. pneumoniae*. MDR *K. pneumoniae*, internationally defined as having non-susceptibility to at least three classes of antibiotics or more [5], poses tremendous threat to clinical setups in hospitals. Resistance to carbapenem by such enzyme is a global concern due to limited therapeutic options and their association with life-threatening infections. Large referral hospitals and teaching institutions are at great risk for a widespread outbreak of infections and responsible for the spread of such strains from one location to another and to other hospitals. Experiences in individual hospitals or in whole countries they have shown how it is possible to eradicate or contain strongly the spread of *Enterobacteria* strains producing carbapenemases through aggressive infection control interventions in the health sector, aimed at promptly identifying cases of clinically evident infections and colonized patients and promptly adopting stringent measures of containment of dissemination (isolation of colonized or infected patients, hand hygiene, cleaning and environmental contamination, etc.) [6].

In this report we present the phenotypic and molecular characterizations of KPC-producing carbapenem-resistant *K. pneumoniae* clinical isolates in an Italian hospital.

The study included thirty-eight isolates obtained from the following samples: rectal swab (n=14), urine (n=10), bronchoalveolar lavage (BAL; n=8) and wound swab (n=6).

Species identification and antimicrobial susceptibility testing were carried out using the Vitek 2 system and the ASTGN201 cards (bio Merieux, Florence, Italy).

Carbapenemase production was confirmed by a disc diffusion synergy test including meropenem and two carbapenemase-inhibiting compounds (ethylenediaminetetraacetic acid EDTA, and boronic acid) [7].

PCR screening was performed with cell lysates for identification of the carbapenemase genes using specific bla primers designed for identifying known β -lactamase genes including, *bla*_{KPC}; *bla*_{IMP}, *bla*_{VIM}, *bla*_{OXA48}, *bla*_{NDM} [8,9] cellular debris were removed by centrifugation at 17,310 g for 5 min. aliquot of 2 μ l of the supernatant used as DNA template for PCR [10]. PCR-positive amplicons were purified by the QIA quick PCR Purification Kit according to the manufacturer's instructions (Qiagen, Milan, Italy) and directly sequenced using amplification primers on the 3130 Genetic Analyzer (Applied Biosystem, Milan, Italy). Purification and sequencing were

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carried out by Genexs.r.o. (CZ, Czech Republic). Sequence alignment and analysis were performed online using the BLAST program of the National Center for Biotechnology Information (www.ncbi.nlm.nih.gov).

The *K. pneumoniae* isolates underwent molecular typing by means of the MLST technique were performed using seven conserved housekeeping genes (*gapA*, *infB*, *mdh*, *pgi*, *phoE*, *rpoB*, *tonB*) [11]. A detailed protocol of the MLST procedure, including allelic type and sequence type (ST) assignment methods, is available in MLST databases from Pasteur Institute, Paris, France at the following website (<http://www.pasteur.fr/recherche/genopole/PF8/mlst/Kpneumoniae.html>).

MICs of imipenem and meropenem in the range of 4 mg/L to ≥ 16 mg/L and 1 mg/L to ≥ 16 mg/L, respectively, were observed. The tested strains were analyzed for the presence of resistance mechanisms against β -lactam antibiotics using PCR amplifications for bla genes responsible for carbapenemases production (*bla*_{KPC}, *bla*_{VIM}, *bla*_{IMP}, *bla*_{NDM} and *bla*_{OXA-48}).

Overall, 38 out of the 34 isolates were positive for the *bla*_{KPC} gene, 2 for the *bla*_{VIM} metallo- β -lactamase and 2 for the *bla*_{OXA-48} gene; while all were negative for NDM gene. Sequence analysis of the entire genes revealed perfect identity with the *bla*_{KPC-2} (20/38), *bla*_{KPC-3} (18/38), *bla*_{VIM-1} (2/18) and *bla*_{OXA-48} (2/38) genes.

Multi Locus Sequence Typing showed that the ST258 clone was typical of the KPC-2 producing strains, while the ST512 clone was characteristic of the 5 *bla*_{KPC-3} producing strains, were previously reported in Italy [12,13].

Recognition of different carbapenem-non-susceptible clones by molecular epidemiological tools is an important step towards tracing transmission routes, developing targeted control and prevention strategies, and monitoring their effectiveness.

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In addition, the KPC and IMP genes are usually associated with mobile genetic elements that can efficiently disseminate to other bacteria in the hospital environment [14,15]. It is therefore important to emphasize the prompt recognition and establishment of proper therapeutic and infection control measures in order to reduce the spread of infections caused by these highly resistant organism.

The accurate detection of this emerging public health threat clone is crucial both for the selection of appropriate therapeutic regimens and for controlling the spread of KPC-type enzymes.

Competing Interests

The authors declare that they have no competing interests.

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