# HEALTH AND NUTRITIONAL BIOMARKERS IN HONEYBEES: OPPORTUNITIES AND CHALLENGES UNDER FIELD CONDITIONS

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### Introduction

The decline of honeybee (Apis *mellifera*) populations has negative consequences not only for agriculture and beekeeping, but also for ecosystems. In human and veterinary medicine, proteomics and metabolomics provide valuable biomarkers to assess the health and nutritional status of organisms. In honeybees, the application of these techniques is still in its infancy and remains underexplored from a clinical perspective.

This study aims to investigate the most abundant proteins of honeybee hemolymph as potential biomarkers of health and nutritional status at the colony level. In addition, an untargeted metabolomics-based approach was applied to honeybee extracts.

### **Materials and Methods**

Samples of hemolymph were collected from honeybees in different apiaries in the province of Bologna in different periods of the year, from April to November. Hemolymph proteins were separated and quantified by 1D SDS-PAGE, as reported by Isani et al. (2023). cytosolic extracts Honeybee were fractionated using size exclusion chromatography (SEC) and metabolites were analyzed in fractions using mass spectrometry (Orbitrap Exploris 480, Thermo Fisher). Metabolites were detected by Compound Discoverer 3.3 software (Thermo Fisher). Different statistical tests were used

depending on whether sample distribution was normal or not.

Differences between honeybee subcastes or seasons were considered statistically significant for p<0.05.

### Results

The five most abundant hemolymph proteins, namely vitellogenin, apolipophorin I and II, transferrin and hexamerin 70a, represent a panel of biomarkers related to key metabolic processes. These proteins show interesting variations depending on many physiological and environmental factors, such as:

- **the age of honeybees:** nurse bees had the highest vitellogenin concentration compared to the other two sub-caste (Figure 1);
- **the season:** in autumn, a peak of vitellogenin and transferrin concentration was observed in winter bees;
- the presence of parasites: a decrease of vitellogenin, apolipophorin II, transferrin, and hexamerin 70a was detected in bees parasitized by Varroa.

After SEC fractionation of cytosolic extracts of honeybees (Figure 3) and mass analysis, one hundred and ninety-eight different pathways and more than 2000 metabolites were identified. The most abundant metabolites belonged to the flavone pathway (Table 1), followed by the lipoxygenase pathway. Most metabolites are of plant origin and may be related to the environmental availability of nectar and pollen, which in turn are essential for honeybee nutrition, suggesting a possible role as biomarkers of nutritional status.

## Proteomics



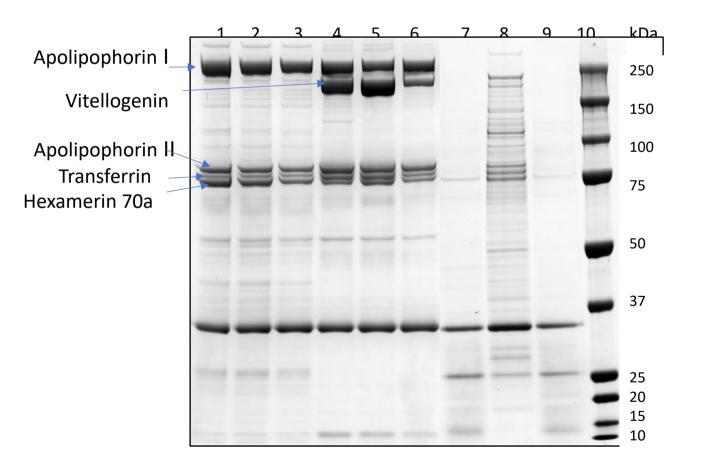


Figure 1. Representative SDS-PAGE of honeybee hemolymph: lanes 1, 2, represent the and 3 profiles of protein newborn bees; lanes 4, 5, and 6 represent the protein profiles of nurse bees; lanes 7, 8, and9 represent protein profiles of foraging bees; lane 10 represents the molecular mass marker.

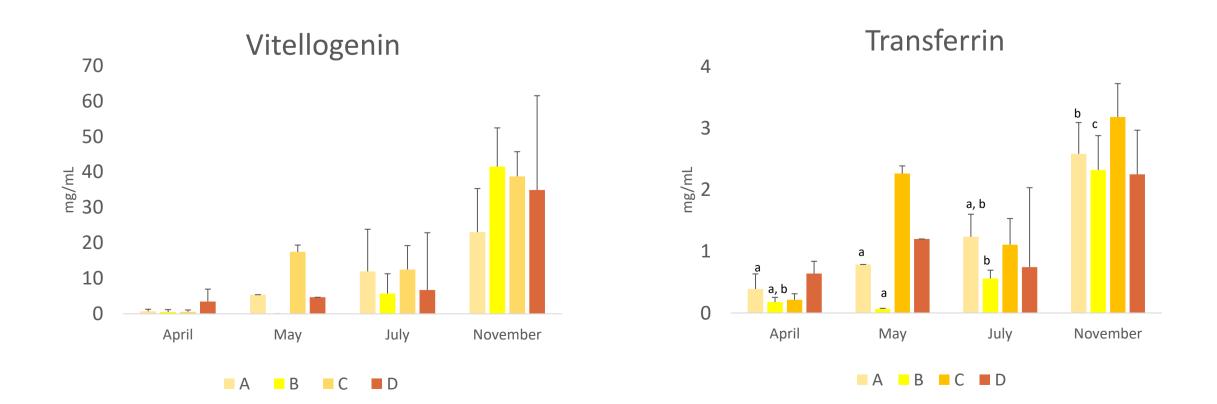
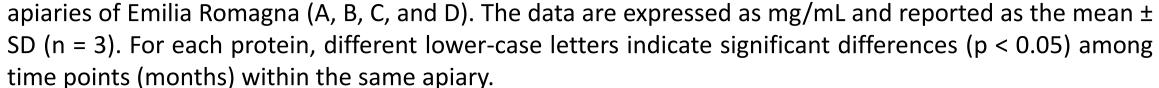


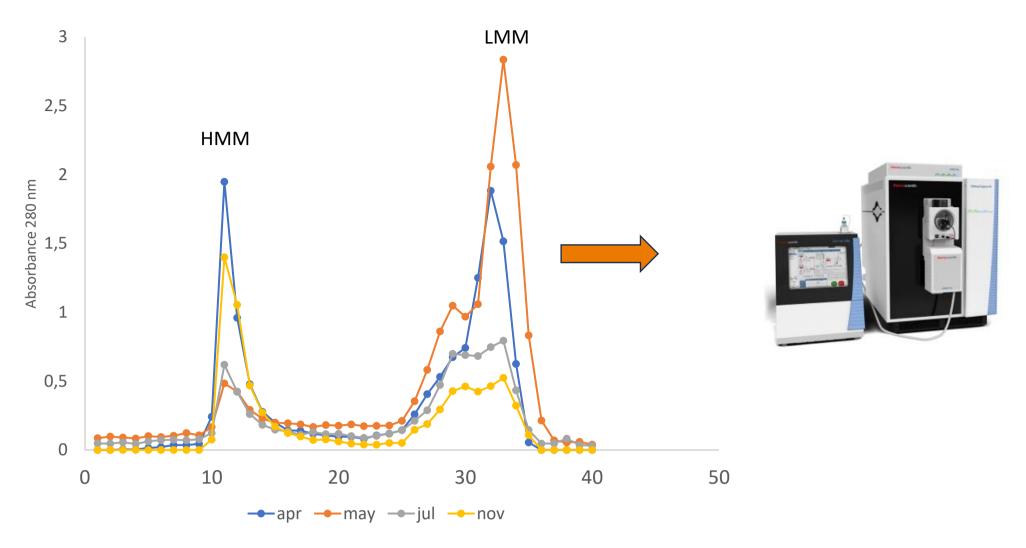
Figure 2. Concentrations of hemolymph vitellogenin and transferrin in different periods of the year in 4



# Metabolomics



**#8** 



**Figure 3**. Chromatographic pattern after size exclusion chromatography (SEC) of extracts of honeybees sampled from the same apiary in different periods of the year. Proteins (HMM), peptides and low molecular mass molecules (LMM) were measured at 280 nm.

Superpathway of flavones and derivatives biosynthesis	#1	#2	#3	#4	#5	#6	#7	7
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	Apr	May	Jul	Nov	Apr	May	Jul	Nov
	Site1	Site1	Site1	Site1	Site2	Site2	Site2	Site2
Kaempferol-3-glucoside-7-rhamnoside	5.8	4.7	2.7	3.6	5.8	3.9	2.7	
Kaempferol 3-O-rhamnoside-7-O-glucoside								
Quercetin 3-O-rhamnoside-7-O-glucoside	6.2	4.3	6.2	3.6	5.5	2.0	3.7	
Kaempferol 3,7-O-diglucoside								
Kaempferol 3-O-β-D-glucosyl-(1->2)-glucoside								
Kaempferol-3-gentiobioside								
Rutin								
Quercetin 3,7-O-diglucoside	3.0	1.7	5.1	6.6	3.2	5.8	4.7	
Quercetin 3-O-sophoroside								
Quercetin 3,5-O-diglucoside								
Quercetin-3-gentiobioside								
Kaempferol-3-rhamnoside-7-rhamnoside		5.2	2.6		6.2	5.5	4.8	4.5
(+)-Dihydromyricetin	3.9	5.8	3.5	2.7	5.1	5.5		
Kaempferol 3-O-rhamnosyl(1->2)glucoside-7-O-rhamnoside	5.8	2.4	3.1	6.2	6.6	2.1	5.1	
Kaempferol-3-O-gentiobioside-7-O-rhamnoside		_	_				_	
Kaempferol 3-O- $\beta$ -D-glucosyl-(1->2)-glucosyl-(1->2)- $\beta$ -D-glucoside		6.2	5.5	5.2	4.7	5.5	6.6	
(+)-Taxifolin	3.5			5.8	3.1	5.4	6.2	5.2
Kaempfenol-3-rhamnoside		1.8	5.0	5.6		5.9	5.8	6.1

**Table 1.** Major metabolites ofthe flavone super-pathwaydetected in LMM fractions ofcytosolic extracts fromhoneybees sampled fromhives of two apiaries in theprovince of Bologna (Site 1and Site 2).

Colored boxes: peak amount of each compound identified. Grey boxes: absence of detection.

### CONCLUSIONS

These are the first data on the concentration of a panel of hemolymph proteins in different phases of the life cycle and seasons in Italian honeybees. They can be considered as a starting point to establish in the future physiological intervals useful in clinical practice to evaluate the health and nutritional status of honeybees. In addition, many of the metabolites identified in honeybees are of plant origin and can also help to assess the availability of environmental resources in different seasons and locations.

**REFERENCES** - Isani G, Bellei E, Rudelli C, Cabbri R, Ferlizza E, Andreani G. SDS-PAGE-Based Quantitative Assay of Hemolymph Proteins in Honeybees: Progress and Prospects for Field Application. Int J Mol Sci. 2023 Jun 16;24(12):10216. doi: 10.3390/ijms241210216. PMID: 37373362; PMCID: PMC10299212.

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