

Para-Cresol and the Brain: Emerging Role in Neurodevelopmental and Neurodegenerative Disorders and Therapeutic Perspectives

Laura Bertarini, Federico Imbeni, Virginia Brighenti, Isabella Martusciello, Federica Pellati,* and Silvia Alboni*



Cite This: *ACS Pharmacol. Transl. Sci.* 2025, 8, 3432–3452



Read Online

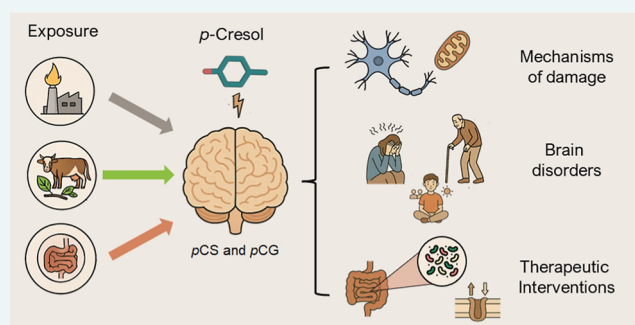
ACCESS |

Metrics & More

Article Recommendations

ABSTRACT: *p*-Cresol (*p*C) is a phenolic compound to which humans can be exposed through both environmental sources, such as a pollutant, and endogenous production by the gut microbiota. Among microbial contributors, *Clostridioides difficile* appears to be a major source of *p*C within the body. Once absorbed, *p*C is highly protein-bound in plasma and predominantly circulates in its hepatic conjugated forms: *p*-cresyl sulfate (*p*CS) and *p*-cresol glucuronide (*p*CG), which are mainly excreted in urine. Accumulation of these metabolites, particularly *p*CS, classified as a protein-bound uremic toxin, has been associated with the progression of chronic kidney disease (CKD) and related complications, due to its pro-oxidant, pro-inflammatory, and pro-apoptotic properties. CKD patients are at increased risk for cognitive impairment, affective disorders, and central nervous system (CNS) dysfunctions. In recent years, increasing evidence has suggested a potential role of *p*C and its metabolites in CNS diseases. Here, we summarize current knowledge on the involvement of these compounds in the pathogenesis and progression of autism spectrum disorder, Parkinson's disease, Alzheimer's disease, and post-traumatic stress disorder. We also discuss how modulating systemic levels of *p*C may represent a promising strategy to improve pathological phenotypes in the context of neurodevelopmental and neurodegenerative disorders.

KEYWORDS: *p*-cresol, autism, Alzheimer's disease, Parkinson's disease, stress, brain



p-Cresol (4-methylphenol, *p*C) is a partially lipophilic compound (log *P* 1.94) with low-molecular-weight, belonging to the large family of phenols.¹ The interest in the role of this compound related to the etiopathogenesis of central diseases, including those associated with neurodevelopment and neurodegeneration, has steadily increased in recent years. Comprehensive data on the general toxicology of *p*C, including its physicochemical properties, environmental distribution, uses, exposure routes, metabolism, and potential health effects, are available from several authoritative sources.^{2,3}

The present review aims to provide comprehensive information on *p*C, its presence and origin in the human body, its pharmacokinetic properties, and its linkage with the pathogenesis of serious central nervous system (CNS) diseases, with regard to autism (ASD), Alzheimer's Disease (AD), Parkinson's Disease (PD), and Post Traumatic Stress Disorder (PTSD). Neurophysiological and neuropathological conditions involving *p*C are listed in Table 1, which will be thoroughly discussed in this review.

■ P-CRESOL EXPOSURE AND DISPOSITION: INSIGHTS INTO ITS ABSORPTION, DISTRIBUTION, METABOLISM, AND ELIMINATION

Given the complex pathways through which *p*C is absorbed, metabolized, distributed, and eliminated, Figure 1 provides a comprehensive overview of its ADME profile and related metabolites, including the main sources of exposure. This facilitates understanding of the multiple processes discussed throughout this section and emphasizes the predominant role of microbial production, first-pass metabolism, distribution, and primary routes of excretion.

*p*C is widely distributed in the environment, originating from both natural and anthropogenic sources. Natural sources include microbial metabolism and plant material degradation,

Received: April 18, 2025

Revised: August 22, 2025

Accepted: August 28, 2025

Published: September 15, 2025



Table 1. Representative Neurophysiological and Neuropathological Conditions for Which an Involvement of the *p*-Cresol and its related metabolites Has Been Suggested

conditions	species	refs
Neurodevelopmental Disease		
Autism Spectrum Disorder (ASD)	human	111
ASD	human	113
ASD	human	123
ASD	human	120
ASD	rat	124
ASD	human	121
ASD	human	95
ASD	mouse	96
ASD	mouse	62
ASD	mouse	60
ASD	human	116
ASD	mouse	94
ASD	human	63
ASD	human	114
Neurodegenerative Disease		
Parkinson's Disease (PD)	human	154
PD	human	161
PD	human	151
PD	human	149
PD	human	127
PD	human	140
PD	human	150
multiple sclerosis	human	93
Alzheimer's Disease (AD)	rat	180
AD	mouse	62
AD	human	187
AD	human	188
Neuropsychiatric Disorder		
Post-Traumatic Stress Disorder (PTSD)	mouse	94
PTSD	mouse	57

whereas anthropogenic contributions stem primarily from industrial activities, such as petrochemical processing, waste incineration, and vehicle emissions (Figure 1).^{2–4} Despite this environmental presence, evidence from germ-free animal models and antibiotic treatment studies indicates that the gut microbiota (GMB) is the primary endogenous source of *p*C.^{5,6} In the gut, phenolic compounds like *p*C are mainly produced through microbial metabolism of L-tyrosine and, to a lesser extent, L-phenylalanine.⁷ At least 55 bacterial species have been identified as *p*C producers, with *Blautia hydrogenotrophica*, *Clostridioides difficile*, *Olsenella uli*, and *Romboutsia ilealis* recognized as major contributors.^{4,8} Supporting the role of the GMB in *p*C production, antimicrobial treatments such as neomycin and vancomycin have been shown to suppress its endogenous synthesis.⁵ Furthermore, metabolomic studies provide additional evidence for the pivotal role of GMB in *p*C production, demonstrating that variations in microbial composition significantly affect systemic levels.⁶

Gut *p*C production is strongly influenced by dietary and physiological factors: high-protein diets promote the growth of proteolytic bacteria, leading to increased *p*C levels,⁹ while fiber-rich diets and probiotic supplementation reduce systemic concentrations by reshaping microbial communities.^{10–13} Fasting has also been shown to elevate *p*C levels in rats, likely due to reduced fiber intake, slower intestinal transit, and greater availability of endogenous proteins, which favor

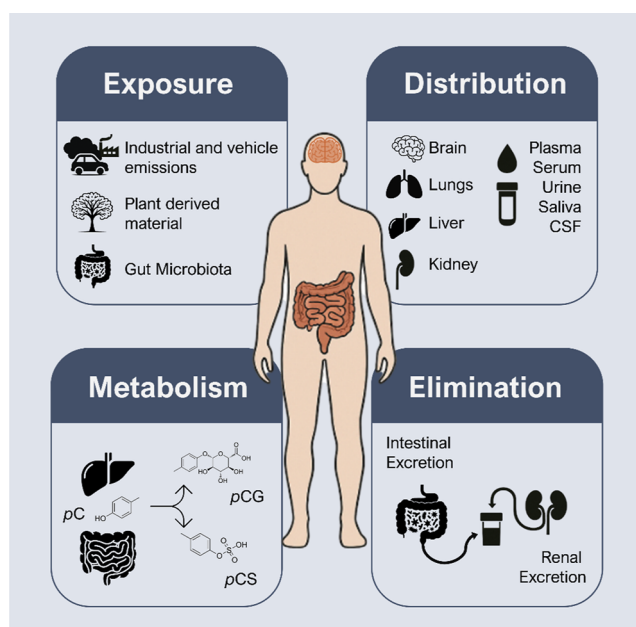


Figure 1. Absorption, distribution, metabolism, and excretion (ADME) of *p*C and its related metabolites. *p*C exposure occurs via inhalation, ingestion, dermal contact, and, mainly, microbial production in the gut. In the body, *p*C is absorbed in the intestine and primarily metabolized into *p*-cresyl sulfate (*p*CS) and glucuronide (*p*CG), with *p*CS being predominant, especially in uremic conditions. Distribution varies by tissues, with *p*CS prevailing in blood, kidneys, and lungs, and free *p*C enriched in the liver. Renal excretion is the main elimination route, with smaller contributions from feces and the lungs. The figure was created using icons from the Mind the Graph platform.

proteolytic activity and *p*C absorption.¹⁴ Additional factors, such as L-tyrosine intake and gastric acid suppression therapy, further influence *p*C production.^{12,13,15,16} Moreover, carbon source availability is a key regulator of *p*C production in the gut, with glucose promoting its synthesis and lactic acid inhibiting it.¹⁷

*p*C is produced through the fermentation of L-tyrosine via the intermediate *p*-hydroxyphenylacetate (*p*HPA), which is then decarboxylated by the glycyl radical enzyme 4-hydroxyphenylacetate decarboxylase (HpdBCA).¹⁸ The detection of *p*HPA in human colon at concentrations up to 19 μM ¹⁹ supports its physiological relevance as a precursor for in vivo *p*C synthesis. Functional studies have demonstrated that disruption of any gene encoding the HpdBCA complex abolishes *p*C production in *C. difficile*.^{20,21}

Following its biosynthesis, *p*C exerts antimicrobial effects against Gram-negative bacteria both *in vitro* and in *in vivo* infection models, with *C. difficile* showing a markedly higher tolerance to *p*C than other common gut microbes, such as *Gammaproteobacteria* and *Bacteroidetes*, thus suggesting a competitive advantage within the intestinal niche.²⁰

Once absorbed, *p*C undergoes extensive first-pass metabolism in the intestine and liver, i.e., conjugation in enterocytes and hepatocytes, primarily forming *p*-cresol glucuronide (*p*CG) and *p*-cresyl sulfate (*p*CS)^{22,23} (Figure 1). Glucuronidation of *p*C is primarily mediated by UDP-glucuronosyl-transferase (UGT) enzymes, with UGT1A6 being the main isoform involved, responsible for over 78% of hepatic and over 54% of renal *p*C conjugation.²⁴ UGT1A6 activity follows a sigmoidal kinetic model, indicating the presence of multiple

substrate-binding sites.²⁵ Conversely, UGT1A9 contributes to a lesser extent and exhibits substrate inhibition at high *pC* concentrations.^{26,27} Although these amounts are unlikely under physiological conditions, levels approaching 700 μM , considered potentially toxic,²⁸ may impair UGT1A9 activity and limit *pC* detoxification via glucuronidation, possibly as a self-limiting or protective mechanism.

Sulfation of *pC* is primarily mediated by the SULT1A1 enzyme, which follows a Michaelis-Menten kinetic in the liver, but exhibits substrate inhibition in the kidneys, suggesting complex regulatory dynamics.²⁹ Even if hepatic sulfation has long been considered the main route for producing *pCS*, emerging evidence highlights a significant contribution from the intestinal epithelium.^{30,31} Indeed, SULT1A1 is expressed not only in the liver but also in the small intestine and colon, where it catalyzes sulfation of phenolic compounds.³⁰ This local intestinal expression aligns with the microbial origin of *pC* in the gut lumen, suggesting that sulfation may begin immediately after absorption. Indeed, transcriptomic and enzymatic studies demonstrate *pC*-induced upregulation of SULT1A1 in gut epithelial tissues, indicating a regulatory role of microbial metabolites on host detoxification enzymes.³²

Due to extensive first-pass metabolism, the predominant circulating forms of *pC* in humans are its conjugated metabolites.³³ Only a minor fraction of unconjugated *pC* remains in plasma, where it circulates predominantly in a protein-bound form.³⁴ Albumin plays a key role in modulating plasma levels, acting as the main carrier for both *pC* and *pCS*,^{34,35} with more than 90% of the latter being in the protein-bound form.²² This has been further confirmed by ultrafiltration studies identifying both high-affinity and low-affinity binding sites.³⁶ In contrast, *pCG* shows a lower binding affinity to albumin, with a larger fraction circulating in its free form.³⁷ However, binding affinity values vary across studies: while some report strong interactions, others describe only weak binding between *pC* or *pCS* and albumin, as observed by Bergé-Lefranc *et al.*³⁴ These discrepancies may be influenced by environmental factors like temperature, which can affect protein binding.³⁴ Notably, conditions such as hypoalbuminemia may alter the distribution of *pC* and its metabolites, increasing their unbound plasma fractions.³⁸

In healthy individuals, unconjugated *pC* is present in blood at very low concentrations, typically around 0.4 nM, while the majority circulates in its conjugated forms as a result of efficient first-pass metabolism.^{33,39} In conditions of impaired renal function, such as chronic kidney disease (CKD) and uremia, blood and organ concentrations of *pC*, *pCS*, and *pCG* rise significantly, due to reduced renal clearance.^{40,41} For example, serum *pCS* increases from 15 to 35 μM in healthy subjects to 116–568 μM in uremic conditions.²² Despite variable and disproportionate patterns described in the literature,^{40,42–45} *pCS* shows the most consistent correlation with CKD progression.^{46,47} Given their predominance in circulation, *pCS* and *pCG* are hypothesized to mediate most of the biological effects in the body.^{22,48}

The elimination of *pC* and its conjugated forms occurs predominantly by renal excretion, with urinary clearance playing a key role in maintaining systemic homeostasis^{15,49,50} (Figure 1). Among the conjugates, *pCS* is the principal urinary metabolite, actively secreted by renal tubular cells via organic anion transporters (OATs), particularly OAT1 and OAT3, which mediate its efficient clearance.⁵¹ In healthy subjects, about 95% of urinary *pC* is excreted as *pCS* and approximately

5% as *pCG*.⁵² Accordingly, reduced renal clearance in pathologies, such as CKD, may explain the increased accumulation of these compounds in both the blood and tissues. Another excretion route is through feces; indeed, *pC* and *pCS* have been detected in healthy individuals.^{53,54}

Alongside uremic conditions, elevated concentrations of *pC* and its conjugated forms have been described in both neurodevelopmental and neurodegenerative disorders, including ASD, PD, AD, and PTSD.^{55–58} The consistent increase in these gut-derived metabolites in these conditions suggests they may not merely reflect microbial imbalance or impaired host detoxification but actively contribute to disease processes. Their augmented presence in plasma, urine, feces, and even cerebrospinal fluid (CSF) underscores their ability to reach the CNS and influence neurological outcomes.^{55–58} As such, these molecules may represent potential mediators in the pathophysiology of gut-brain axis disorders.

MECHANISMS OF DAMAGE CAUSED BY *P*-CRESOL AND METABOLITES

pC and its primary metabolites, particularly *pCS*, have been associated with multiple pathological mechanisms across different organ systems and models.^{4,41,55,59–61} These compounds are increasingly recognized as contributors to neurotoxicity and systemic inflammation, all of which may be relevant in the context of neurodevelopmental and neurodegenerative disorders.^{55,59,62,63}

A summary of the main molecular pathways through which *pC* and its metabolites are implicated in mechanisms of toxicity and dysfunction is provided in Figure 2, along with an overview of the major disorders in which they are involved. The main molecular mechanisms underlying the *in vitro/in vivo* effects of *pC* and its metabolites are summarized in Table 2.

Among the various cellular targets affected by *pC* and its metabolites, the vascular endothelium, and, in particular, the blood-brain barrier (BBB), emerges as a critical site of vulnerability.^{64–66} *pCS* can impair BBB integrity by activating epidermal growth factor receptor (EGFR) signaling and promoting matrix metalloproteinase release, leading to extracellular matrix breakdown, cytoskeletal changes, and weakened cell junctions that increase permeability.⁶⁴ In brain microvascular endothelial cells, these effects translate into a direct compromise of BBB integrity, allowing peripheral pro-inflammatory factors and neurotoxic metabolites to penetrate into the CNS.⁶⁷ This loss of barrier selectivity is not only a consequence of systemic toxicity but a key amplifier of central damage, facilitating the widespread neuronal, metabolic, and immune alterations that will be discussed in the following sections.⁶⁷

Oxidative Stress (A). *pC* and *pCS* contribute to cellular and tissue damage, both systemically and in the CNS, primarily by inducing oxidative stress. Evidence, including a recent systematic review, shows that these uremic toxins disrupt redox balance, weaken antioxidant defenses, and increase reactive oxygen species (ROS), causing extensive molecular and mitochondrial damage.⁶⁸

One important mechanism by which *pCS* contributes to oxidative stress is through activation of NADPH oxidase (NOX), leading to the production of superoxide anions and other ROS.^{59,69,70} This NOX-mediated ROS generation, particularly in microglia and astrocytes, has been strongly linked to neuroinflammation and disruption of the BBB, processes involved in neurodegenerative and neurodevelop-

Table 2. Main *In Vitro* and *In Vivo* Effects of *pC* and Its Metabolites^a

experimental model	exposure	effect	refs
<i>In Vitro</i>			
hepatic tumor cell line (HepaRG)	<i>pC</i>	↑ DCF formation ↑ lactate dehydrogenase (LDH) release ↓ GSH concentration	41
human liver microsomes	<i>pC</i>	↑ cytochrome P450-mediated aromatic oxidation	74
mesenchymal stem cells (MSC)	<i>pC</i>	↑ apoptosis ↑ mitofusion ↓ mitophagy ↓ mitochondrial complexes I and IV activity	86
primary oligodendrocytes (from P6 mouse pup brain)	<i>pC</i>	impaired oligodendrocytes differentiation ↓ myelin gene expression	90
neuron2a neuroblastoma (N2a)	<i>pC</i>	↓ cell differentiation ↓ neurite length	92
pheochromocytoma (PC-12)	<i>pC</i>	↓ neurite length	92
rat hippocampal neurons	<i>pC</i>	↓ dendritic arborization ↓ synaptic density ↓ neuronal activity ↓ intracellular Ca ²⁺ signaling	92
rat pheochromocytoma cells (PC-12)	<i>pC</i> (low doses)	↑ neurite outgrowth ↑ NF expression ↑ BDNF secretion	101
human brain capillary endothelium model (hCMEC/D3)	<i>pC</i>	↑ paracellular permeability	109
conditionally immortalized human renal proximal tubule epithelial cells (ciPTEC)	<i>pC/pCS/pCG</i>	↓ mitochondrial succinate dehydrogenase activity	85
splenocytes	<i>pC</i>	↓ IFN- γ production ↑ IL-4 production	105
	<i>pCS</i>	↓ IFN- γ production ↑ IL-4 production ↓ % Th1 cells and Th1/Th2 ratio ↑ % Th2 cells	
human proximal tubular cells (HK-2)	<i>pCS</i>	↑ ROS production ↑ NOX activity ↑ mRNA levels of inflammatory cytokines ↑ active TGF- β 1 protein secretion	59
human brain capillary endothelium model (hCMEC/D3)	<i>pCS</i>	↑ BBB permeability ↓ endothelial junction disruption ↑ metalloproteinases activity	64
human endothelial cell line (EA.hy926)	<i>pCS</i>	↑ CREB/ATF1 signaling activation ↑ ATF1 protein levels ↑ endothelial inflammation ↑ oxidative stress	69
mouse 3T3-L1 adipocytes	<i>pCS</i>	↑ ROS production ↑ NADPH activation ↓ intracellular GSH content	70
human endothelial cell line (EA.hy926)	<i>pCS</i>	formation of extracellular vesicles (EVs)	81
human renal tubular cells (HK2)	<i>pCS</i>	aerobic/anaerobic metabolism impairment ↓ mitochondrial mass ↑ autophagy ↑ mitochondrial fission	84
primary cultures of hippocampal and cortical neurons	<i>pCS</i>	↑ neuronal dysfunction ↓ firing rate ↓ spikes/sec ↓ network bursts/sec	93
macrophage-like cell line (RAW246.7)	<i>pCS</i>	↓ IL-12 p40 production ↑ IL-10 production ↓ NO production ↓ LPS-induced CD40 expression on the cell surface	104
primary peritoneal macrophages	<i>pCS</i>	↓ IL-12 p40 production ↑ IL-10 production	104
neuroinflammation model of LPS-activated BV2 microglia	<i>pCS</i>	↓ ADAM10 and ADAM17 expression ↓ TNF- α and IL-6 release	106

Table 2. continued

experimental model	exposure	effect	refs
human brain capillary endothelium model (hCMEC/D3)	<i>pCG</i>	↓ constitutive microglia phagocytosis ↑ transendothelial electrical resistance	109
	<i>In Vivo</i>		
C57BL6/J mice	<i>pC</i>	↓ social interaction ↑ stereotyped/perseverative behaviors	60
Wistar rats	<i>pC</i>	↓ VTA dopaminergic neurons activity microbiota remodeling ↑ GLUN2B/GLUN2A ratio in Nac ↓ GLUN2B/GLUN2A ratio in hippocampus ↓ CREB phosphorylation in Nac ↓ Rac1 activity in Nac ↑ Rac1 activity in hippocampus	88
audiogenic seizure-prone Krushinski–Molodkina (KM) rats	<i>pC</i>	↓ GLUN2B/GLUN2A ratio in Nac ↑ GLUN2B/GLUN2A ratio in hippocampus ↓ CREB phosphorylation in hippocampus ↓ Rac1 activity in Nac ↑ Rac1 activity in hippocampus	88
inbred murine model of ASD (BTBR mice)	<i>pC</i>	↓ social interaction stereotyped/perseverative behaviors ↑ anxiety-like behavior ↑ DA turnover	96
Wistar rats	<i>pC</i>	↓ Na ⁺ /K ⁺ -ATPase activity ↓ total ATPase activity ↑ lipid peroxidation	102
5/6 nephrectomy Sprague–Dawley rats CKD model	<i>pCS</i>	↑ oxidative stress ↑ NOX activity ↑ upregulation of NOX4 ↑ renal damage ↑ TGF- β 1 production	59
unilateral nephrectomized C57BL/6 mice	<i>pCS</i>	↑ depression/anxiety-like and cognitive impairment ↑ oxidative stress ↑ neuroinflammation ↓ serum levels of BDNF and 5-HT ↑ serum levels of corticosterone ↑ pro-inflammatory cytokines ↓ neurogenesis and cell survival ↓ GSH and MDA	62
C57BL6/J mice	<i>pCS</i>	↑ BBB permeability ↓ neuronal activity	64
B-6 mice with 1/2-nephrectomy mice	<i>pCS</i>	mitochondrial function impairment	84
C57BL6/J mice	<i>pCG</i>	↓ BBB permeability ↓ pathways associated with cellular degradation and metabolism ↑ pathways associated with growth factor/transcription factor signaling	109

^aADAM: a disintegrin and metalloprotease; ATF1: activating transcription factor 1; BBB: blood–brain barrier; BDNF: brain-derived neurotrophic factor; CD40: cluster of differentiation 40; CREB: cAMP response element-binding protein; DA: dopamine; DCF: 2'-7'-dichlorofluorescein; GLUN2A: NMDAR subunit 2A; GLUN2B: NMDAR subunit 2B; GSH: glutathione; 5-HT: serotonin; IFN- γ : interferon-gamma; IL: interleukin; LDH: lactate dehydrogenase; LPS: lipopolysaccharide; MDA: malondialdehyde; Nac: Nucleus accumbens; NF: neurofilament; NO: nitric oxide; NOX:NADPH oxidase; NOX4: NADPH oxidase-4; ROS: reactive oxygen species; TGF- β 1:transforming growth factor β 1; TNF- α : tumor necrosis factor-alpha; and VTA: ventral tegmental area.

less clear, warranting further investigation to understand its precise role in neuroinflammation.

Synaptic Alterations (D). Guzmán-Salas *et al.* have studied *pC* effects on neuronal cells and hippocampal cultures, finding that this compound significantly reduced synaptic density by downregulating key synaptic markers (PCLO, SHANK, PSD-9S, VGAT), which are vital for synapse formation and neurotransmission.⁹² Prolonged exposure to *pC* was also associated with reduced spontaneous neuronal activity and lower intracellular calcium levels, factors that may underlie the compound's proposed role in exacerbating ASD symptoms, as

they compromise the functional dynamics of neural circuits.⁹² Additionally, even short-term *pC* exposure reduced neurite length for at least 24 h, while higher concentrations impaired dendritic arborization by inhibiting primary and secondary dendrite growth, changes similar to those seen in ASD.⁹² This supports the hypothesis that *pC* may play a role in abnormal neurodevelopment, consistent with earlier findings of reduced Rac activity, a protein important for dendritic growth and synaptic plasticity, following *pC* exposure.⁸⁸ Similarly, chronic exposure to *pCS* can reduce neuronal network activity by impairing the firing rate and burst frequency, interestingly,

without affecting mitochondrial function or oxidative stress.⁹³ Overall, these effects support a direct role for *pC* and *pCS* in functional synaptic disruption, potentially contributing to broader neurodevelopmental and neurodegenerative disorders.

Neurotransmitter Metabolism Dysfunction (E). Another key neurotoxic action of *pC* is its competitive inhibition of DBH, the enzyme that converts DA into norepinephrine (NE), an essential step for maintaining catecholamine balance.^{63,75,76,94} By covalent binding to DBH, *pC* alters normal DA metabolism, leading to its accumulation and reduced levels of downstream catecholamines. Clinical studies support this mechanism, showing that individuals with ASD have elevated urinary levels of both DA and *pC*, alongside reduced NE and epinephrine (EPI), suggesting functional DBH inhibition *in vivo*.⁹⁵ Experimental models further confirm that high *pC* exposure increases DA as well as its metabolites 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA) in various brain regions, while NE levels remain unchanged.⁹⁶ Furthermore, *pC* exposure was found to suppress the activity of dopaminergic neurons in the ventral tegmental area (VTA), reinforcing its role in dopaminergic dysregulation.⁶⁰

In contrast, *pCS* can interfere with critical neurotrophic and monoaminergic systems. Specifically, *pCS* reduces the levels of serotonin (5-HT) and brain-derived neurotrophic factor (BDNF), which are essential for neuronal survival, plasticity, mood regulation, and cognitive functions.⁶² In the prefrontal cortex, *pCS* lowers 5-HT levels, thus affecting serotonergic transmission,⁶² a process closely linked to depression, anxiety, social dysfunction, and the cognitive decline observed in neurodegenerative diseases, including both AD and PD.^{97,98} At the same time, *pCS* downregulates BDNF, which not only leads to cognitive decline, brain atrophy, and increased risk of psychiatric disorders but also impairs serotonergic neurons themselves.^{99,100} These effects are reinforced by the upregulation of REST, a transcriptional repressor that silences genes involved in both 5-HT and BDNF pathways, worsening neuropsychiatric vulnerability.⁶² Interestingly, *pCS* may exert a dual effect depending on its concentration; mild oxidative stress induced by low *pCS* levels can transiently stimulate BDNF release and activate redox-sensitive genes, promoting neuroplasticity, whereas higher levels can cause detrimental effects.¹⁰¹

Receptor Activity Disruption (F). The impact of this toxin on NMDA receptors (NMDARs) was explored by Gigi *et al.*,⁸⁸ who found that exposure to *pC* significantly altered the composition of NMDAR subunits, particularly increasing the GLUN2B/GLUN2A ratio in a region-specific manner. This effect is partly mediated by *pC* modulation of the interaction between NMDAR subunits and dopamine D1 receptors, affecting NMDAR surface expression and localization.⁹² Additionally, *pC* can affect the expression of neurofilaments (NFs), proteins that interact with both glutamatergic and D1 receptors and play a vital role in neuronal structural remodeling, further amplifying the changes induced by *pC*.¹⁰¹ Alongside these effects on receptor composition and structural proteins, *pC* is also associated with reduced activity of Na⁺/K⁺-ATPase, an essential enzyme for maintaining neuronal excitability and signaling, which may further contribute to its neurotoxic impact.¹⁰²

Systemic Immunosuppression (G). The potential damage caused by *pC* and, mainly, its metabolite *pCS*, appears to be significantly related to alterations in immune responses

across different cell types.^{103–105} *pCS* can suppress Th1-type cellular immune responses, a key component of the body's defense against infections and inflammation.¹⁰⁵ This was demonstrated *in vivo* through hypersensitivity tests, where *pCS* exposure led to decreased IFN- γ production and increased IL-4 levels in T cells, indicating a shift toward a Th2-type immune response that weakens effective Th1 defenses.¹⁰⁵ Moreover, *pCS* is able to alter macrophage functions by suppressing the production of pro-inflammatory cytokines, such as IL-12, while promoting the production of anti-inflammatory cytokines, like IL-10.¹⁰⁵ This imbalance, alongside a reduction in nitric oxide (NO) production and decreased CD40 expression on macrophages, further impairs Th1-type immune responses, potentially undermining the body's defense mechanisms.¹⁰⁴

Microglial Dysfunction and Impaired Brain Immune Responses (H). This immunosuppressive profile of *pCS* extends to the CNS, where it directly impacts microglia, the brain resident immune cells. In BV2 microglial cells, *pCS* inhibits both constitutive and lipopolysaccharide (LPS)-induced production of pro-inflammatory cytokines (TNF- α and IL-6) by inhibiting ADAM10 and ADAM17 enzymes, which are essential for cytokine processing and release.¹⁰⁶ This reduction suggests that *pCS* disrupts cytokine-mediated immune responses, potentially impairing immune signaling and contributing to neurological conditions, such as ASD, where immune dysregulation plays a key role.¹⁰⁶ Additionally, increased microglial CD68 expression after *in vivo* *pCS* exposure suggests microglial involvement in *pCS*-driven neuroinflammation.⁶²

***pCG* between Detoxification and Toxicity.** While the toxic effects of *pC* and, more consistently, its sulfate conjugate *pCS* have been well documented, the role of its glucuronide counterpart *pCG* remains less clearly defined. Several studies have demonstrated that *pCG* is significantly less toxic than both *pC* and *pCS* across various human cell types, including kidney, liver, and blood cells.^{41,44,85,107,108} In hepatocyte models, for example, *pCG* showed minimal cytotoxicity, even at elevated intracellular concentrations, suggesting that glucuronidation serves as an important detoxification mechanism to neutralize the harmful effects of its parent compound.⁴¹ In agreement with this, inhibiting *pCG* formation results in increased cellular damage, as indicated by a higher LDH release and the accumulation of free *pC*.⁴¹ Beyond its role in detoxification, these findings also challenge the traditional assumption that glucuronide conjugates are biologically inert and simply destined for renal excretion. Notably, *pCG* appears to work as a functional antagonist of toll-like receptor 4 (TLR4), preventing the receptor activation by bacterial endotoxins, such as LPS, thereby protecting the BBB from LPS-induced permeability.¹⁰⁹ This suggests a possible neuroprotective effect against pro-inflammatory stimuli of *pCG* under physiological conditions, particularly within the context of gut-brain communication. Nevertheless, *pCG* is classified as a uremic toxin, and it has been found at elevated levels either in patients with CKD or in those undergoing hemodialysis, where it may exert harmful effects.^{43,85,110} These findings highlight the need to re-evaluate glucuronide conjugates not merely as passive metabolic byproducts but as active modulators, particularly at the level of the cerebral vasculature. The contradictory nature of the current data highlights the need for further research to fully understand its impact on human health.

■ EFFECT OF *p*-CRESOL AND CONJUGATED METABOLITES ON NEURODEVELOPMENT: AUTISM

ASD comprises a range of neurodevelopmental disorders including autistic disorder, Asperger syndrome, and pervasive developmental disorder. Typically manifesting in early childhood, this pathology is primarily characterized by deficits in social interaction and communication, as well as by restricted interests and stereotyped behavior.¹¹¹ Clinical manifestations of ASD exhibit significant interindividual variability in terms of symptom presentation, severity, developmental trajectory, and treatment response. Over the past two decades, the incidence of ASD has increased markedly: recent estimates from the Autism and Developmental Disabilities Monitoring (ADDM) Network indicate that approximately one in every 36 children aged eight or older in the United States is diagnosed with ASD.¹¹² Consequently, a condition once considered rare has emerged as one of the most prevalent disorders in child neuropsychiatry.

Both genetic and environmental factors are implicated in the pathogenesis of ASD. Recent research has strongly highlighted the role of *p*C and its conjugated metabolites in ASD.⁵⁵ Elevated urinary levels of these compounds have been detected in the urine of a subset of ASD individuals. A study carried out in Italy by Altieri *et al.* in 2011, later replicated in France by Gabriele *et al.* in 2014, has highlighted significantly higher urinary levels of *p*C, *p*CS, and *p*CG in autistic children.^{111,113} In these studies, higher levels of total *p*C in urine were observed in ASD children,^{111,113} with particular regard to *p*CS, leading to the conclusion that *p*CS may be the real toxin under ASD onset.¹¹³ In addition, increased levels of *p*CS were found in females with ASD compared to males, as consistently reported across multiple studies,⁵⁵ including those by Altieri *et al.*,¹¹¹ Gabriele *et al.*,¹¹³ and more recently Osredkar *et al.*,¹¹⁴ suggesting possible sex-related differences in metabolism or microbiota composition rather than inherent severity. Nevertheless, more studies with larger and more gender-balanced cohorts are needed to verify this gender-related difference.

Other studies have pointed out that *p*CS levels are significantly higher in children with ASD children and comorbid conditions, such as epilepsy, hearing loss, developmental delays, ADHD, and tic disorders;⁵⁵ higher urinary levels of both *p*C and *p*CS were also positively correlated with the severity of ASD symptoms.^{55,114,115} It should be pointed out that in one of the studies cited it is not clear whether the authors refer to the total *p*C level in urine or only to the unconjugated form,¹¹⁵ making it difficult to clearly state if either *p*C or *p*CS or both are correlated to the severity of ASD. It is worth noting that urinary *p*C levels are especially high in ASD subjects with chronic gastrointestinal (GI) conditions linked to gut dysbiosis and abnormal abundance of *Clostridium* species,¹¹⁶ among the main producers of this toxin. Many studies have described a correlation between ASD and increased presence of *C. difficile*, contributing to explaining the increased levels of total *p*C in these individuals.^{117–119} Specifically, an abnormal abundance of *Clostridium* species in the intestines has been correlated with increased total urinary *p*C levels in autistic children under the age of 8.¹¹¹ Additionally, a correlation between environmental glyphosate exposure and *C. difficile* proliferation has been pointed out, suggesting that glyphosate, a common herbicide, may

contribute to gut dysbiosis by promoting the overgrowth of *Clostridium* species.¹²⁰

It has been shown that *p*C levels are also higher in the feces of autistic children compared to controls, with a negative correlation between these levels and age:¹²¹ this suggests that younger children with ASD may be exposed to greater concentrations of this toxin, potentially making it a useful biomarker for autism in early childhood.^{106,111,113,121} Meta-analyses were carried out on the studies correlating fecal levels of *p*C and ASD, but they were not conclusive; *p*C levels were consistently higher in the ASD group, but they did not reach a statistically significant difference.¹²²

Increased total urinary *p*C concentrations have been associated with stool consistency, indicating that slow intestinal transit and chronic constipation, both common in individuals with ASD, are key factors influencing its accumulation.^{116,123} In fact, approximately 40% of children with ASD have obvious functional GI issues such as diarrhea, constipation, bloating, and abdominal pain, which may be linked to an altered GMB and the overproduction of *p*C.⁵⁵ Further supporting this, a longitudinal study on chronically constipated autistic children (aged 2–8) has shown that accelerating intestinal transit through polyethylene glycol (PEG) administration led to a noticeable reduction in anxiety, hyperactivity, social interaction deficits, and stereotyped behaviors.¹¹⁶ This clinical improvement was accompanied by a decrease in the urinary *p*C level, although with some variability, likely due to the unpredictable effects of PEG on GMB composition.¹¹⁶ By shortening the intestinal transit time, PEG may either promote or inhibit *p*C production, depending on individual microbiome characteristics. Regardless of whether *p*C originates primarily from environmental exposure or gut bacterial metabolism, these findings highlight the crucial role of intestinal transit in determining its systemic levels, reinforcing the idea that GI health is closely linked to behavioral symptoms in ASD.¹¹⁶ It should be emphasized that the study by Turriziani *et al.*¹¹⁶ refers to total *p*C levels in urine and they do not discriminate among *p*CS, *p*CG, and free *p*C; the increased level of *p*C in urine, therefore, cannot be attributed to a specific compound.

The causative correlation between exposure to *p*C and ASD development has been increasingly highlighted through studies showing its potential to impair brain function and behavior. It has been observed that mice exposed to *p*C for 4 weeks in drinking water presented autistic-like symptoms, including social deficits, stereotypies, and perseverative behaviors, but no changes in anxiety, locomotion, or cognition were observed.⁶⁰ Similarly, BTBR mice, a murine model of ASD-like phenotype, displayed heightened anxiety and hyperactivity when administered a low dose of *p*C (1 mg/kg), while a higher dose (10 mg/kg) exacerbated typical ASD traits, including stereotypic behaviors and social deficits.⁹⁶ This increase in severity was accompanied by accelerated dopaminergic turnover in the amygdala and dorsal striatum, critical areas for emotional and motor control, providing further insight into how the exposure to *p*C alters brain chemistry in a way that mirrors the primary and secondary symptoms seen in humans with ASD.⁹⁶ In other studies, intraperitoneal injection of *p*C for 21 days considerably induces autism-like behavioral alterations in rats¹²⁴ and increases their seizure readiness, suggesting a possibility of a common mechanism involved in the *p*C-induced development of both autism and epilepsy.¹²⁵ Confirming a possible correlation between the two disorders, 40% of autistic children experience epilepsy.¹²⁵ Behavioral abnormalities were paral-

led by neurochemical alterations in mice, mainly involving the dopaminergic turnover.⁹⁶ This interpretation aligns with long-standing evidence of DBH inhibition exerted by *pC* and with the proportionate increase in DA and its metabolites, supporting increased DA accumulation, release, and catabolism (intra- and extra-cellular).⁹⁶ This has been proposed as a possible cause of the social and sensory issues observed in individuals with ASD.¹¹² Nevertheless, in all these studies it was not investigated whether *pC* itself or its metabolites was responsible for the effects.^{60,96,124,125} A study by Vinithakumari *et al.*, involving C57BL6/J mice treated with a mixture of antibiotics and *C. difficile* spores, has revealed neurotoxic effects after the induction of *C. difficile* infection.⁹⁴ These mice developed symptoms like diarrhea, weight loss, and increased DA levels in the nigrostriatal pathway.⁹⁴ Despite the elevated DA, a significant decrease in hippocampal NE was also observed, impairing their ability to consolidate social memories.⁹⁴ The authors observed that *C. difficile* infection leads to an increase in serum and intestinal *pC* levels, which could exert its effects on neurotransmission after crossing the BBB.⁹⁴

All of these findings suggest that *pC* has a profound impact on both neurotransmitter systems and the neural circuits that regulate behavior and cognition, suggesting that it may interfere with the neural circuits involved in phenotypical core aspects of ASD.

These biochemical changes, observed in murine models, are not limited to animals, as human studies mirror these findings. Indeed, abnormal levels of DA and NE have been observed in autistic children, strengthening the connection between neurotransmitter imbalances and ASD.⁹⁵ The analysis of urine samples from 40 autistic children has shown significantly higher levels of DA and HVA, coupled with lower levels of NE.⁹⁵ The cause of this neurotransmitter imbalance is thought to be linked to the ability of *pC* to inhibit DBH, as previously described in the “Mechanisms of damage caused by *p*-cresol and metabolites” section. This accumulation, particularly in the form of HVA, has been strongly correlated with the severity of ASD symptoms, including heightened agitation and stereotypic behaviors.⁹⁵

The existing studies on autism, taken together, strongly suggest that *pC* and its metabolites may play a crucial role in the pathogenesis of ASD. Human studies showed elevated urine levels of *pC* and its metabolites in certain ASD patients, particularly those with chronic GI issues linked to dysbiosis, indicating that these compounds could play a significant role in the development and progression of the disorder.¹²² A very recent meta-analysis on urine studies highlighted that the levels of *pC* and its conjugated metabolites could be a potential biomarker for ASD, also considering its strict link to the increased relative abundance of specific intestinal bacteria.¹²²

The mechanism through which *pC* and its conjugated metabolites exert their detrimental effects are likely to be related to their ability to boost oxidative stress in several districts, comprising brain tissues, and to alter the neurotransmitter system,⁶⁸ as described in the dedicated section of this review. Collectively, these effects exacerbate the pathophysiological onset, progression, and severity of a broad spectrum of diseases, including neurodevelopmental conditions, such as ASD.⁶⁸ Their occurrence in specific brain tissues could be due to the disruption of the BBB or mediated by OATs.⁶⁸

The presence of *pC* in autistic children, specifically produced by *C. difficile*, in an altered GMB scenario, further supports the concept of a gut-brain axis and offers new insights into possible therapeutic intervention, both pharmacological and nutritional.⁹⁵

Unfortunately, the available literature often does not specify or investigate whether the observed effects are exerted by unconjugated *pC* or by its conjugated metabolites, making it difficult to speculate which is most responsible for neurotoxicity. Nevertheless, recent findings emphasize the importance of considering environmental factors, such as *pC* exposure, when investigating the broader causes of ASD.

■ P-CRESOL AND CONJUGATED METABOLITES IN PARKINSON'S DISEASE

PD is a progressive neurodegenerative disorder that symptomatically evolves through a combination of motor symptoms, such as tremors and bradykinesia, and nonmotor symptoms, including GI dysfunctions.¹²⁶ Growing evidence indicates that PD can be viewed as a multifactorial disorder, with its onset and progression likely driven by an interplay of age, genetic factors, and environmental influences.^{127,128}

Etiopathogenesis of PD, notably the formation of Lewy bodies and the α -synuclein pathology, has been linked to the enteric nervous system, with emerging evidence suggesting that these structures may spread to the brain, further propagating the disease via cell-to-cell transmission.^{129,130} This gut-brain connection is further supported by clinical observations that GI symptoms often precede motor impairments by years, highlighting a potential role for the GMB both at the onset and throughout the course of the disease.^{131,132} Increasing evidence suggests that GI comorbidities, such as constipation, delayed colonic transit, and small intestinal bacterial overgrowth, are highly prevalent in PD patients.^{133–139} Among the many functions of the GMB, its role in metabolism is particularly relevant to PD, as it aids in the breakdown of dietary components, fermenting carbohydrates into short-chain fatty acids (SCFAs) and proteins into aromatic amino acid (AAA) metabolites.¹⁴⁰ These metabolites, including L-phenylalanine, L-tyrosine, and L-tryptophan, are involved in various physiological processes that regulate immune, inflammatory, metabolic, and neuronal responses in the gut and the brain. The dysregulation of these metabolic pathways is thought to contribute to the pathophysiology of neurodegenerative diseases, including PD.^{141,142,153} Imbalanced GMB may help explain the elevated levels of *pC* and its metabolites observed in PD patients, with these changes mainly attributed to bacteria from the *Lactobacillaceae* and *Bifidobacteriaceae* families.¹⁴³ Increased gut permeability, a feature also described in chronic CKD, further facilitates the circulation of uremic toxins and has similarly been reported in PD patients.¹⁴⁴ Environmental exposures to pesticides (such as paraquat and rotenone) and pollution have also been increasingly linked to PD,^{145–148} suggesting that microbial byproducts, including *pC* and its derivatives, are potentially influenced by these exposures and they may play a significant role in the pathogenesis of the disease.

In light of this, several studies have sought to deepen our understanding of how the GMB and its metabolites may contribute to PD. Cirstea *et al.*¹⁴⁹ recently provided evidence linking altered gut microbial metabolism, disturbed gut function, and PD. Compared to healthy controls, PD patients exhibited a reduction in butyrate-producing taxa, alongside an

increase in bacterial groups linked to *pC* synthesis. These microbial shifts were reflected in increased serum levels of *pC*, *pCS*, and phenylacetyl-L-glutamine, a metabolite derived from microbial processing of phenylalanine, which positively correlated with PD status as well as the severity of GI dysfunction, firmer stools, and constipation severity.¹⁶²

Untargeted metabolomic analyses of serum and plasma further supported the role of *pC* and its derivatives in PD-associated metabolic disturbances.^{25,150,151} Elevated levels of *pC* were observed only in serum, whereas increased *pCS* and *pCG* were detected in both serum and plasma of PD patients. The serum analysis also revealed a positive correlation between *pC* and its metabolites and age across both PD patients and controls, suggesting that aging might influence the metabolism or accumulation of these compounds. Interestingly, previous studies also reported that *pCS* levels do not correlate with its metabolic precursor, L-tyrosine, further supporting the idea that its accumulation may depend more on age-related changes than on precursor availability.¹⁵² On the contrary, this age-related correlation was not observed in plasma.¹⁵⁰

Notably, circulating *pCS* levels across studies were strongly associated with GI disturbances, such as constipation, but they were not related to motor impairments, indicating that these microbial metabolites may primarily contribute to GI dysfunction in PD.^{147,149} In contrast, *pCG* was consistently elevated in plasma and serum of PD patients and it positively correlated with motor symptom severity. This supports a potential link between *pCG* accumulation and the progression of motor symptoms in PD.^{150,151}

Beyond peripheral circulation, two recent studies have described elevated levels of *pCS* also in the CSF of PD patients compared to controls.^{56,154} Targeted metabolomics revealed that CSF levels of *pCS* in PD patients were approximately eight times higher than those in plasma, suggesting that individuals with PD accumulate more *pCS* in the brain than their healthy counterparts.⁵⁶ One likely contributor to the increased entry of potentially neurotoxic microbial metabolites into the brain is a compromised BBB, which is known to be more permeable in PD patients.^{155,156} In addition to passive diffusion through a more permeable BBB observed in PD patients, Sankowski *et al.* suggested that specific transport mechanisms might facilitate *pCS* entry into the brain.⁵⁶ OATs, particularly OAT3, are believed to be involved in this process. While primarily known for renal tubular secretion, some OAT isoforms, including OAT3, are also expressed in the brain, where they regulate metabolite exchange between the blood, cerebrospinal fluid, and brain tissue.^{157,158} Notably, OAT3 has been implicated in the clearance of other uremic toxins in the brain, such as IS, making it a likely candidate for mediating *pCS* transport as well.^{159,160} The disproportionate CSF-to-plasma ratio of *pCS* in PD patients may reflect a potential dysfunction in its distribution, potentially resulting from impaired OAT3 activity. This dysregulation could compromise the efflux of *pCS* from the CSF to the blood, thereby possibly contributing to its accumulation within the CNS.⁵⁶ Nevertheless, the role of OATs in mediating *pCS* transport across the BBB has not yet been directly investigated in the context of PD, and no studies have so far demonstrated altered expression or activity of these transporters in PD patients; their proposed involvement thus remains a hypothesis by the authors. Moreover, the significant correlation observed between *pCS* levels in plasma and CSF suggests that the systemic accumulation of this metabolite may influence its central concentration. This finding, together with

the markedly elevated CSF-to-plasma *pCS* ratio described in PD patients, points to possible impairments in its distribution or clearance at the brain-blood interface.⁵⁶ Notably, this imbalance was also associated with reduced kidney function, as indicated by lower estimated glomerular filtration rate (eGFR), suggesting that compromised renal clearance may increase circulating *pCS* levels and thereby promote its accumulation in the CNS. Taken together, these observations support a model in which both systemic (renal) and central (BBB and transporter-mediated) mechanisms may contribute to elevated levels of *pCS* in the PD brain.

Elevated *pCS* concentrations in CSF have also been detected in metabolomic-metallomic studies of PD patients, supporting the role of this metabolite in PD pathogenesis and progression.¹⁶¹ In PD, *pCS* accumulation appears to positively correlate with metallomic disturbances, especially involving copper and iron, metals known to contribute to oxidative stress and neurotoxicity. Moreover, elevated *pCS* levels were observed in adults with high plasma lead concentrations and reduced cognitive performance, suggesting that *pCS* could also be involved in lead-associated cognitive decline and broader aging-related processes such as those regulated by longevity-regulating pathways.

In summary, gut-derived metabolites, including *pC*, *pCS*, and *pCG*, eventually influenced by environmental exposure, are increasingly recognized as key players in PD, linking microbial dysbiosis, age-related changes, renal function, and BBB integrity to both motor and nonmotor symptoms. These effects may be mediated by mechanisms previously described, involving oxidative stress, mitochondrial dysfunction, neuroinflammation, and impairment of the BBB. While their exact mechanisms and transport pathways, such as the potential involvement of OAT3, remain to be fully clarified, these findings highlight the importance of the gut-brain axis in PD pathogenesis and open new avenues for therapeutic intervention targeting microbiome-related pathways.

■ P-CRESOL AND CONJUGATED METABOLITES IN ALZHEIMER'S DISEASE

AD is the most common form of neurodegenerative disorder in the elderly and the leading cause of dementia with few effective treatments currently available. The disease is characterized by pathological features, including amyloid- β -peptide ($A\beta$) plaques, tau protein neurofibrillary tangles, and neuroinflammation, all of which contribute to neurodegeneration. Increasing evidence points to a pivotal role of neuroinflammation in the pathogenesis and progression of AD.^{163–165} It is now known that the GMB plays a pivotal role in regulating neuroinflammation in various neurological conditions, including AD.^{58,166–169}

Recent studies indicate that patients with AD exhibit an altered GMB composition compared to individuals without AD,^{168,170,171} and experimental manipulations of the GMB in mouse models have demonstrated its capacity to influence AD-related pathology and neuroinflammation.^{172,173}

Even if the exact mechanisms linking GMB alterations to AD progression remain unclear, modifications of GMB composition can affect microbial-derived metabolites, which eventually modulate CNS immune responses in neurological diseases, including AD. In an AD mouse model, it has been observed that GMB dysbiosis is required to infiltrate peripheral immune cells into the brain.¹⁷⁴ Once they enter the CNS, immune cells contribute to neuroinflammation and associated cognitive

deficits. On the other hand, restoring GMB composition in APP/PS1 mice, a transgenic model of AD, mitigates neuroinflammation and reverses cognitive impairments.¹⁷⁴

Evidence suggests a heightened risk of developing AD and other neurological conditions among patients with CKD,^{175–177} possibly due to the accumulation of protein-bound uremic toxins, such as *pCS* and IS.¹⁷⁸ These toxins, primarily derived from gut microbial metabolism, are known to contribute to cognitive decline and neurodegenerative processes.^{62,179} On the other hand, the treatment with AST-120, an oral spherical activated carbon that acts as a uremic toxin adsorbent, resulted in protection against cognitive and emotional impairment in CKD patients,^{179,180} thus confirming the involvement of these molecules in impaired CNS functioning.

Overall, gut dysbiosis, involved in both CKD and AD conditions, exacerbates the production of uremic toxins like *pCS*, which, in turn, are known to increase vascular permeability, including that of the BBB.^{64,65,181,182} This results in higher levels of uremic toxins not only in the periphery (i.e., serum/plasma) but also in the brain,⁹¹ where these toxins can induce oxidative stress, mitochondrial dysfunction, and neuroinflammation, hallmark features of AD.^{84,183,184}

The work performed by Sun *et al.* has further highlighted the potential role of *pCS* in the pathogenesis of AD.⁶² *In vivo* behavioral analyses revealed that mice exposed to *pCS* developed anxiety- and depressive-like behaviors, symptoms frequently observed in AD. Moreover, exogenous administration of *pCS* induced notable cognitive impairments, alongside increased oxidative stress, neuroinflammation, and reduced levels of BDNF and 5-HT, all of which are hallmark features of neurodegenerative diseases.⁶² Consistently, clinical studies have described decreased serum levels of both BDNF and 5-HT in AD patients,^{185,186} further supporting a mechanistic link between *pCS* exposure and neurodegenerative pathologies. Despite the limited number of studies focusing specifically on the relationship between *pC* and AD, metabolomic analyses have consistently shown elevated levels of its primary derivatives, *pCS* and *pCG*, in AD patients.^{187,188} A targeted metabolomic analysis of AD plasma and brain tissues performed by Kalecký *et al.* in nonhispanic whites has revealed notable changes in small molecules linked to GMB activity, reinforcing the critical role of the gut-brain axis in disease progression.¹⁸⁷ Among these metabolites, elevated levels of *pCS* were found in plasma and approached statistical significance in the frontal cortex.¹⁸⁷

Similarly, Gordon *et al.* have found that higher *pCG* levels were strongly associated with adverse brain aging and cognitive decline in AD patients.¹⁸⁸ This was further supported by a cross-sectional study carried out in Japan, that showed increased fecal *pC* levels in dementia patients compared to those without dementia (57.5 $\mu\text{g/g}$ vs 0.29 $\mu\text{g/g}$), thus reinforcing the growing evidence for the role of this compound and its metabolites in neurodegenerative diseases.¹⁸⁹

Overall, emerging evidence supports a potential link between gut-derived uremic toxins, particularly *pC* and its metabolites, and the pathogenesis of AD. *pCS* and *pCG*, generated by gut microbial fermentation, accumulate in circulation under conditions such as gut dysbiosis and CKD, both linked to AD. Mechanistically, they increase BBB permeability, allowing their entry into the brain, where they trigger synaptic impairment and cognitive decline. Elevated *pCS* and *pCG* levels in AD patients further support their

involvement, highlighting the gut–brain axis as a potential therapeutic target in AD.

Despite these findings, the relationship between *pC* and AD remains less thoroughly investigated than its correlation to ASD and PD, highlighting a gap in the current research.

■ EFFECT OF P-CRESOL AND CONJUGATED METABOLITES ON POST-TRAUMATIC STRESS DISORDER

PTSD is a trauma- and stressor-related disorder characterized by symptoms, such as intrusion, avoidance, negative alterations in cognition and mood, and hyperarousal, as defined by the Diagnostic and Statistical Manual of Mental Disorders.¹⁹⁰ These symptoms significantly affect the quality of life and often persist long after a traumatic event. While much of the early research on PTSD focused on psychological and neurological mechanisms, recent advances highlight the role of the GMB axis in mediating the disorder.^{191,192} Mendelian randomization studies in humans have demonstrated a causal association between specific bacterial taxa and PTSD risk.¹⁹³ Protective taxa, such as *Porphyromonadaceae* and *Veillonellaceae*, contrast with others like *Phascolarctobacterium* and *Ruminococcaceae*.¹⁹³ Malan-Muller *et al.* have identified a consortium of bacterial genera associated with PTSD severity, suggesting that GMB dysbiosis may exacerbate trauma-related symptoms.¹⁹¹

The interplay between GMB and mental health, with regard to PTSD, has emerged as a critical field of investigation.¹⁹⁴ Indeed, GMB plays a key role in understanding the pathophysiology of PTSD and related neuropsychiatric conditions.¹⁹⁵ Several studies underscore the critical role of GMB in regulating stress responses, trauma recovery, and neuropsychiatric health.^{196–198} The findings of these studies suggest that microbial alterations may contribute to the onset and persistence of PTSD. In particular, GMB can influence inflammation, stress responses, and neurotransmitter signaling, thus contributing to PTSD onset; in addition, microbial metabolites, immune system activation, and the vagus nerve may represent the mechanism through which the bidirectional communication between the gut and brain is implemented.¹⁹⁴ The interplay between the microbiome and the host autonomic, neuroendocrine, and immune systems may impact stress resilience and susceptibility to trauma- and stressor-related disorders, including PTSD.¹⁹⁹

Recent research has focused on the role of microbial metabolites, such as *pC*, in oligodendrocyte differentiation and myelination, which represent critical processes for maintaining neural connectivity and behavioral stability.¹⁹³ This interaction underscores the microbiota's influence on brain regions, such as the PFC, which is pivotal for higher-order cognitive functions and emotional regulation.

In a PTSD preclinical mouse model, it has been demonstrated that changes in gut microbial composition may influence neuropsychiatric outcomes, including resilience or susceptibility to stress.⁵⁷ Alterations in GMB, whether pre-existing or induced by trauma, can increase trauma susceptibility by enhancing the production of certain metabolites such as *pC*.⁵⁷ These metabolites can influence the dopaminergic system, in a manner that varies depending on the brain region. A study by Laudani *et al.*, using the arousal-based individual screening model, has provided evidence for pretrauma and post-trauma GMB alterations in susceptible mice exhibiting persistent PTSD-related phenotypes.⁵⁷ A more in-depth analysis revealed an increased abundance of pro-

inflammatory bacteria (*Ruminococcaceae* and *Lachnospiraceae*) affecting brain processes, including myelination, and brain systems such as the dopaminergic neurotransmission.⁵⁷ Whether these alterations in GMB composition could be associated with abnormal levels of metabolites that induce dopaminergic dysfunctions was investigated in the study of Laudani *et al.*⁵⁷ High levels of *pC* were detected exclusively in the PFC of susceptible mice.⁵⁷ In addition, abnormal levels of DA and DOPAC were observed, together with a detrimental increase of D3 receptor expression in the same area.⁵⁷ Conversely, either resilience mechanisms aimed at counteracting these *pC*-induced dopaminergic dysfunctions or myelination-related resilience mechanisms were observed only in the PFC of resilient mice.⁵⁷ These findings are in agreement with previous evidence on *pC*, which was demonstrated to be able to affect myelination by blocking myelin gene expression and differentiation⁹⁰ and to affect the dopaminergic system.⁷⁶

PTSD is associated with significant alterations not only in GMB composition but also in immune system function. A work carried out by Petakh *et al.* has pointed out the interaction between PTSD, gut dysbiosis, and elevated inflammatory biomarkers, suggesting that microbial metabolites modulate immune and brain functions in PTSD.²⁰⁰

pCS has also been linked with behavioral disorders and neuroinflammation.⁶² In a study by Sun *et al.* on unilateral nephrectomized mice, it was observed that, after administration of the protein-bound uremic toxin *pCS* at a dose of 100 mg/kg/day, the serum *pCS* concentration was progressively increased; in particular, there was an accumulation of *pCS* in the PFC tissues, and this was associated with several behavioral disorders, such as depression, anxiety, and cognitive impairment.⁶² In addition, the authors have demonstrated that *pCS* exacerbates oxidative stress and neuroinflammation in CNS.⁶² It should be pointed out that this study relied on the administration of a supraphysiological dose of *pCS* and that the end points were not focused on PTSD onset. Nevertheless, these findings suggest that *pCS* and similar neurotoxic metabolites further complicate the gut-brain axis in chronic conditions and they may contribute to neurodegenerative processes observed in PTSD.⁶²

The damages at the brain level exerted by *pC* and *pCS* could be explained by their capacity to cross BBB by various mechanisms.⁶⁸ Even if only the study by Laudani *et al.*⁵⁷ to date has demonstrated a possible link between the presence of *pC* in specific brain regions and the onset of typical PTSD symptoms, this study has opened new perspectives for investigation and potential therapeutic interventions. Clearly, further research is needed to confirm these results and to elucidate the mechanisms through which *pC* may contribute to neuropathological processes in the context of PTSD.

The therapeutic implications of targeting GMB in PTSD are relevant. Strategies based on probiotics, prebiotics, and dietary modifications can potentially modulate gut dysbiosis, thus reducing inflammation and potentially alleviating psychiatric symptoms.¹⁹⁴

■ THERAPEUTIC INTERVENTIONS

Given the neurotoxic role of *pC*, a reduction in its central concentration could help mitigate the molecular, cellular, and, consequently, behavioral effects associated with this metabolite.^{62,193,201} This goal could be achieved through various strategies, including modifying the composition of the GMB

(i.e., fecal transplantation, prebiotics and/or probiotics administration, and diet) or modulating its distribution and elimination.

In nephrectomized mice, an animal model of impaired kidney functionality, central *pCS* accumulation has been associated with depressive-like and anxiety-like behavior and cognitive impairments.⁶² These effects were alleviated by the administration of the uremic toxin adsorbent AST-120, which also reduces neuroinflammation, oxidative stress, apoptosis, and the negative impact on neurogenesis and neurotrophic support induced by *pCS*.⁶² Clinically, AST-120 is used to slow the progression of kidney dysfunction in CKD due to its ability to bind uremic toxins like *pC* and IS. Recently, it has been demonstrated that the administration of AST-120 is also effective in improving cognitive and emotional functions in CKD animal models.¹⁷⁹ Although these effects may be related to the reduction of other uremic toxins, mainly IS, it has been shown that an AST-120 treatment can prevent CKD-induced hippocampal damage (i.e., gliosis and impaired plasticity) and *pCS* accumulation within the hippocampus.^{202,203}

Another strategy to modify systemic and central *pC* levels and related metabolites could be fecal microbiota transplantation (FMT). Indeed, FMT has emerged as a potential therapy for several diseases associated with dysbiotic GMB, including neurodevelopmental and neurodegenerative diseases.²⁰⁴ In recurrent patients suffering from *C. difficile* infection (rCDI), FMT rapidly increases urinary levels of *pCS*, *pCG*, and fecal *pC* levels.⁵ Data from an animal model also confirm that *pC* biogenesis is more responsive to FMT treatment than other microbial metabolic pathways. These findings not only suggest that these metabolites can serve as reliable markers of the response to FMT treatment but also indicate that the effectiveness of these therapies may depend on their enhanced elimination.⁵

In patients treated with *Lactobacillus gasseri* CP2305, the fecal levels of *pC* significantly decreased compared to the control group, and this effect is paralleled by an improvement of intestinal functionality. Moreover, it is interesting to note that treatment with CP2305 has also been shown to improve sleep quality in adults with mild to moderate stress²⁰⁵ and to inhibit the effects on food intake in a subchronic and mild social defeat stress mouse model.²⁰⁶

Finally, a strategy to limit *pCS* toxicity may involve the modulation of OAT, as they are involved in the uptake and elimination of uremic toxins, including *pCS*.¹⁵⁷ They work by mediating the active transport of organic anions from the bloodstream into cells, where they can be further metabolized or excreted through the kidneys.^{157,207} Specifically, *pCS* represents a substrate for OATs, being preferentially recognized by OAT1 and OAT3, as demonstrated using *in vitro* and *ex vivo* models.^{51,206}

The relationship between the OAT transporters and *pCS* highlights an important aspect of kidney disease management. Understanding the mechanisms behind *pCS* transport and accumulation can provide insights into potential therapeutic strategies for patients with CKD. For example, modulating the activity of the OAT transporter or developing inhibitors of *pCS* production might offer novel ways to alleviate uremic toxicity and improve patient outcomes. Additionally, the study of the transporters of the OAT in relation to *pCS* can be crucial for developing new pharmacological treatments to enhance the clearance of uremic toxins. These interventions could help

improve the quality of life for individuals with impaired renal function, especially those undergoing dialysis.

Moreover, the involvement of OAT transporters in regulating the influx and efflux of uremic toxins, such as *pCS*, across the BBB is of significant interest.^{160,208} If OAT transporters in the brain, particularly OAT1 and OAT3 are impaired, whether due to kidney dysfunction or other factors, *pCS* could accumulate within the brain, exacerbating its neurotoxic effects and contributing to cognitive and mood disorders.⁵⁶ Further research is necessary and, aimed at investigating potential therapeutic strategies to enhance the function of the OAT transporter and to mitigate the detrimental effects of *pC* and its metabolites in the CNS, thus reducing the neurotoxic burden of this compound and improving cognitive function and mental health in individuals affected by the neurological pathologies described in this review. Specifically, molecular dynamics simulations can reveal the conformational changes of the transporters and identify the interactions between the substrates and the protein during the binding, translocation in the transporter cavity, and release of the substrate on the other side of the membrane.²⁰⁹ By combining the proteomics of OATs together with computational drug discovery, the rational design of transporter-modulating compounds could be successfully achieved.

These approaches could help reduce systemic and central levels of *pC* and may represent effective therapeutic strategies to improve cognitive and emotional functions in patients with neurodevelopmental or neurodegenerative disorders associated with this compound.

■ CRITICAL APPRAISAL OF THE LITERATURE

Although there is increasing evidence on the relevance of *pC* and its conjugated metabolites (*pCS* and *pCG*) in the onset of different neurodevelopmental, neurodegenerative, and neuropsychiatric disorders, several limitations of the cited studies are worth mentioning, in order to have a critical view on the reported results.

First, several studies have described the detection of free *pC* in plasma or serum; however, the accuracy of these findings has been increasingly questioned, due to significant methodological limitations. Indeed, many studies do not rely on an analytical methodology capable of discriminating between *pC* itself and its conjugated metabolites. Furthermore, analytical artifacts, due to the spontaneous hydrolysis of *pCS* into *pC* during sample preparation, should be considered, leading to a possible misinterpretation. These methodological issues can give the false impression that free *pC* is present in the samples analyzed. This aspect is crucial given the fact that *pC* undergoes massive sulfation, limiting the effective amount of circulating unconjugated compound. These insights highlight the need for appropriate analytical methods and support the use of validated, highly specific analytical platforms, particularly HPLC-MS/MS assays with targeted detection of conjugated forms. In addition, frequently authors do not provide any quantitative data on average concentrations detected, making a comparison of the studies and of the results almost impossible.

As to the *in vitro* and *in vivo* mechanistic studies, the main limitations are related to either a supraphysiological exposure, far exceeding expected systemic levels, or to an inappropriate administration route selected by the authors (i.e., intraperitoneal). Even if these models shed light on cellular and molecular pathways, they do not accurately mirror the typical exposure profiles in humans.

For what concerns studies on humans, the possibility to make conclusions either on sex-related differences or on the correlation between the toxin level and the severity of the disease is limited by the unbalance of the cohorts considered and by the small number of studies described in the literature, as in the case of ASD. For other pathologies, such as AD and PTSD, the number of studies investigating the role of *pC* and conjugated metabolites is still very limited, and therefore, a causation-effect correlation is still preliminary.

■ CONCLUSIONS


An increasing number of studies strongly support the role of *pC* and its primary human metabolites (*pCS* and *pCG*), in the development and progression of neurodevelopmental and neurodegenerative disorders. These multifactorial conditions share common features, including the involvement of environmental risk factors and GMB alterations, which are often characterized by dysbiosis. Indeed, *pCS* is considered a biomarker of dysbiosis and it may act as a key mediator of damage within the gut–brain axis. Elevated levels of this metabolite, detected both in the periphery and in the CNS, have been reported in several CNS diseases. Both *pCS* and *pC* have been shown to induce cellular damage in neuronal and glial cells.


Mechanistically, oxidative stress, mitochondrial dysfunction, and altered inflammatory signaling triggered by *pC* may contribute to its detrimental effects on CNS functioning, impairing cell differentiation, survival, and brain plasticity.

Further studies are needed to better elucidate causal relationships, rather than merely correlative links, between *pC* and the pathogenesis of brain disorders. These insights could significantly advance the development of preventive and therapeutic strategies, including lifestyle interventions and targeted approaches to restore eubiosis, aiming to mitigate or even prevent the progression of these complex and highly prevalent and debilitating diseases.


■ AUTHOR INFORMATION

Corresponding Authors

Federica Pellati – Department of Life Sciences, University of Modena and Reggio Emilia, 41125 Modena, Italy;  orcid.org/0000-0002-9822-6862; Phone: +39 059 2058565; Email: federica.pellati@unimore.it; <https://personale.unimore.it/rubrica/dettaglio/fpellati>

Silvia Albani – Department of Life Sciences, University of Modena and Reggio Emilia, 41125 Modena, Italy; Center for Neuroscience and Neurotechnology, University of Modena and Reggio Emilia, 41121 Modena, Italy;  orcid.org/0000-0002-2332-3166; Phone: +39 059 2055360; Email: salboni@unimore.it; <https://personale.unimore.it/rubrica/dettaglio/salboni>

Authors

Laura Bertarini – Department of Life Sciences, University of Modena and Reggio Emilia, 41125 Modena, Italy; Clinical and Experimental Medicine PhD Program, Department of Biomedical, Metabolic and Neural Sciences, University of Modena and Reggio Emilia, 41125 Modena, Italy;  orcid.org/0009-0006-9576-2551

Federico Imbeni – Department of Life Sciences, University of Modena and Reggio Emilia, 41125 Modena, Italy

Virginia Brighenti – Department of Life Sciences, University of Modena and Reggio Emilia, 41125 Modena, Italy

Isabella Martusciello – Department of Life Sciences,
University of Modena and Reggio Emilia, 41125 Modena,
Italy; orcid.org/0009-0006-0673-7494

Complete contact information is available at:
<https://pubs.acs.org/10.1021/acspptscli.5c00289>

Author Contributions

S.A. and F.P. were involved in the conceptualization, reviewed and edited the manuscript for intellectual content, and approved the final draft; L.B., F.I., V.B., and I.M. wrote the first draft of the paper and contributed to the literature search, manuscript formatting, and figure design. All authors agree to be accountable for all aspects of the work.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or no-profit sectors.

Notes

The authors declare no competing financial interest.

■ ABBREVIATIONS

AAA	aromatic amino acid
A β	amyloid- β peptide
AD	Alzheimer's disease
ADAM	a disintegrin and metalloprotease
ADDM	autism and developmental disabilities monitoring
ADHD	attention-deficit/hyperactivity disorder
ADME	absorption-distribution-metabolism-excretion
Akt	protein kinase B
ASD	autism spectrum disorder
ATF1	activating transcription factor 1
ATP	adenosine triphosphate
BBB	blood–brain barrier
BDNF	brain-derived neurotrophic factor
CD40	cluster of differentiation 40
CKD	chronic kidney disease
CNS	central nervous system
CREB	cAMP response element-binding protein
CSF	cerebrospinal fluid
D1	dopamine receptor D1
D3	dopamine receptor D3
DA	dopamine
DBH	dopamine β -hydroxylase
DCF	2'-7'-dichlorofluorescein;
DOPAC	3,4-dihydroxyphenylacetic acid
eGFR	estimated glomerular filtration rate
EGFR	epidermal growth factor receptor
EPI	epinephrine
EVs	extracellular vesicles
FMT	fecal microbiota transplantation
GI	gastrointestinal
GLUN2A	NMDAR subunit 2A
GLUN2B	NMDAR subunit 2B
GMB	gut microbiome
GSH	glutathione
HpdBCA	4-hydroxyphenylacetate decarboxylase
HPLC–MS/MS	high-performance liquid chromatography-tandem mass spectrometry 5-HT, serotonin
HVA	homovanillic acid
IFN- γ	interferon-gamma
IL	interleukin

IS	indoxyl-sulfate
JNK	c-Jun N-terminal kinase
LDH	lactate dehydrogenase
LPS	lipopolysaccharide
MAP-2	microtubule-associated protein 2
MDA	malonyldialdehyde
Nac	nucleus accumbens
NE	norepinephrine
NF	neurofilament
NF- κ B	nuclear factor kappa B
NMDA	N-methyl-D-aspartate
NMDAR	N-methyl-D-aspartate receptor
NLRP3	NLR family pyrin domain containing 3
NO	nitric oxide
NOX	NADPH oxidase
NOX4	NADPH oxidase-4
OAT	organic anion transporter
p38	p38 mitogen-activated protein kinases
pC	p-cresol
pCG	p-cresol glucuronide
PCLO	piccolo
pCS	p-cresyl sulfate
PD	Parkinson's disease
PEG	polyethylene glycol
PFC	prefrontal cortex
pHPA	p-hydroxyphenylacetic acid
PKA	protein kinase A
PSD-95	postsynaptic density protein 95
PTSD	post-traumatic stress disorder
REST	repressor element-1 silencing transcription factor
ROS	reactive oxygen species
SULT1A1	sulfotransferase-1A
SCFAs	short-chain fatty acids
SHANK	SH3 and multiple ankyrin repeat domains protein
TGF- β 1	transforming growth factor β 1
TLR4	toll-like receptor 4
TNF- α	tumor necrosis factor-alpha
UGT	UDP-glucuronosyltransferase
VGAT	vesicular GABA transporter
VTA	ventral tegmental area

■ REFERENCES

- (1) Kuzminskis, V.; Ziginskiene, E.; Arune Bumblyte, I. A Registry of Haemodialysis Patients and the Progress of Haemodialysis Services in Lithuania. *Nephrol., Dial., Transplant.* **2005**, *20* (12), 2623–2628.
- (2) U.S. Department of Health and Human Services Toxicological Profile for Cresols, 2008. <https://www.atsdr.cdc.gov/toxprofiles/tp34.pdf>.
- (3) World Health Organization, International Programme on Chemical Safety, United Nations Environment Programme & International Labour Organization *Cresols: Health and Safety Guide*; World Health Organization: Geneva, 1996.
- (4) Zheng, Y.; Bek, M. K.; Prince, N. Z.; Peralta Marzal, L. N.; Garsen, J.; Perez Pardo, P.; Kraneveld, A. D. The Role of Bacterial-Derived Aromatic Amino Acids Metabolites Relevant in Autism Spectrum Disorders: A Comprehensive Review. *Front. Neurosci.* **2021**, *15*, 738220.
- (5) Zhou, Y.; Bi, Z.; Hamilton, M. J.; Zhang, L.; Su, R.; Sadowsky, M. J.; Roy, S.; Khoruts, A.; Chen, C. P-Cresol Sulfate Is a Sensitive Urinary Marker of Fecal Microbiota Transplantation and Antibiotics Treatments in Human Patients and Mouse Models. *Int. J. Mol. Sci.* **2023**, *24* (19), 14621.

- (6) Wikoff, W. R.; Anfora, A. T.; Liu, J.; Schultz, P. G.; Lesley, S. A.; Peters, E. C.; Siuzdak, G. Metabolomics Analysis Reveals Large Effects of Gut Microflora on Mammalian Blood Metabolites. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106* (10), 3698–3703.
- (7) Windey, K.; De Preter, V.; Verbeke, K. Relevance of Protein Fermentation to Gut Health. *Mol. Nutr. Food Res.* **2012**, *56* (1), 184–196.
- (8) Saito, Y.; Sato, T.; Nomoto, K.; Tsuji, H. Identification of Phenol- and p-Cresol-Producing Intestinal Bacteria by Using Media Supplemented with Tyrosine and Its Metabolites. *FEMS Microbiol. Ecol.* **2018**, *94* (9), fiy125.
- (9) Salmean, Y. A.; Segal, M. S.; Palii, S. P.; Dahl, W. J. Fiber Supplementation Lowers Plasma P-Cresol in Chronic Kidney Disease Patients. *J. Renal Nutr.* **2015**, *25* (3), 316–320.
- (10) Nakabayashi, I.; Nakamura, M.; Kawakami, K.; Ohta, T.; Kato, I.; Uchida, K.; Yoshida, M. Effects of Synbiotic Treatment on Serum Level of P-Cresol in Haemodialysis Patients: A Preliminary Study. *Nephrol., Dial., Transplant.* **2011**, *26* (3), 1094–1098.
- (11) Kawakami, K.; Marino, I.; Asahara, T.; Kato, I.; Onoue, M. Dietary Galacto-Oligosaccharides Mixture Can Suppress Serum Phenol and p-Cresol Levels in Rats Fed Tyrosine Diet. *J. Nutr. Sci. Vitaminol.* **2005**, *51* (3), 182–186.
- (12) Evenepoel, P.; Claus, D.; Geypens, B.; Maes, B.; Hiele, M.; Rutgeerts, P.; Ghoo, Y. Evidence for Impaired Assimilation and Increased Colonic Fermentation of Protein, Related to Gastric Acid Suppression Therapy. *Aliment. Pharmacol. Ther.* **1998**, *12* (10), 1011–1019.
- (13) Cummings, J.; Hill, M.; Bone, E.; Branch, W.; Jenkins, D. J. A. The Effect of Meat Protein and Dietary Fiber on Colonic Function and Metabolism II. Bacterial Metabolites in Feces and Urine. *Am. J. Clin. Nutr.* **1979**, *32* (10), 2094–2101.
- (14) Kawakami, K.; Kojima, K.; Makino, I.; Kato, I.; Onoue, M. Fasting Enhances P-Cresol Production in the Rat Intestinal Tract. *Exp. Anim.* **2007**, *56* (4), 301–307.
- (15) Bone, E.; Tamm, A.; Hill, M. The Production of Urinary Phenols by Gut Bacteria and Their Possible Role in the Causation of Large Bowel Cancer. *Am. J. Clin. Nutr.* **1976**, *29* (12), 1448–1454.
- (16) Bakke, O. M. Urinary Simple Phenols in Rats Fed Diets Containing Different Amounts of Casein and 10% Tyrosine. *J. Nutr.* **1969**, *98* (2), 217–221.
- (17) Ji, M.; Du, H.; Xu, Y. Structural and Metabolic Performance of P-Cresol Producing Microbiota in Different Carbon Sources. *Food Res. Int.* **2020**, *132*, 109049.
- (18) Elsdon, S. R.; Hilton, M. G.; Waller, J. M. The End Products of the Metabolism of Aromatic Amino Acids by Clostridia. *Arch. Microbiol.* **1976**, *107* (3), 283–288.
- (19) Jenner, A. M.; Rafta, J.; Halliwell, B. Human Fecal Water Content of Phenolics: The Extent of Colonic Exposure to Aromatic Compounds. *Free Radicals Biol. Med.* **2005**, *38* (6), 763–772.
- (20) Passmore, I. J.; Letertre, M. P. M.; Preston, M. D.; Bianconi, I.; Harrison, M. A.; Nasher, F.; Kaur, H.; Hong, H. A.; Baines, S. D.; Cutting, S. M.; Swann, J. R.; Wren, B. W.; Dawson, L. F. Para-Cresol Production by Clostridium Difficile Affects Microbial Diversity and Membrane Integrity of Gram-Negative Bacteria. *PLoS Pathog.* **2018**, *14* (9), No. e1007191.
- (21) Dawson, L. F.; Donahue, E. H.; Cartman, S. T.; Barton, R. H.; Bundy, J.; McEnerney, R.; Minton, N. P.; Wren, B. W. The Analysis of Para-Cresol Production and Tolerance in Clostridium Difficile O27 and O12 Strains. *BMC Microbiol.* **2011**, *11* (1), 86.
- (22) Grypp, T.; Vanholder, R.; Vanechoutte, M.; Glorieux, G. P-Cresol Sulfate. *Toxins* **2017**, *9* (2), 52.
- (23) Kuzminskis, V.; Bumblytė, I. A.; Šurkus, J. Development of Nephrology and Renal Replacement Therapy in Lithuania since 1989—an Update. *Nephrol., Dial., Transplant.* **1999**, *14* (12), 2846–2848.
- (24) Rong, Y.; Kiang, T. K. L. Characterizations of Human UDP-Glucuronosyltransferase Enzymes in the Conjugation of p-Cresol. *Toxicol. Sci.* **2020**, *176* (2), 285–296.
- (25) Shou, M.; Dai, R.; Cui, D.; Korzekwa, K. R.; Baillie, T. A.; Rushmore, T. H. A Kinetic Model for the Metabolic Interaction of Two Substrates at the Active Site of Cytochrome P450 3A4. *J. Biol. Chem.* **2001**, *276* (3), 2256–2262.
- (26) Uchaipichat, V.; Mackenzie, P. I.; Guo, X.-H.; Gardner-Stephen, D.; Galetin, A.; Houston, J. B.; Miners, J. O. Human Glucuronosyltransferase: Isoform Selectivity and Kinetics of 4-Methylumbelliferone and 1-Naphthol Glucuronidation, Effects of Organic Solvents, and Inhibition by Diclofenac and Probenecid. *Drug Metab. Dispos.* **2004**, *32* (4), 413–423.
- (27) Houston, J. B.; Kenworthy, K. E. In Vitro-In Vivo Scaling of CYP Kinetic Data Not Consistent with the Classical Michaelis-Menten Model. *Drug Metab. Dispos.* **2000**, *28* (3), 246–254.
- (28) Cuoghi, A.; Caiazzo, M.; Bellei, E.; Monari, E.; Bergamini, S.; Palladino, G.; Ozben, T.; Tomasi, A. Quantification of P-Cresol Sulfate in Human Plasma by Selected Reaction Monitoring. *Anal. Bioanal. Chem.* **2012**, *404* (6–7), 2097–2104.
- (29) Rong, Y.; Kiang, T. K. L. Characterization of Human Sulfotransferases Catalyzing the Formation of P-Cresol Sulfate and Identification of Mefenamic Acid as a Potent Metabolism Inhibitor and Potential Therapeutic Agent for Detoxification. *Toxicol. Appl. Pharmacol.* **2021**, *425*, 115553.
- (30) Gamage, N.; Barnett, A.; Hempel, N.; Duggleby, R. G.; Windmill, K. F.; Martin, J. L.; McManus, M. E. Human Sulfotransferases and Their Role in Chemical Metabolism. *Toxicol. Sci.* **2006**, *90* (1), 5–22.
- (31) Al-Dajani, A. R.; Hou, Q. K.; Kiang, T. K. L. Liquid Chromatography-Mass Spectrometry Analytical Methods for the Quantitation of p-Cresol Sulfate and Indoxyl Sulfate in Human Matrices: Biological Applications and Diagnostic Potentials. *Pharmaceutics* **2024**, *16* (6), 743.
- (32) Peng, B.; Zhao, H.; Keerthisinghe, T. P.; Yu, Y.; Chen, D.; Huang, Y.; Fang, M. Gut Microbial Metabolite P-Cresol Alters Biotransformation of Bisphenol A: Enzyme Competition or Gene Induction? *J. Hazard. Mater.* **2022**, *426*, 128093.
- (33) De Loor, H.; Bammens, B.; Evenepoel, P.; De Preter, V.; Verbeke, K. Gas Chromatographic–Mass Spectrometric Analysis for Measurement of p-Cresol and Its Conjugated Metabolites in Uremic and Normal Serum. *Clin. Chem.* **2005**, *51* (8), 1535–1538.
- (34) Bergé-Lefranc, D.; Chaspoul, F.; Calaf, R.; Charpiot, P.; Brunet, P.; Gallice, P. Binding of P-Cresylsulfate and p-Cresol to Human Serum Albumin Studied by Microcalorimetry. *J. Phys. Chem. B* **2010**, *114* (4), 1661–1665.
- (35) Viaene, L.; Annaert, P.; De Loor, H.; Poesen, R.; Evenepoel, P.; Meijers, B. Albumin Is the Main Plasma Binding Protein for Indoxyl Sulfate and p-cresyl Sulfate. *Biopharm. Drug Dispos.* **2013**, *34* (3), 165–175.
- (36) Watanabe, H.; Noguchi, T.; Miyamoto, Y.; Kadowaki, D.; Kotani, S.; Nakajima, M.; Miyamura, S.; Ishima, Y.; Otagiri, M.; Maruyama, T. Interaction between Two Sulfate-Conjugated Uremic Toxins, p-Cresyl Sulfate and Indoxyl Sulfate, during Binding with Human Serum Albumin. *Drug Metab. Dispos.* **2012**, *40* (7), 1423–1428.
- (37) Yi, D.; Monteiro, E. B.; Chambert, S.; Soula, H. A.; Daleprane, J. B.; Soulage, C. O. Determination of the Binding Properties of P-Cresyl Glucuronide to Human Serum Albumin. *Biochimie* **2018**, *150*, 1–7.
- (38) De Smet, R.; Van Kaer, J.; Van Vlem, B.; De Cubber, A.; Brunet, P.; Lameire, N.; Vanholder, R. Toxicity of Free P-Cresol: A Prospective and Cross-Sectional Analysis. *Clin. Chem.* **2003**, *49* (3), 470–478.
- (39) Ikematsu, N.; Kashiwagi, M.; Hara, K.; Waters, B.; Matsusue, A.; Takayama, M.; Kubo, S. Organ Distribution of Endogenous p-Cresol in Hemodialysis Patients. *J. Med. Invest.* **2019**, *66* (1.2), 81–85.
- (40) Poesen, R.; Evenepoel, P.; De Loor, H.; Kuypers, D.; Augustijns, P.; Meijers, B. Metabolism, Protein Binding, and Renal Clearance of Microbiota-Derived p-Cresol in Patients with CKD. *Clin. J. Am. Soc. Nephrol.* **2016**, *11* (7), 1136–1144.

- (41) Zhu, S.; Rong, Y.; Kiang, T. K. L. Effects of P-Cresol on Oxidative Stress, Glutathione Depletion, and Necrosis in HepaRG Cells: Comparisons to Other Uremic Toxins and the Role of p-Cresol Glucuronide Formation. *Pharmaceutics* **2021**, *13* (6), 857.
- (42) Chinnappa, S.; Tu, Y.-K.; Yeh, Y. C.; Glorieux, G.; Vanholder, R.; Mooney, A. Association between Protein-Bound Uremic Toxins and Asymptomatic Cardiac Dysfunction in Patients with Chronic Kidney Disease. *Toxins* **2018**, *10* (12), 520.
- (43) Liabeuf, S.; Glorieux, G.; Lenglet, A.; Diouf, M.; Schepers, E.; Desjardins, L.; Choukroun, G.; Vanholder, R.; Massy, Z. A. on behalf of the European Uremic Toxin (EUTox) Work Group. Does P-Cresylglucuronide Have the Same Impact on Mortality as Other Protein-Bound Uremic Toxins? *PLoS One* **2013**, *8* (6), No. e67168.
- (44) Meert, N.; Schepers, E.; Glorieux, G.; Van Landschoot, M.; Goeman, J. L.; Waterloos, M.-A.; Dhondt, A.; Van Der Eycken, J.; Vanholder, R. Novel Method for Simultaneous Determination of P-Cresylsulphate and p-Cresylglucuronide: Clinical Data and Pathophysiological Implications. *Nephrol., Dial., Transplant.* **2012**, *27* (6), 2388–2396.
- (45) Itoh, Y.; Ezawa, A.; Kikuchi, K.; Tsuruta, Y.; Niwa, T. Protein-Bound Uremic Toxins in Hemodialysis Patients Measured by Liquid Chromatography/Tandem Mass Spectrometry and Their Effects on Endothelial ROS Production. *Anal. Bioanal. Chem.* **2012**, *403* (7), 1841–1850.
- (46) Corradi, V.; Caprara, C.; Barzon, E.; Mattarollo, C.; Zanetti, F.; Ferrari, F.; Husain-Syed, F.; Giavarina, D.; Ronco, C.; Zanella, M. A Possible Role of P-Cresyl Sulfate and Indoxyl Sulfate as Biomarkers in the Prediction of Renal Function According to the GFR (G) Categories. *Integr. Renephrol. Androl.* **2024**, *11* (3), No. e24-00002.
- (47) Lin, C.-J.; Chen, H.-H.; Pan, C.-F.; Chuang, C.-K.; Wang, T.-J.; Sun, F.-J.; Wu, C.-J. P-Cresylsulfate and Indoxyl Sulfate Level at Different Stages of Chronic Kidney Disease. *J. Clin. Lab. Anal.* **2011**, *25* (3), 191–197.
- (48) Vanholder, R.; Bammens, B.; De Loor, H.; Glorieux, G.; Meijers, B.; Schepers, E.; Massy, Z.; Evenepoel, P. Warning: The Unfortunate End of p-Cresol as a Uraemic Toxin. *Nephrol., Dial., Transplant.* **2011**, *26* (5), 1464–1467.
- (49) King, R. A.; May, B. L.; Davies, D. A.; Bird, A. R. Measurement of Phenol and P-Cresol in Urine and Feces Using Vacuum Microdistillation and High-Performance Liquid Chromatography. *Anal. Biochem.* **2009**, *384* (1), 27–33.
- (50) Renwick, A. G.; Thakrar, A.; Lawrie, C. A.; George, C. F. Microbial Amino Acid Metabolites and Bladder Cancer: No Evidence of Promoting Activity in Man. *Hum. Toxicol.* **1988**, *7* (3), 267–272.
- (51) Miyamoto, Y.; Watanabe, H.; Noguchi, T.; Kotani, S.; Nakajima, M.; Kadowaki, D.; Otagiri, M.; Maruyama, T. Organic Anion Transporters Play an Important Role in the Uptake of P-Cresyl Sulfate, a Uremic Toxin, in the Kidney. *Nephrol., Dial., Transplant.* **2011**, *26* (8), 2498–2502.
- (52) Persico, A. M.; Napolioni, V. Urinary P-Cresol in Autism Spectrum Disorder. *Neurotoxicol. Teratol.* **2013**, *36*, 82–90.
- (53) Candelieri, F.; Simone, M.; Leonardi, A.; Rossi, M.; Amaretti, A.; Raimondi, S. Indole and P-Cresol in Feces of Healthy Subjects: Concentration, Kinetics, and Correlation with Microbiome. *Front. Mol. Med.* **2022**, *2*, 959189.
- (54) Kang, D.-W.; Adams, J. B.; Vargason, T.; Santiago, M.; Hahn, J.; Krajmalnik-Brown, R. Distinct Fecal and Plasma Metabolites in Children with Autism Spectrum Disorders and Their Modulation after Microbiota Transfer Therapy. *mSphere* **2020**, *5* (5), No. e00314-20.
- (55) Flynn, C. K.; Adams, J. B.; Krajmalnik-Brown, R.; Khoruts, A.; Sadowsky, M. J.; Nirmalkar, K.; Takyi, E.; Whiteley, P. Review of Elevated Para-Cresol in Autism and Possible Impact on Symptoms. *Int. J. Mol. Sci.* **2025**, *26* (4), 1513.
- (56) Sankowski, B.; Książarczyk, K.; Raćkowska, E.; Szlufik, S.; Koziorowski, D.; Giebultowicz, J. Higher Cerebrospinal Fluid to Plasma Ratio of P-Cresol Sulfate and Indoxyl Sulfate in Patients with Parkinson's Disease. *Clin. Chim. Acta* **2020**, *501*, 165–173.
- (57) Laudani, S.; Torrisi, S. A.; Alboni, S.; Bastiaanssen, T. F. S.; Benatti, C.; Rivi, V.; Moloney, R. D.; Fuochi, V.; Furneri, P. M.; Drago, F.; Salomone, S.; Tascadda, F.; Cryan, J. F.; Leggio, G. M. Gut Microbiota Alterations Promote Traumatic Stress Susceptibility Associated with P-Cresol-Induced Dopaminergic Dysfunctions. *Brain, Behav., Immun.* **2023**, *107*, 385–396.
- (58) Denman, C. R.; Park, S. M.; Jo, J. Gut-Brain Axis: Gut Dysbiosis and Psychiatric Disorders in Alzheimer's and Parkinson's Disease. *Front. Neurosci.* **2023**, *17*, 1268419.
- (59) Watanabe, H.; Miyamoto, Y.; Honda, D.; Tanaka, H.; Wu, Q.; Endo, M.; Noguchi, T.; Kadowaki, D.; Ishima, Y.; Kotani, S.; Nakajima, M.; Kataoka, K.; Kim-Mitsuyama, S.; Tanaka, M.; Fukagawa, M.; Otagiri, M.; Maruyama, T. P-Cresyl Sulfate Causes Renal Tubular Cell Damage by Inducing Oxidative Stress by Activation of NADPH Oxidase. *Kidney Int.* **2013**, *83* (4), 582–592.
- (60) Bermudez-Martin, P.; Becker, J. A. J.; Caramello, N.; Fernandez, S. P.; Costa-Campos, R.; Canaguier, J.; Barbosa, S.; Martinez-Gili, L.; Myridakis, A.; Dumas, M.-E.; Bruneau, A.; Cherbuy, C.; Langella, P.; Callebert, J.; Launay, J.-M.; Chabry, J.; Barik, J.; Le Merrer, J.; Glaichenhaus, N.; Davidovic, L. The Microbial Metabolite P-Cresol Induces Autistic-like Behaviors in Mice by Remodeling the Gut Microbiota. *Microbiome* **2021**, *9* (1), 157.
- (61) Blachier, F.; Andriamihaja, M. Effects of the L-Tyrosine-Derived Bacterial Metabolite p-Cresol on Colonic and Peripheral Cells. *Amino Acids* **2022**, *54* (3), 325–338.
- (62) Sun, C.-Y.; Li, J.-R.; Wang, Y.-Y.; Lin, S.-Y.; Ou, Y.-C.; Lin, C.-J.; Wang, J.-D.; Liao, S.-L.; Chen, C.-J. P-Cresol Sulfate Caused Behavior Disorders and Neurodegeneration in Mice with Unilateral Nephrectomy Involving Oxidative Stress and Neuroinflammation. *Int. J. Mol. Sci.* **2020**, *21* (18), 6687.
- (63) Shaw, W. Inhibition of the Beta-Oxidation Pathway of Fatty Acids and Dopamine-Beta-Hydroxylase by Phenyl Derivatives of Short-Chain Fatty Acids from Gastrointestinal Clostridia Bacteria Is a (the) Major Cause of Autism. *Integr. Med.* **2023**, *22* (2), 18–25.
- (64) Shah, S. N.; Knausenberger, T. B.-A.; Pontifex, M. G.; Connell, E.; Le Gall, G.; Hardy, T. A. J.; Randall, D. W.; McCafferty, K.; Yaqoob, M. M.; Solito, E.; Müller, M.; Stachulski, A. V.; Glen, R. C.; Vauzour, D.; Hoyles, L.; McArthur, S. Cerebrovascular Damage Caused by the Gut Microbe/Host Co-Metabolite p-Cresol Sulfate Is Prevented by Blockade of the EGF Receptor. *Gut Microbes* **2024**, *16* (1), 2431651.
- (65) Mankhong, S.; Den-Udom, T.; Tanawattanasuntorn, T.; Suriyun, T.; Muta, K.; Kitiyakara, C.; Ketsawatsomkron, P. The Microbial Metabolite P-Cresol Compromises the Vascular Barrier and Induces Endothelial Cytotoxicity and Inflammation in a 3D Human Vessel-on-a-Chip. *Sci. Rep.* **2024**, *14* (1), 18553.
- (66) Dou, L.; Anfosso, F.; Sabatier, F.; Moal, V.; Glorieux, G.; De Smet, R.; Vanholder, R.; Dignat-George, F.; Sampol, J.; Berland, Y.; Brunet, P.; Cerini, C. P-Cresol a Uremic Retention Solute, Alters the Endothelial Barrier Function in Vitro. *Thromb. Haemostasis* **2004**, *92* (07), 140–150.
- (67) Takata, F.; Nakagawa, S.; Matsumoto, J.; Dohgu, S. Blood-Brain Barrier Dysfunction Amplifies the Development of Neuroinflammation: Understanding of Cellular Events in Brain Microvascular Endothelial Cells for Prevention and Treatment of BBB Dysfunction. *Front. Cell. Neurosci.* **2021**, *15*, 661838.
- (68) Renaldi, R.; Wiguna, T.; Persico, A. M.; Tanra, A. J. p-Cresol and p-Cresyl Sulphate Boost Oxidative Stress: A Systematic Review of Recent Evidence. *Basic Clin. Pharmacol. Toxicol.* **2025**, *137* (1), No. e70065.
- (69) Stafim Da Cunha, R.; Gregório, P. C.; Maciel, R. A. P.; Favretto, G.; Franco, C. R. C.; Gonçalves, J. P.; De Azevedo, M. L. V.; Pecoits-Filho, R.; Stighen, A. E. M. Uremic Toxins Activate CREB/ATF1 in Endothelial Cells Related to Chronic Kidney Disease. *Biochem. Pharmacol.* **2022**, *198*, 114984.
- (70) Koppe, L.; Croze, M. L.; Monteiro, E. B.; Benoit, B.; Bres, E.; Guebre-Egziabher, F.; Daleprane, J. B.; Fouque, D.; Soulage, C. O. The Protein-Bound Uremic Toxin p-Cresyl-Sulfate Promotes Intra-

cellular ROS Production and Lipid Peroxidation in 3T3-L1 Adipose Cells. *Biochimie* **2021**, *189*, 137–143.

(71) Dustin, C. M.; Shiva, S. S.; Vazquez, A.; Saeed, A.; Pascoal, T.; Cifuentes-Pagano, E.; Pagano, P. J. NOX2 in Alzheimer's and Parkinson's Disease. *Redox Biol.* **2024**, *78*, 103433.

(72) Simpson, D. S. A.; Oliver, P. L. ROS Generation in Microglia: Understanding Oxidative Stress and Inflammation in Neurodegenerative Disease. *Antioxidants* **2020**, *9* (8), 743.

(73) Tarafdar, A.; Pula, G. The Role of NADPH Oxidases and Oxidative Stress in Neurodegenerative Disorders. *Int. J. Mol. Sci.* **2018**, *19* (12), 3824.

(74) Yan, Z.; Zhong, H. M.; Maher, N.; Torres, R.; Leo, G. C.; Caldwell, G. W.; Huebert, N. Bioactivation of 4-Methylphenol (p-Cresol) via Cytochrome p450-Mediated Aromatic Oxidation in Human Liver Microsomes. *Drug Metab. Dispos.* **2005**, *33* (12), 1867–1876.

(75) DeWolf, W. E.; Carr, S. A.; Varrichio, A.; Goodhart, P. J.; Mentzer, M. A.; Roberts, G. D.; Southan, C.; Dolle, R. E.; Kruse, L. I. Inactivation of Dopamine- β -Hydroxylase by p-Cresol: Isolation and Characterization of Covalently Modified Active Site Peptides. *Biochemistry* **1988**, *27* (26), 9093–9101.

(76) Southan, C.; DeWolf, W. E.; Kruse, L. I. Inactivation of Dopamine β -Hydroxylase by p-Cresol: Evidence for a Second, Minor Site of Covalent Modification at Tyrosine 357. *Biochim. Biophys. Acta, Protein Struct. Mol. Enzymol.* **1990**, *1037* (2), 256–258.

(77) Juárez Olguín, H.; Calderón Guzmán, D.; Hernández García, E.; Barragán Mejía, G. The Role of Dopamine and Its Dysfunction as a Consequence of Oxidative Stress. *Oxid. Med. Cell. Longevity* **2016**, *2016* (1), 9730467.

(78) Muñoz, P.; Huenchuguala, S.; Paris, I.; Segura-Aguilar, J. Dopamine Oxidation and Autophagy. *Parkinson's Dis.* **2012**, *2012*, 1–13.

(79) Forman, H. J.; Zhang, H.; Rinna, A. Glutathione: Overview of Its Protective Roles, Measurement, and Biosynthesis. *Mol. Aspects Med.* **2009**, *30* (1–2), 1–12.

(80) Olufunmilayo, E. O.; Gerke-Duncan, M. B.; Holsinger, R. M. D. Oxidative Stress and Antioxidants in Neurodegenerative Disorders. *Antioxidants* **2023**, *12* (2), 517.

(81) Favretto, G.; Da Cunha, R. S.; Flores Santos, A.; Leitolis, A.; Schiefer, E. M.; Gregório, P. C.; Franco, C. R. C.; Massy, Z.; Dalboni, M. A.; Stinghen, A. E. M. Uremic Endothelial-Derived Extracellular Vesicles: Mechanisms of Formation and Their Role in Cell Adhesion, Cell Migration, Inflammation, and Oxidative Stress. *Toxicol. Lett.* **2021**, *347*, 12–22.

(82) Guo, C.; Sun, L.; Chen, X.; Zhang, D. Oxidative Stress, Mitochondrial Damage and Neurodegenerative Diseases. *Neural Regener. Res.* **2013**, *8* (21), 2003–2014.

(83) Koziel, R.; Pircher, H.; Kratochwil, M.; Lener, B.; Hermann, M.; Dencher, N. A.; Jansen-Dürr, P. Mitochondrial Respiratory Chain Complex I Is Inactivated by NADPH Oxidase Nox4. *Biochem. J.* **2013**, *452* (2), 231–239.

(84) Sun, C.-Y.; Cheng, M.-L.; Pan, H.-C.; Lee, J.-H.; Lee, C.-C. Protein-Bound Uremic Toxins Impaired Mitochondrial Dynamics and Functions. *Oncotarget* **2017**, *8* (44), 77722–77733.

(85) Mutsaers, H. A. M.; Wilmer, M. J. G.; Reijnders, D.; Jansen, J.; Van Den Broek, P. H. H.; Forkink, M.; Schepers, E.; Glorieux, G.; Vanholder, R.; Van Den Heuvel, L. P.; Hoenderop, J. G.; Masereeuw, R. Uremic Toxins Inhibit Renal Metabolic Capacity through Interference with Glucuronidation and Mitochondrial Respiration. *Biochim. Biophys. Acta, Mol. Basis Dis.* **2013**, *1832* (1), 142–150.

(86) Yoon, Y. M.; Han, Y.-S.; Yun, C. W.; Lee, J. H.; Kim, R.; Lee, S. H. Pioglitazone Protects Mesenchymal Stem Cells against P-Cresol-Induced Mitochondrial Dysfunction via Up-Regulation of PINK-1. *Int. J. Mol. Sci.* **2018**, *19* (10), 2898.

(87) Huang, T.; Yip, H.; Sun, C.; Chen, Y.; Yang, C.; Lee, F. P-cresyl Sulfate Causes Mitochondrial Hyperfusion in H9C2 Cardiomyoblasts. *J. Cell. Mol. Med.* **2020**, *24* (15), 8379–8390.

(88) Gigi, T.; Elene, Z.; Tamar, B.; Lali, S.; Davit, D.; Zaqaria, N.; Tamar, L.; David, M. Gut Neurotoxin P-Cresol Induces Differential

Expression of GLUN2B and GLUN2A Subunits of the NMDA Receptor in the Hippocampus and Nucleus Accumbens in Healthy and Audiogenic Seizure-Prone Rats. *AIMS Neurosci.* **2020**, *7* (1), 30–42.

(89) Mayr, B.; Montminy, M. Transcriptional Regulation by the Phosphorylation-Dependent Factor CREB. *Nat. Rev. Mol. Cell Biol.* **2001**, *2* (8), 599–609.

(90) Gacias, M.; Gaspari, S.; Santos, P.-M. G.; Tamburini, S.; Andrade, M.; Zhang, F.; Shen, N.; Tolstikov, V.; Kiebish, M. A.; Dupree, J. L.; Zachariou, V.; Clemente, J. C.; Casaccia, P. Microbiota-Driven Transcriptional Changes in Prefrontal Cortex Override Genetic Differences in Social Behavior. *eLife* **2016**, *5*, No. e13442.

(91) Li, L.-C.; Chen, W.-Y.; Chen, J.-B.; Lee, W.-C.; Chang, C.-C.; Tzeng, H.-T.; Huang, C.-C.; Chang, Y.-J.; Yang, J.-L. The AST-120 Recovers Uremic Toxin-Induced Cognitive Deficit via NLRP3 Inflammasome Pathway in Astrocytes and Microglia. *Biomedicines* **2021**, *9* (9), 1252.

(92) Guzmán-Salas, S.; Weber, A.; Malci, A.; Lin, X.; Herrera-Molina, R.; Cerpa, W.; Dorador, C.; Signorelli, J.; Zamorano, P. The Metabolite p-cresol Impairs Dendritic Development, Synaptogenesis, and Synapse Function in Hippocampal Neurons: Implications for Autism Spectrum Disorder. *J. Neurochem.* **2022**, *161* (4), 335–349.

(93) Ntranos, A.; Park, H.-J.; Wentling, M.; Tolstikov, V.; Amatruda, M.; Inbar, B.; Kim-Schulze, S.; Frazier, C.; Button, J.; Kiebish, M. A.; Lublin, F.; Edwards, K.; Casaccia, P. Bacterial Neurotoxic Metabolites in Multiple Sclerosis Cerebrospinal Fluid and Plasma. *Brain* **2022**, *145* (2), 569–583.

(94) Vinithakumari, A. A.; Padhi, P.; Hernandez, B.; Lin, S. J.-H.; Dunkerson-Kurzhumov, A.; Showman, L.; Breitzman, M.; Stokes, C.; Sulaiman, Y.; Tangudu, C.; Kuttappan, D. A.; Muyyarikkandy, M. S.; Willette, A. A.; Phillips, G. J.; Anantharam, V.; Perera, A.; Sponseller, B. A.; Kanthasamy, A.; Mooyottu, S. Clostridioides Difficile Infection Dysregulates Brain Dopamine Metabolism. *Microbiol. Spectrum* **2022**, *10* (2), No. e00073-22.

(95) Gevi, F.; Belardo, A.; Zolla, L. A Metabolomics Approach to Investigate Urine Levels of Neurotransmitters and Related Metabolites in Autistic Children. *Biochim. Biophys. Acta, Mol. Basis Dis.* **2020**, *1866* (10), 165859.

(96) Pascucci, T.; Colamartino, M.; Fiori, E.; Sacco, R.; Coviello, A.; Ventura, R.; Puglisi-Allegra, S.; Turriziani, L.; Persico, A. M. P-Cresol Alters Brain Dopamine Metabolism and Exacerbates Autism-Like Behaviors in the BTBR Mouse. *Brain Sci.* **2020**, *10* (4), 233.

(97) Politis, M.; Niccolini, F. Serotonin in Parkinson's Disease. *Behav. Brain Res.* **2015**, *277*, 136–145.

(98) Aaldijk, E.; Vermeiren, Y. The Role of Serotonin within the Microbiota-Gut-Brain Axis in the Development of Alzheimer's Disease: A Narrative Review. *Ageing Res. Rev.* **2022**, *75*, 101556.

(99) Correia, A. S.; Cardoso, A.; Vale, N. BDNF Unveiled: Exploring Its Role in Major Depression Disorder Serotonergic Imbalance and Associated Stress Conditions. *Pharmacaceutics* **2023**, *15* (8), 2081.

(100) Ibrahim, A. M.; Chauhan, L.; Bhardwaj, A.; Sharma, A.; Fayaz, F.; Kumar, B.; Alhashmi, M.; AlHajri, N.; Alam, M. S.; Potttoo, F. H. Brain-Derived Neurotrophic Factor in Neurodegenerative Disorders. *Biomedicines* **2022**, *10* (5), 1143.

(101) Tevzadze, G.; Barbakadze, T.; Kvergelidze, E.; Zhuravliova, E.; Shanshiashvili, L.; Mikeladze, D. Gut Neurotoxin P-Cresol Induces Brain-Derived Neurotrophic Factor Secretion and Increases the Expression of Neurofilament Subunits in PC-12 Cells. *AIMS Neurosci.* **2021**, *9* (1), 12–23.

(102) Calderón-Guzmán, D.; Hernández-Islas, J. L.; Espitia Vázquez, I. R.; Barragán-Mejía, G.; Hernández-García, E.; Del Angel, D. S.; Juárez-Olguín, H. Effect of Toluene and Cresols on Na⁺,K⁺-ATPase, and Serotonin in Rat Brain. *Regul. Toxicol. Pharmacol.* **2005**, *41* (1), 1–5.

(103) Fu, H.-Y.; Xu, J.-M.; Ai, X.; Dang, F.-T.; Tan, X.; Yu, H.-Y.; Feng, J.; Yang, W.-X.; Ma, H.-T.; Tu, R.-F.; Gupta, A. K.; Manandhar, L. K.; Bao, W.-M.; Tang, Y.-M. The Clostridium Metabolite P-Cresol Sulfate Relieves Inflammation of Primary Biliary Cholangitis by Regulating Kupffer Cells. *Cells* **2022**, *11* (23), 3782.

- (104) Shiba, T.; Makino, I.; Kawakami, K.; Kato, I.; Kobayashi, T.; Kaneko, K. P-Cresyl Sulfate Suppresses Lipopolysaccharide-Induced Anti-Bacterial Immune Responses in Murine Macrophages in Vitro. *Toxicol. Lett.* **2016**, *245*, 24–30.
- (105) Shiba, T.; Kawakami, K.; Sasaki, T.; Makino, I.; Kato, I.; Kobayashi, T.; Uchida, K.; Kaneko, K. Effects of Intestinal Bacteria-Derived p-Cresyl Sulfate on Th1-Type Immune Response in Vivo and in Vitro. *Toxicol. Appl. Pharmacol.* **2014**, *274* (2), 191–199.
- (106) Zheng, Y.; Prince, N. Z.; Peralta Marzal, L. N.; Ahmed, S.; Garssen, J.; Perez Pardo, P.; Kraneveld, A. D. The Autism Spectrum Disorder-Associated Bacterial Metabolite p-Cresol Derails the Neuro-immune Response of Microglial Cells Partially via Reduction of ADAM17 and ADAM10. *Int. J. Mol. Sci.* **2022**, *23* (19), 11013.
- (107) London, J. A.; Wang, E. C. S.; Barsukov, I. L.; Yates, E. A.; Stachulski, A. V. Synthesis and Toxicity Profile in 293 Human Embryonic Kidney Cells of the β d-Glucuronide Derivatives of Ortho-, Meta- and Para-Cresol. *Carbohydr. Res.* **2021**, 499, 108225.
- (108) Weigand, K. M.; Schirris, T. J. J.; Houweling, M.; Van Den Heuvel, J. J. M. W.; Koenderink, J. B.; Dankers, A. C. A.; Russel, F. G. M.; Greupink, R. Uremic Solutes Modulate Hepatic Bile Acid Handling and Induce Mitochondrial Toxicity. *Toxicol. In Vitro* **2019**, *56*, 52–61.
- (109) Stachulski, A. V.; Knausenberger, T. B.-A.; Shah, S. N.; Hoyles, L.; McArthur, S. A Host–Gut Microbial Amino Acid Co-Metabolite, p -Cresol Glucuronide, Promotes Blood–Brain Barrier Integrity in Vivo. *Tissue Barriers* **2023**, *11* (1), 2073175.
- (110) Glorieux, G.; Vanholder, R.; Van Biesen, W.; Pletinck, A.; Schepers, E.; Neiryck, N.; Speeckaert, M.; De Bacquer, D.; Verbeke, F. Free p -Cresyl Sulfate Shows the Highest Association with Cardiovascular Outcome in Chronic Kidney Disease. *Nephrol., Dial., Transplant.* **2021**, *36* (6), 998–1005.
- (111) Altieri, L.; Neri, C.; Sacco, R.; Curatolo, P.; Benvenuto, A.; Muratori, F.; Santocchi, E.; Bravaccio, C.; Lenti, C.; Saccani, M.; Rigardetto, R.; Gandione, M.; Urbani, A.; Persico, A. M. Urinary p -Cresol Is Elevated in Small Children with Severe Autism Spectrum Disorder. *Biomarkers* **2011**, *16* (3), 252–260.
- (112) Basra, M.; Miceli, L.; Mundra, V.; Stern-Harbutte, A.; Patel, H.; Haynes, J.; Parmar, M. S. Exploring the Neurotoxic Effects of Microbial Metabolites: A Potential Link between p-Cresol and Autism Spectrum Disorders? *Brain Res.* **2025**, *1850*, 149427.
- (113) Gabriele, S.; Sacco, R.; Cerullo, S.; Neri, C.; Urbani, A.; Tripi, G.; Malvy, J.; Barthelemy, C.; Bonnet-Brihault, F.; Persico, A. M. Urinary p -Cresol Is Elevated in Young French Children with Autism Spectrum Disorder: A Replication Study. *Biomarkers* **2014**, *19* (6), 463–470.
- (114) Osredkar, J.; Baškovič, B. Z. v.; Finderle, P.; Bobrowska-Korczak, B.; Gałarek, P.; Rosiak, A.; Giebultowicz, J.; Vrhovšek, M. J.; Kałużna-Czaplińska, J. Relationship between Excreted Uremic Toxins and Degree of Disorder of Children with ASD. *Int. J. Mol. Sci.* **2023**, *24* (8), 7078.
- (115) Mussap, M.; Siracusano, M.; Noto, A.; Fattuoni, C.; Riccioni, A.; Rajula, H. S. R.; Fanos, V.; Curatolo, P.; Barberini, L.; Mazzone, L. The Urine Metabolome of Young Autistic Children Correlates with Their Clinical Profile Severity. *Metabolites* **2020**, *10* (11), 476.
- (116) Turriziani, L.; Ricciardello, A.; Cucinotta, F.; Bellomo, F.; Turturo, G.; Boncoddio, M.; Mirabelli, S.; Scattoni, M. L.; Rossi, M.; Persico, A. M. Gut Mobilization Improves Behavioral Symptoms and Modulates Urinary P-cresol in Chronically Constipated Autistic Children: A Prospective Study. *Autism Res.* **2022**, *15* (1), 56–69.
- (117) Kandeel, W. A.; Meguid, N. A.; Bjørklund, G.; Eid, E. M.; Farid, M.; Mohamed, S. K.; Wakeel, K. E.; Chirumbolo, S.; Elsaedi, A.; Hammad, D. Y. Impact of Clostridium Bacteria in Children with Autism Spectrum Disorder and Their Anthropometric Measurements. *J. Mol. Neurosci.* **2020**, *70* (6), 897–907.
- (118) Vendrik, K. E. W.; Ooijevaar, R. E.; De Jong, P. R. C.; Laman, J. D.; Van Oosten, B. W.; Van Hilten, J. J.; Ducarmon, Q. R.; Keller, J. J.; Kuijper, E. J.; Contarino, M. F. Fecal Microbiota Transplantation in Neurological Disorders. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 98.
- (119) Li, Q.; Han, Y.; Dy, A. B. C.; Hagerman, R. J. The Gut Microbiota and Autism Spectrum Disorders. *Front. Cell. Neurosci.* **2017**, *11*, 120.
- (120) Shaw, W. Elevated Urinary Glyphosate and Clostridia Metabolites With Altered Dopamine Metabolism in Triplets With Autistic Spectrum Disorder or Suspected Seizure Disorder: A Case Study. *Integr. Med.* **2017**, *16* (1), 50–57.
- (121) Kang, D.-W.; İlhan, Z. E.; Isern, N. G.; Hoyt, D. W.; Howsmen, D. P.; Shaffer, M.; Lozupone, C. A.; Hahn, J.; Adams, J. B.; Krajmalnik-Brown, R. Differences in Fecal Microbial Metabolites and Microbiota of Children with Autism Spectrum Disorders. *Anaerobe* **2018**, *49*, 121–131.
- (122) Serrano-Tomás, M. I.; Contreras-Romero, P.; Parellada, M.; Chaves-Cordero, J.; Zamora, J.; Hengst, M.; Pozo, P.; Del Campo, R.; Guzmán-Salas, S. Recognition of the Microbial Metabolite P-Cresol in Autism Spectrum Disorder: Systematic Review and Meta-Analysis. *Front. Mol. Neurosci.* **2025**, *18*, 1576388.
- (123) Gabriele, S.; Sacco, R.; Altieri, L.; Neri, C.; Urbani, A.; Bravaccio, C.; Riccio, M. P.; Iovene, M. R.; Bombace, F.; De Magistris, L.; Persico, A. M. Slow Intestinal Transit Contributes to Elevate Urinary p -cresol Level in Italian Autistic Children. *Autism Res.* **2016**, *9* (7), 752–759.
- (124) Tevzadze, G.; Oniani, N.; Zhuravliova, E.; Darchia, N.; Eliozishvili, M.; Gogichadze, M.; Lortkipanidze, N.; Oniani, T.; Kakabadze, A.; Kakabadze, Z.; Karalashvili, L.; Kikvidze, Z.; Mikeladze, D. Effects of a Gut Microbiome Toxin, p-Cresol, on the Indices of Social Behavior in Rats. *Neurophysiology* **2018**, *50* (5), 372–377.
- (125) Tevzadze, G.; Nanobashvili, Z.; Zhuravliova, E.; Bilanishvili, I.; Shanshiashvili, L.; Kikvidze, Z.; Mikeladze, D. Effects of a Gut Microbiome Toxin, p-Cresol, on the Susceptibility to Seizures in Rats. *Neurophysiology* **2018**, *50* (6), 424–427.
- (126) Kalia, L. V.; Lang, A. E. Parkinson's Disease. *Lancet* **2015**, *386* (9996), 896–912.
- (127) Shao, Y.; Le, W. Recent Advances and Perspectives of Metabolomics-Based Investigations in Parkinson's Disease. *Mol. Neurodegener.* **2019**, *14* (1), 3.
- (128) Powers, R.; Lei, S.; Anandhan, A.; Marshall, D.; Worley, B.; Cerny, R.; Dodds, E.; Huang, Y.; Panayiotidis, M.; Pappa, A.; Franco, R. Metabolic Investigations of the Molecular Mechanisms Associated with Parkinson's Disease. *Metabolites* **2017**, *7* (2), 22.
- (129) Borghammer, P.; Horsager, J.; Andersen, K.; Van Den Berge, N.; Raunio, A.; Murayama, S.; Parkkinen, L.; Myllykangas, L. Neuropathological Evidence of Body-First vs. Brain-First Lewy Body Disease. *Neurobiol. Dis.* **2021**, *161*, 105557.
- (130) Braak, H.; Rüb, U.; Gai, W. P.; Del Tredici, K. Idiopathic Parkinson's Disease: Possible Routes by Which Vulnerable Neuronal Types May Be Subject to Neuroinvasion by an Unknown Pathogen. *J. Neural Transm.* **2003**, *110* (5), 517–536.
- (131) Liddle, R. A. Parkinson's Disease from the Gut. *Brain Res.* **2018**, *1693*, 201–206.
- (132) Savica, R.; Carlin, J. M.; Grossardt, B. R.; Bower, J. H.; Ahlskog, J. E.; Maraganore, D. M.; Bharucha, A. E.; Rocca, W. A. Medical Records Documentation of Constipation Preceding Parkinson Disease: A Case-Control Study. *Neurology* **2009**, *73* (21), 1752–1758.
- (133) Barichella, M.; Severgnini, M.; Cilia, R.; Cassani, E.; Bolliri, C.; Caronni, S.; Ferri, V.; Canello, R.; Ceccarani, C.; Faierman, S.; Pinelli, G.; De Bellis, G.; Zecca, L.; Cereda, E.; Consolandi, C.; Pezzoli, G. Unraveling Gut Microbiota in Parkinson's Disease and Atypical Parkinsonism. *Mov. Disord.* **2019**, *34* (3), 396–405.
- (134) Boertien, J. M.; Pereira, P. A. B.; Aho, V. T. E.; Scheperjans, F. Increasing Comparability and Utility of Gut Microbiome Studies in Parkinson's Disease: A Systematic Review. *J. Parkinson's Dis.* **2019**, *9* (s2), S297–S312.
- (135) Hill-Burns, E. M.; Debelius, J. W.; Morton, J. T.; Wissemann, W. T.; Lewis, M. R.; Wallen, Z. D.; Peddada, S. D.; Factor, S. A.; Molloy, E.; Zabetian, C. P.; Knight, R.; Payami, H. Parkinson's Disease

and Parkinson's Disease Medications Have Distinct Signatures of the Gut Microbiome. *Mov. Disord.* **2017**, *32* (5), 739–749.

(136) Bedarf, J. R.; Hildebrand, F.; Coelho, L. P.; Sunagawa, S.; Bahram, M.; Goeser, F.; Bork, P.; Wüllner, U. Functional Implications of Microbial and Viral Gut Metagenome Changes in Early Stage L-DOPA-Naïve Parkinson's Disease Patients. *Genome Med.* **2017**, *9* (1), 39.

(137) Keshavarzian, A.; Green, S. J.; Engen, P. A.; Voigt, R. M.; Naqib, A.; Forsyth, C. B.; Mutlu, E.; Shannon, K. M. Colonic Bacterial Composition in Parkinson's Disease. *Mov. Disord.* **2015**, *30* (10), 1351–1360.

(138) Scheperjans, F.; Aho, V.; Pereira, P. A. B.; Koskinen, K.; Paulin, L.; Pekkonen, E.; Haapaniemi, E.; Kaakkola, S.; Eerola-Rautio, J.; Pohja, M.; Kinnunen, E.; Murros, K.; Auvinen, P. Gut Microbiota Are Related to Parkinson's Disease and Clinical Phenotype. *Mov. Disord.* **2015**, *30* (3), 350–358.

(139) Fasano, A.; Visanji, N. P.; Liu, L. W. C.; Lang, A. E.; Pfeiffer, R. F. Gastrointestinal Dysfunction in Parkinson's Disease. *Lancet Neurol.* **2015**, *14* (6), 625–639.

(140) Tran, S. M.-S.; Mohajeri, M. H. The Role of Gut Bacterial Metabolites in Brain Development, Aging and Disease. *Nutrients* **2021**, *13* (3), 732.

(141) Jain, A.; Madkan, S.; Patil, P. The Role of Gut Microbiota in Neurodegenerative Diseases: Current Insights and Therapeutic Implications. *Cureus* **2023**, *15*, No. e47861.

(142) Philip Mani, A.; Balasubramanian, B.; Mali, L. A.; Joseph, K. S.; Meyyazhagan, A.; Pappuswamy, M.; Joseph, B. V. The Role of the Gut Microbiota in Neurodegenerative Diseases. *Microbiol. Res.* **2024**, *15* (2), 489–507.

(143) Romano, S.; Savva, G. M.; Bedarf, J. R.; Charles, I. G.; Hildebrand, F.; Narbad, A. Meta-Analysis of the Parkinson's Disease Gut Microbiome Suggests Alterations Linked to Intestinal Inflammation. *npj Parkinson's Dis.* **2021**, *7* (1), 27.

(144) Clairembault, T.; Leclair-Visonneau, L.; Coron, E.; Bourreille, A.; Le Dily, S.; Vavasseur, F.; Heymann, M.-F.; Neunlist, M.; Derkinderen, P. Structural Alterations of the Intestinal Epithelial Barrier in Parkinson's Disease. *Acta Neuropathol. Commun.* **2015**, *3* (1), 12.

(145) Dorsey, E. R.; Bloem, B. R. Parkinson's Disease Is Predominantly an Environmental Disease. *J. Parkinson's Dis.* **2024**, *14* (3), 451–465.

(146) Yuan, X.; Tian, Y.; Liu, C.; Zhang, Z. Environmental Factors in Parkinson's Disease: New Insights into the Molecular Mechanisms. *Toxicol. Lett.* **2022**, *356*, 1–10.

(147) Rehman, K.; Irshad, K.; Kamal, S.; Imran, I.; Akash, M. S. H. Exposure of Environmental Contaminants and Development of Neurological Disorders. *Crit. Rev. Eukaryotic Gene Expression* **2021**, *31* (2), 35–53.

(148) Adams, J. D. Possible Causes of Parkinson's Disease. *Front. Biosci.-Landmark* **2021**, *26* (8), 387.

(149) Cirstea, M. S.; Yu, A. C.; Golz, E.; Sundvick, K.; Kligler, D.; Radisavljevic, N.; Foulger, L. H.; Mackenzie, M.; Huan, T.; Finlay, B. B.; Appel-Cresswell, S. Microbiota Composition and Metabolism Are Associated With Gut Function in Parkinson's Disease. *Mov. Disord.* **2020**, *35* (7), 1208–1217.

(150) Chen, S.-J.; Wu, Y.-J.; Chen, C.-C.; Wu, Y.-W.; Liou, J.-M.; Wu, M.-S.; Kuo, C.-H.; Lin, C.-H. Plasma Metabolites of Aromatic Amino Acids Associate with Clinical Severity and Gut Microbiota of Parkinson's Disease. *npj Parkinson's Dis.* **2023**, *9* (1), 165.

(151) Paul, K. C.; Zhang, K.; Walker, D. I.; Sinsheimer, J.; Yu, Y.; Kusters, C.; Del Rosario, I.; Folle, A. D.; Keener, A. M.; Bronstein, J.; Jones, D. P.; Ritz, B. Untargeted Serum Metabolomics Reveals Novel Metabolite Associations and Disruptions in Amino Acid and Lipid Metabolism in Parkinson's Disease. *Mol. Neurodegener.* **2023**, *18* (1), 100.

(152) Wyczalkowska-Tomasik, A.; Czarkowska-Paczek, B.; Giebultowicz, J.; Wroczynski, P.; Paczek, L. Age-dependent Increase in Serum Levels of Indoxyl Sulphate and P-cresol Sulphate Is Not

Related to Their Precursors: Tryptophan and Tyrosine. *Geriatr. Gerontol. Int.* **2017**, *17* (6), 1022–1026.

(153) Chen, S.-J.; Wu, Y.-J.; Chen, C.-C.; Wu, Y.-W.; Liou, J.-M.; Wu, M.-S.; Kuo, C.-H.; Lin, C.-H. Plasma Metabolites of Aromatic Amino Acids Associate with Clinical Severity and Gut Microbiota of Parkinson's Disease. *npj Parkinson's Dis.* **2023**, *9* (1), 165.

(154) Willkommen, D.; Lucio, M.; Moritz, F.; Forcisi, S.; Kanawati, B.; Smirnov, K. S.; Schroeter, M.; Sigaroudi, A.; Schmitt-Kopplin, P.; Michalke, B. Metabolomic Investigations in Cerebrospinal Fluid of Parkinson's Disease. *PLoS One* **2018**, *13* (12), No. e0208752.

(155) Gray, M. T.; Woulfe, J. M. Striatal Blood–Brain Barrier Permeability in Parkinson's Disease. *J. Cereb. Blood Flow Metab.* **2015**, *35* (5), 747–750.

(156) Lau, K.; Kotzur, R.; Richter, F. Blood–Brain Barrier Alterations and Their Impact on Parkinson's Disease Pathogenesis and Therapy. *Transl. Neurodegener.* **2024**, *13* (1), 37.

(157) Wu, W.; Bush, K. T.; Nigam, S. K. Key Role for the Organic Anion Transporters, OAT1 and OAT3, in the in Vivo Handling of Uremic Toxins and Solutes. *Sci. Rep.* **2017**, *7* (1), 4939.

(158) Urquhart, B. L.; Kim, R. B. Blood–brain Barrier Transporters and Response to CNS-Active Drugs. *Eur. J. Clin. Pharmacol.* **2009**, *65* (11), 1063–1070.

(159) Ohtsuki, S.; Asaba, H.; Takayama, H.; Deguchi, T.; Hosoya, K.; Otogiri, M.; Terasaki, T. Role of Blood–Brain Barrier Organic Anion Transporter 3 (OAT3) in the Efflux of Indoxyl Sulfate, a Uremic Toxin: Its Involvement in Neurotransmitter Metabolite Clearance from the Brain. *J. Neurochem.* **2002**, *83* (1), 57–66.

(160) Hosoya, K.; Tachikawa, M. Roles of Organic Anion/Cation Transporters at the Blood–Brain and Blood–Cerebrospinal Fluid Barriers Involving Uremic Toxins. *Clin. Exp. Nephrol.* **2011**, *15* (4), 478–485.

(161) Lucio, M.; Willkommen, D.; Schroeter, M.; Sigaroudi, A.; Schmitt-Kopplin, P.; Michalke, B. Integrative Metabolomic and Metallomic Analysis in a Case–Control Cohort With Parkinson's Disease. *Front. Aging Neurosci.* **2019**, *11*, 331.

(162) Wang, Y.; Nong, Y.; Zhang, X.; Mai, T.; Cai, J.; Liu, J.; Lai, K. P.; Zhang, Z. Comparative Plasma Metabolomic Analysis to Identify Biomarkers for Lead-Induced Cognitive Impairment. *Chem.-Biol. Interact.* **2022**, *366*, 110143.

(163) Wong-Guerra, M.; Calfio, C.; Maccioni, R. B.; Rojo, L. E. Revisiting the Neuroinflammation Hypothesis in Alzheimer's Disease: A Focus on the Druggability of Current Targets. *Front. Pharmacol.* **2023**, *14*, 1161850.

(164) Thakur, S.; Dhapola, R.; Sarma, P.; Medhi, B.; Reddy, D. H. Neuroinflammation in Alzheimer's Disease: Current Progress in Molecular Signaling and Therapeutics. *Inflammation* **2023**, *46* (1), 1–17.

(165) Long, J. M.; Holtzman, D. M. Alzheimer Disease: An Update on Pathobiology and Treatment Strategies. *Cell* **2019**, *179* (2), 312–339.

(166) Jung, Y. H.; Chae, C. W.; Han, H. J. The Potential Role of Gut Microbiota-Derived Metabolites as Regulators of Metabolic Syndrome-Associated Mitochondrial and Endolysosomal Dysfunction in Alzheimer's Disease. *Exp. Mol. Med.* **2024**, *56* (8), 1691–1702.

(167) Seo, D.; Holtzman, D. M. Current Understanding of the Alzheimer's Disease-Associated Microbiome and Therapeutic Strategies. *Exp. Mol. Med.* **2024**, *56* (1), 86–94.

(168) Chandra, S.; Sisodia, S. S.; Vassar, R. J. The Gut Microbiome in Alzheimer's Disease: What We Know and What Remains to Be Explored. *Mol. Neurodegener.* **2023**, *18* (1), 9.

(169) Minter, M. R.; Zhang, C.; Leone, V.; Ringus, D. L.; Zhang, X.; Oyler-Castrillo, P.; Musch, M. W.; Liao, F.; Ward, J. F.; Holtzman, D. M.; Chang, E. B.; Tanzi, R. E.; Sisodia, S. S. Antibiotic-Induced Perturbations in Gut Microbial Diversity Influences Neuro-Inflammation and Amyloidosis in a Murine Model of Alzheimer's Disease. *Sci. Rep.* **2016**, *6* (1), 30028.

(170) Vogt, N. M.; Kerby, R. L.; Dill-McFarland, K. A.; Harding, S. J.; Merluzzi, A. P.; Johnson, S. C.; Carlsson, C. M.; Asthana, S.; Zetterberg, H.; Blennow, K.; Bendlin, B. B.; Rey, F. E. Gut

- Microbiome Alterations in Alzheimer's Disease. *Sci. Rep.* **2017**, *7* (1), 13537.
- (171) Cattaneo, A.; Cattane, N.; Galluzzi, S.; Provasi, S.; Lopizzo, N.; Festari, C.; Ferrari, C.; Guerra, U. P.; Paghera, B.; Muscio, C.; Bianchetti, A.; Volta, G. D.; Turla, M.; Cotelli, M. S.; Gennuso, M.; Prele, A.; Zanetti, O.; Lussignoli, G.; Mirabile, D.; Bellandi, D.; Gentile, S.; Belotti, G.; Villani, D.; Harach, T.; Bolmont, T.; Padovani, A.; Boccardi, M.; Frisoni, G. B. Association of Brain Amyloidosis with Pro-Inflammatory Gut Bacterial Taxa and Peripheral Inflammation Markers in Cognitively Impaired Elderly. *Neurobiol. Aging* **2017**, *49*, 60–68.
- (172) Mezö, C.; Dokalis, N.; Mossad, O.; Staszewski, O.; Neuber, J.; Yilmaz, B.; Schnepf, D.; De Agüero, M. G.; Ganal-Vonarburg, S. C.; Macpherson, A. J.; Meyer-Luehmann, M.; Staeheli, P.; Blank, T.; Prinz, M.; Erny, D. Different Effects of Constitutive and Induced Microbiota Modulation on Microglia in a Mouse Model of Alzheimer's Disease. *Acta Neuropathol. Commun.* **2020**, *8* (1), 119.
- (173) Harach, T.; Marungruang, N.; Duthilleul, N.; Cheatham, V.; Mc Coy, K. D.; Frisoni, G.; Neher, J. J.; Fák, F.; Jucker, M.; Lasser, T.; Bolmont, T. Reduction of Abeta Amyloid Pathology in APPS1 Transgenic Mice in the Absence of Gut Microbiota. *Sci. Rep.* **2017**, *7* (1), 41802.
- (174) Wang, X.; Sun, G.; Feng, T.; Zhang, J.; Huang, X.; Wang, T.; Xie, Z.; Chu, X.; Yang, J.; Wang, H.; Chang, S.; Gong, Y.; Ruan, L.; Zhang, G.; Yan, S.; Lian, W.; Du, C.; Yang, D.; Zhang, Q.; Lin, F.; Liu, J.; Zhang, H.; Ge, C.; Xiao, S.; Ding, J.; Geng, M. Sodium Oligomannate Therapeutically Remodels Gut Microbiota and Suppresses Gut Bacterial Amino Acids-Shaped Neuroinflammation to Inhibit Alzheimer's Disease Progression. *Cell Res.* **2019**, *29* (10), 787–803.
- (175) Kwon, M. J.; Song, Y. R.; Kim, J.-H.; Kim, J. H.; Kang, H. S.; Lim, H.; Kim, M.-J.; Kim, N. Y.; Hong, S.; Choi, Y.; Min, K.-W.; Choi, H. G.; Kim, E. S. Exploring the Link between Chronic Kidney Disease and Alzheimer's Disease: A Longitudinal Follow-Up Study Using the Korean National Health Screening Cohort. *Biomedicines* **2023**, *11* (6), 1606.
- (176) Zhang, C.-Y.; He, F.-F.; Su, H.; Zhang, C.; Meng, X.-F. Association between Chronic Kidney Disease and Alzheimer's Disease: An Update. *Metab. Brain Dis.* **2020**, *35* (6), 883–894.
- (177) McAdams-DeMarco, M. A.; Bae, S.; Chu, N.; Gross, A. L.; Brown, C. H.; Oh, E.; Rosenberg, P.; Neufeld, K. J.; Varadhan, R.; Albert, M.; Walston, J.; Segev, D. L. Dementia and Alzheimer's Disease among Older Kidney Transplant Recipients. *J. Am. Soc. Nephrol.* **2017**, *28* (5), 1575–1583.
- (178) Lim, Y. J.; Sidor, N. A.; Tonial, N. C.; Che, A.; Urquhart, B. L. Uremic Toxins in the Progression of Chronic Kidney Disease and Cardiovascular Disease: Mechanisms and Therapeutic Targets. *Toxins* **2021**, *13* (2), 142.
- (179) Yu, Y. H.; Im, H.; Park, S.; Song, B.; Park, D.-K.; Kim, D.-S.; Gil, H.-W. AST-120 Protects Cognitive and Emotional Impairment in Chronic Kidney Disease Induced by 5/6 Nephrectomy. *Brain Sci.* **2024**, *14* (11), 1043.
- (180) Liu, W.-C.; Tomino, Y.; Lu, K.-C. Impacts of Indoxyl Sulfate and p-Cresol Sulfate on Chronic Kidney Disease and Mitigating Effects of AST-120. *Toxins* **2018**, *10* (9), 367.
- (181) Bobot, M.; Thomas, L.; Moyon, A.; Fernandez, S.; McKay, N.; Balasse, L.; Garrigue, P.; Brige, P.; Chopinet, S.; Poitevin, S.; Cérimi, C.; Brunet, P.; Dignat-George, F.; Burtey, S.; Guillet, B.; Hache, G. Uremic Toxic Blood-Brain Barrier Disruption Mediated by AhR Activation Leads to Cognitive Impairment during Experimental Renal Dysfunction. *J. Am. Soc. Nephrol.* **2020**, *31* (7), 1509–1521.
- (182) Tang, W.-H.; Wang, C.-P.; Yu, T.-H.; Tai, P.-Y.; Liang, S.-S.; Hung, W.-C.; Wu, C.-C.; Huang, S.-H.; Lee, Y.-J.; Chen, S.-C. Protein-Bounded Uremic Toxin p-Cresylsulfate Induces Vascular Permeability Alternations. *Histochem. Cell Biol.* **2018**, *149* (6), 607–617.
- (183) Edamatsu, T.; Fujieda, A.; Itoh, Y. Phenyl Sulfate Indoxyl Sulfate and p-Cresyl Sulfate Decrease Glutathione Level to Render Cells Vulnerable to Oxidative Stress in Renal Tubular Cells. *PLoS One* **2018**, *13* (2), No. e0193342.
- (184) Azevedo, M. L. V.; Bonan, N. B.; Dias, G.; Brehm, F.; Steiner, T. M.; Souza, W. M.; Stinghen, A. E. M.; Barreto, F. C.; Elifio-Esposito, S.; Pecoito-Filho, R.; Moreno-Amaral, A. N. p-Cresyl sulfate affects the oxidative burst, phagocytosis process, and antigen presentation of monocyte-derived macrophages. *Toxicol. Lett.* **2016**, *263*, 1–5.
- (185) Sim, M. A.; Liao, Y.; Chan, S. P.; Tan, E. S. J.; Kan, C. N.; Chong, J. R.; Chai, Y. L.; Venketasubramanian, N.; Tan, B. Y.; Hilal, S.; Xu, X.; Chen, C. L. H.; Lai, M. K. P. Low Serum Serotonin Is Associated with Functional Decline, Mild Behavioural Impairment and Brain Atrophy in Dementia-Free Subjects. *Brain Commun.* **2024**, *7* (1), fcaf005.
- (186) Gao, L.; Zhang, Y.; Sterling, K.; Song, W. Brain-Derived Neurotrophic Factor in Alzheimer's Disease and Its Pharmaceutical Potential. *Transl. Neurodegener.* **2022**, *11* (1), 4.
- (187) Kalecký, K.; German, D. C.; Montillo, A. A.; Bottiglieri, T. Targeted Metabolomic Analysis in Alzheimer's Disease Plasma and Brain Tissue in Non-Hispanic Whites. *J. Alzheimer's Dis.* **2022**, *86* (4), 1875–1895.
- (188) Gordon, S.; Lee, J. S.; Scott, T. M.; Bhupathiraju, S.; Ordovas, J.; Kelly, R. S.; Bhadelia, R.; Koo, B.-B.; Bigornia, S.; Tucker, K. L.; Palacios, N. Metabolites and MRI-Derived Markers of AD/ADRD Risk in a Puerto Rican Cohort. *Res. Sq.* **2024**, rs.3.rs3941791.
- (189) Saji, N.; Murotani, K.; Hisada, T.; Kunihiro, T.; Tsuduki, T.; Sugimoto, T.; Kimura, A.; Niida, S.; Toba, K.; Sakurai, T. Relationship between Dementia and Gut Microbiome-Associated Metabolites: A Cross-Sectional Study in Japan. *Sci. Rep.* **2020**, *10* (1), 8088.
- (190) Stein, D. J.; McLaughlin, K. A.; Koenen, K. C.; Atwoli, L.; Friedman, M. J.; Hill, E. D.; Maercker, A.; Petukhova, M.; Shahly, V.; Van Ommeren, M.; Alonso, J.; Borges, G.; De Girolamo, G.; De Jonge, P.; Demyttenaere, K.; Florescu, S.; Karam, E. G.; Kawakami, N.; Matschinger, H.; Okoliyski, M.; Posada-Villa, J.; Scott, K. M.; Viana, M. C.; Kessler, R. C. DSM-5 and ICD-11 definitions of posttraumatic stress disorder: investigating “narrow” and “broad” approaches: Research Article: DSM-5 and ICD-11 Definitions of PTSD. *Depression Anxiety* **2014**, *31* (6), 494–505.
- (191) Malan-Muller, S.; Valles-Colomer, M.; Foxx, C. L.; Vieira-Silva, S.; Van Den Heuvel, L. L.; Raes, J.; Seedat, S.; Lowry, C. A.; Hemmings, S. M. J. Exploring the Relationship between the Gut Microbiome and Mental Health Outcomes in a Posttraumatic Stress Disorder Cohort Relative to Trauma-Exposed Controls. *Eur. Neuropsychopharmacol.* **2022**, *56*, 24–38.
- (192) Yehuda, R.; Hoge, C. W.; McFarlane, A. C.; Vermetten, E.; Lanius, R. A.; Nievergelt, C. M.; Hobfoll, S. E.; Koenen, K. C.; Neylan, T. C.; Hyman, S. E. Post-Traumatic Stress Disorder. *Nat. Rev. Dis. Primers* **2015**, *1* (1), 15057.
- (193) Ntranos, A.; Casaccia, P. The Microbiome–Gut–Behavior Axis: Crosstalk Between the Gut Microbiome and Oligodendrocytes Modulates Behavioral Responses. *Neurotherapeutics* **2018**, *15* (1), 31–35.
- (194) Ke, S.; Hartmann, J.; Ressler, K. J.; Liu, Y.-Y.; Koenen, K. C. The Emerging Role of the Gut Microbiome in Posttraumatic Stress Disorder. *Brain, Behav., Immun.* **2023**, *114*, 360–370.
- (195) He, Q.; Wang, W.; Xu, D.; Xiong, Y.; Tao, C.; You, C.; Ma, L.; Ma, J.; Nievergelt, C. M.; Maihofer, A. X.; Klengel, T.; Atkinson, E. G.; Chen, C.-Y.; Choi, K. W.; Coleman, J. R. I.; Dalvie, S.; Duncan, L. E.; Logue, M. W.; Provost, A. C.; Ratanatharathorn, A.; Stein, M. B.; Torres, K.; Aiello, A. E.; Almli, L. M.; Amstadter, A. B.; Andersen, S. B.; Andreassen, O. A.; Arbis, P. A.; Ashley-Koch, A. E.; Austin, S. B.; Avdibegovic, E.; Babić, D.; Bækvad-Hansen, M.; Baker, D. G.; Beckham, J. C.; Bierut, L. J.; Bisson, J. I.; Boks, M. P.; Bolger, E. A.; Børglum, A. D.; Bradley, B.; Brashear, M.; Breen, G.; Bryant, R. A.; Bustamante, A. C.; Bybjerg-Grauholm, J.; Calabrese, J. R.; Caldas-de-Almeida, J. M.; Dale, A. M.; Daly, M. J.; Daskalakis, N. P.; Deckert, J.; Delahanty, D. L.; Dennis, M. F.; Disner, S. G.; Domschke, K.; Dzubur-Kulenovic, A.; Erbes, C. R.; Evans, A.; Farrer, L. A.; Feeny, N.

- C.; Flory, J. D.; Forbes, D.; Franz, C. E.; Galea, S.; Garrett, M. E.; Gelaye, B.; Gelernter, J.; Geuze, E.; Gillespie, C.; Uka, A. G.; Gordon, S. D.; Guffanti, G.; Hammamieh, R.; Harnal, S.; Hauser, M. A.; Heath, A. C.; Hemmings, S. M. J.; Hougaard, D. M.; Jakovljevic, M.; Jett, M.; Johnson, E. O.; Jones, I.; Jovanovic, T.; Qin, X.-J.; Junglen, A. G.; Karstoft, K.-I.; Kaufman, M. L.; Kessler, R. C.; Khan, A.; Kimbrel, N. A.; King, A. P.; Koen, N.; Kranzler, H. R.; Kremen, W. S.; Lawford, B. R.; Lebois, L. A. M.; Lewis, C. E.; Linnstaedt, S. D.; Lori, A.; Lugonja, B.; Luyck, J. J.; Lyons, M. J.; Maples-Keller, J.; Marmar, C.; Martin, A. R.; Martin, N. G.; Maurer, D.; Mavissakalian, M. R.; McFarlane, A.; McGlinchey, R. E.; McLaughlin, K. A.; McLean, S. A.; McLeay, S.; Mehta, D.; Milberg, W. P.; Miller, M. W.; Morey, R. A.; Morris, C. P.; Mors, O.; Mortensen, P. B.; Neale, B. M.; Nelson, E. C.; Nordentoft, M.; Norman, S. B.; O'Donnell, M.; Orcutt, H. K.; Panizzon, M. S.; Peters, E. S.; Peterson, A. L.; Peverill, M.; Pietrzak, R. H.; Polusny, M. A.; Rice, J. P.; Ripke, S.; Risbrough, V. B.; Roberts, A. L.; Rothbaum, A. O.; Rothbaum, B. O.; Roy-Byrne, P.; Ruggiero, K.; Rung, A.; Rutten, B. P. F.; Saccone, N. L.; Sanchez, S. E.; Schijven, D.; Seedat, S.; Seligowski, A. V.; Seng, J. S.; Sheerin, C. M.; Silove, D.; Smith, A. K.; Smoller, J. W.; Solovieff, N.; Sponheim, S. R.; Stein, D. J.; Sumner, J. A.; Teicher, M. H.; Thompson, W. K.; Trapido, E.; Uddin, M.; Ursano, R. J.; Van Den Heuvel, L. L.; Van Hooff, M.; Vermetten, E.; Vinkers, C. H.; Voisey, J.; Wang, Y.; Wang, Z.; Werge, T.; Williams, M. A.; Williamson, D. E.; Winternitz, S.; Wolf, C.; Wolf, E. J.; Wolff, J. D.; Yehuda, R.; Young, K. A.; Young, R. M.; Zhao, H.; Zoellner, L. A.; Liberzon, I.; Ressler, K. J.; Haas, M.; Koenen, K. C. Potential Causal Association between Gut Microbiome and Posttraumatic Stress Disorder. *Transl. Psychiatry* **2024**, *14* (1), 67.
- (196) Zeamer, A. L.; Salive, M.-C.; An, X.; Beaudoin, F. L.; House, S. L.; Stevens, J. S.; Zeng, D.; Neylan, T. C.; Clifford, G. D.; Linnstaedt, S. D.; Rauch, S. L.; Storrow, A. B.; Lewandowski, C.; Musey, P. I.; Hendry, P. L.; Sheikh, S.; Jones, C. W.; Panches, B. E.; Swor, R. A.; Hudak, L. A.; Pascual, J. L.; Seamon, M. J.; Harris, E.; Pearson, C.; Peak, D. A.; Merchant, R. C.; Domeier, R. M.; Rathlev, N. K.; O'Neil, B. J.; Sergot, P.; Sanchez, L. D.; Bruce, S. E.; Kessler, R. C.; Koenen, K. C.; McLean, S. A.; Bucci, V.; Haran, J. P. Association between Microbiome and the Development of Adverse Posttraumatic Neuropsychiatric Sequelae after Traumatic Stress Exposure. *Transl. Psychiatry* **2023**, *13* (1), 354.
- (197) Tanelian, A.; Nankova, B.; Miar, M.; Nahvi, R. J.; Sabban, E. L. Resilience or Susceptibility to Traumatic Stress: Potential Influence of the Microbiome. *Neurobiol. Stress* **2022**, *19*, 100461.
- (198) Kelly, L. S.; Apple, C. G.; Gharaibeh, R.; Pons, E. E.; Thompson, C. W.; Kannan, K. B.; Darden, D. B.; Efron, P. A.; Thomas, R. M.; Mohr, A. M. Stress-Related Changes in the Gut Microbiome after Trauma. *J. Trauma Acute Care Surg.* **2021**, *91* (1), 192–199.
- (199) Loupy, K. M.; Lowry, C. A. Posttraumatic Stress Disorder and the Gut Microbiome. In *The Oxford Handbook of the Microbiome-Gut-Brain Axis*; Burnet, P. W. J., Ed.; Oxford University Press, 2020.
- (200) Petakh, P.; Oksenysh, V.; Kamyshna, I.; Boisak, I.; Lyubomirskaya, K.; Kamyshnyi, O. Exploring the Interplay between Posttraumatic Stress Disorder, Gut Microbiota, and Inflammatory Biomarkers: A Comprehensive Meta-Analysis. *Front. Immunol.* **2024**, *15*, 1349883.
- (201) Leclercq, S.; Ahmed, H.; Amadiou, C.; Petit, G.; Koistinen, V.; Leyrolle, Q.; Poncin, M.; Stärkel, P.; Kok, E.; Karhunen, P. J.; De Timary, P.; Laye, S.; Neyrinck, A. M.; Kärkkäinen, O. K.; Hanhineva, K.; Delzenne, N. Blood Metabolomic Profiling Reveals New Targets in the Management of Psychological Symptoms Associated with Severe Alcohol Use Disorder. *eLife* **2024**, *13*, RP96937.
- (202) Yeh, Y.-C.; Huang, M.-F.; Liang, S.-S.; Hwang, S.-J.; Tsai, J.-C.; Liu, T.-L.; Wu, P.-H.; Yang, Y.-H.; Kuo, K.-C.; Kuo, M.-C.; Chen, C.-S. Indoxyl Sulfate, Not p-Cresyl Sulfate, Is Associated with Cognitive Impairment in Early-Stage Chronic Kidney Disease. *Neurotoxicology* **2016**, *53*, 148–152.
- (203) Lin, Y.-T.; Wu, P.-H.; Liang, S.-S.; Mubanga, M.; Yang, Y.-H.; Hsu, Y.-L.; Kuo, M.-C.; Hwang, S.-J.; Kuo, P.-L. Protein-Bound Uremic Toxins Are Associated with Cognitive Function among Patients Undergoing Maintenance Hemodialysis. *Sci. Rep.* **2019**, *9* (1), 20388.
- (204) Yassin, L. K.; Nakhal, M. M.; Alderei, A.; Almehairbi, A.; Mydeen, A. B.; Akour, A.; Hamad, M. I. K. Exploring the Microbiota-Gut-Brain Axis: Impact on Brain Structure and Function. *Front. Neuroanat.* **2025**, *19*, 1504065.
- (205) Chu, A.; Samman, S.; Galland, B.; Foster, M. Daily Consumption of Lactobacillus Gasseri CP2305 Improves Quality of Sleep in Adults – A Systematic Literature Review and Meta-Analysis. *Clin. Nutr.* **2023**, *42* (8), 1314–1321.
- (206) Watanabe, H.; Sakaguchi, Y.; Sugimoto, R.; Kaneko, K.; Iwata, H.; Kotani, S.; Nakajima, M.; Ishima, Y.; Otagiri, M.; Maruyama, T. Human Organic Anion Transporters Function as a High-Capacity Transporter for p-Cresyl Sulfate, a Uremic Toxin. *Clin. Exp. Nephrol.* **2014**, *18* (5), 814–820.
- (207) Nigam, S. K.; Bush, K. T.; Martovetsky, G.; Ahn, S.-Y.; Liu, H. C.; Richard, E.; Bhatnagar, V.; Wu, W. The Organic Anion Transporter (OAT) Family: A Systems Biology Perspective. *Physiol. Rev.* **2015**, *95* (1), 83–123.
- (208) Parvez, M. M.; Sadighi, A.; Ahn, Y.; Keller, S. F.; Enoru, J. O. Uptake Transporters at the Blood–Brain Barrier and Their Role in Brain Drug Disposition. *Pharmaceutics* **2023**, *15* (10), 2473.
- (209) Huttunen, K. M.; Terasaki, T.; Urtti, A.; Montaser, A. B.; Uchida, Y. Pharmacoproteomics of Brain Barrier Transporters and Substrate Design for the Brain Targeted Drug Delivery. *Pharm. Res.* **2022**, *39* (7), 1363–1392.