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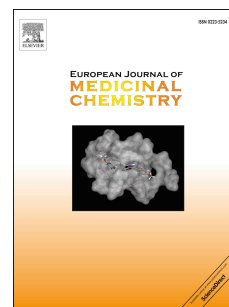
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Synthesis, Biological Evaluation and Molecular Modelling of 1-oxa-4-thiaspiro- and 1,4-dithiaspiro[4.5]decane derivatives as potent and selective 5-HT_{1A} Receptor Agonists

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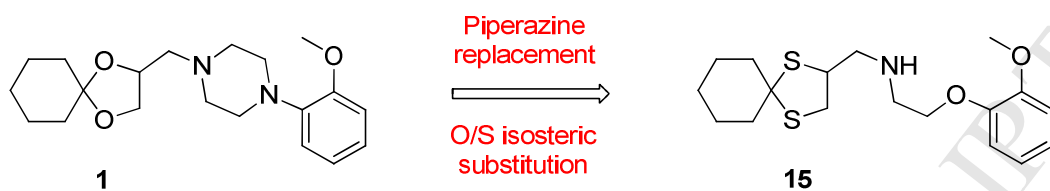
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Key words: 5-HT_{1A} receptor; Agonist; Neuroprotection; BBB penetration; Analgesic activity;

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5-HT_{1A}: pK_i = 8.29, pD₂ = 8.61, E_{max} = 48%

5-HT_{1A}: pK_i = **9.01**, pD₂ = **9.58**, E_{max} = **73%**

ABSTRACT

Recently, 1-(1,4-dioxaspiro[4,5]dec-2-ylmethyl)-4-(2-methoxyphenyl)piperazine (**1**) was reported as a potent 5-HT_{1A}R agonist with a moderate 5-HT_{1A}R selectivity. In an extension of this work a series of derivatives of **1**, obtained by combining different heterocyclic rings with a more flexible amine chain, was synthesized and tested for binding affinity and activity at 5-HT_{1A}R and α_1 adrenoceptors. The results led to the identification of **14** and **15** as novel 5-HT_{1A}R partial agonists, the first being outstanding for selectivity (5-HT_{1A}/ α_{1d} = 80), the latter for potency (pD₂ = 9.58) and efficacy (E_{max} = 74%,). Theoretical studies of ADME properties shows a good profile for the entire series and MDCKII-MDR1 cells permeability data predict a good *BBB* permeability of compound **15**, which possess a promising neuroprotective activity. Furthermore, in mouse formalin test, compound **15** shows a potent antinociceptive activity suggesting a new strategy for pain control.

Introduction

Serotonin (5-hydroxytryptamine, 5-HT) is a relevant neurotransmitter both in the central nervous system and in periphery. It mediates several physiological effects through at least 14 receptor subtypes (5-HT₁₋₇: 5-HT_{1A-F}, 5-HT_{2A-C}, 5-HT₃, 5-HT₄, 5-HT_{5A}, 5-HT₆, 5-HT₇). With the exception of 5-HT₃, they belong to the seven-transmembrane-spanning receptor or the G-protein-coupled receptor (GPCR) family [1,2]. The 5-HT_{1A} receptor (5-HT_{1A}R) was the first subtype to be isolated and completely sequenced [3], its pharmacology has been extensively studied and a number of selective ligands have been discovered (Figure 1). Nowadays this receptor still represents a new attractive target for drug discovery [4]. 5-HT_{1A}R agonists and partial agonists have been initially employed for the treatment of anxiety, depression, and psychosis [5-9]. Moreover, 5-HT_{1A}R agonists have shown neuroprotective properties indicating their utility for the treatment of many neurodegenerative disorders, including Parkinson's disease (PD) and ischemic stroke [10-32]. More recently, it has been shown that 5-HT_{1A}R is involved at multiple level in the regulation of nociception and 5-HT_{1A}R agonists may represent a new approach in pain relief therapy [33-36].

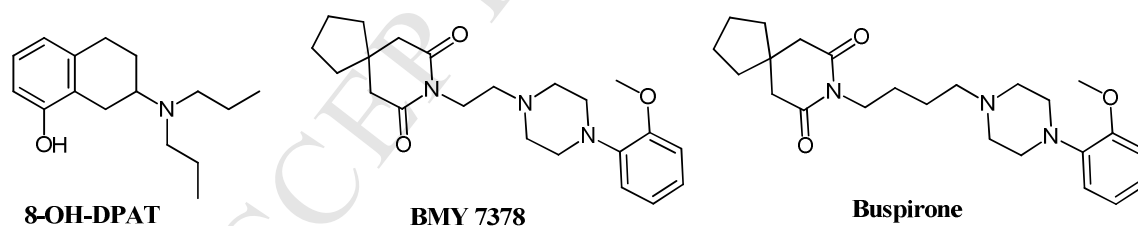


Figure 1. 5-HT_{1A}R selective ligands

Among the 5-HT_{1A}R ligands, N-1-substituted N-4-arylpiperazines (so-called “long-chain arylpiperazines”) have been extensively studied and a generally accepted pharmacophoric model for the recognition of the agonist has been drawn (Figure 2, panel a) [4,37]. Recently, our

research group reported 1-(1,4-dioxaspiro[4.5]dec-2-ylmethyl)-4-(2-methoxyphenyl)piperazine (**1**) as a potent 5-HT_{1A}R partial agonist ($pD_2 = 8.61$) with a moderate selectivity with respect to α_1 adrenoceptors (5-HT_{1A}R/ α_{1a} = 18) (Figure 2, panel b) [38]. In a more recent paper, docking of **1** on the newly published 5-HT_{1A}R model was performed and a set of structural analogues, substituted at C8 position of the 1,4-dioxaspiro[4.5]decane moiety was explored [39]. All compounds displayed low affinity and activity at 5-HT_{1A}R, indicating that only small substituents are allowed, while a higher affinity was observed at α_1 -adrenoceptors, resulting in a significant reversal of selectivity.

With the aim of improving 5-HT_{1A}R/ α_1 selectivity as well as potency and efficacy, in this work we designed a new set of structural analogues of **1** focusing the attention on both the 1,4-dioxaspiro[4.5]decane and arylpiperazine moieties (Figure 2, panel c). As regard the first portion, on the basis of previously published data showing that the replacement of one or two ring oxygen atoms with sulfur leads to a progressive decrease of α_1 affinity, the 1-oxa-4-thiaspiro- and 1,4-dithiaspiro[4.5]decane-analogues were synthesized and tested [38]. In addition, the replacement of the piperazine ring with a more flexible basic chain was investigated. By merging these two structural modifications, compound **15** emerged as a potent and selective 5-HT_{1A}R agonist endowed with neuroprotective activity *in-vitro* and a potent antinociceptive activity in *in-vivo* model. Finally, molecular modelling on the two GPCRs was performed to better understand the basis of activity and selectivity.

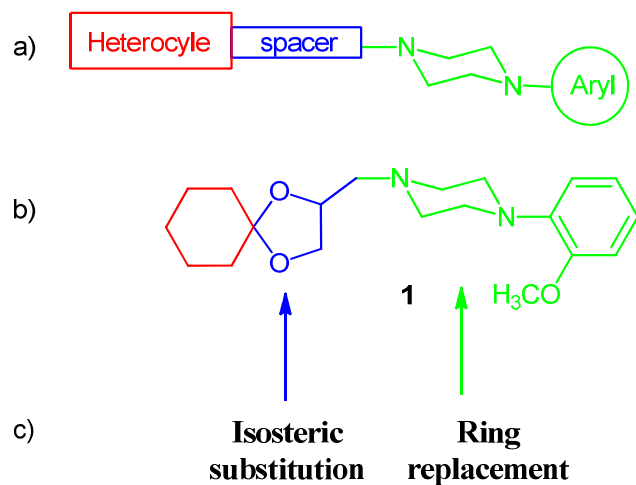


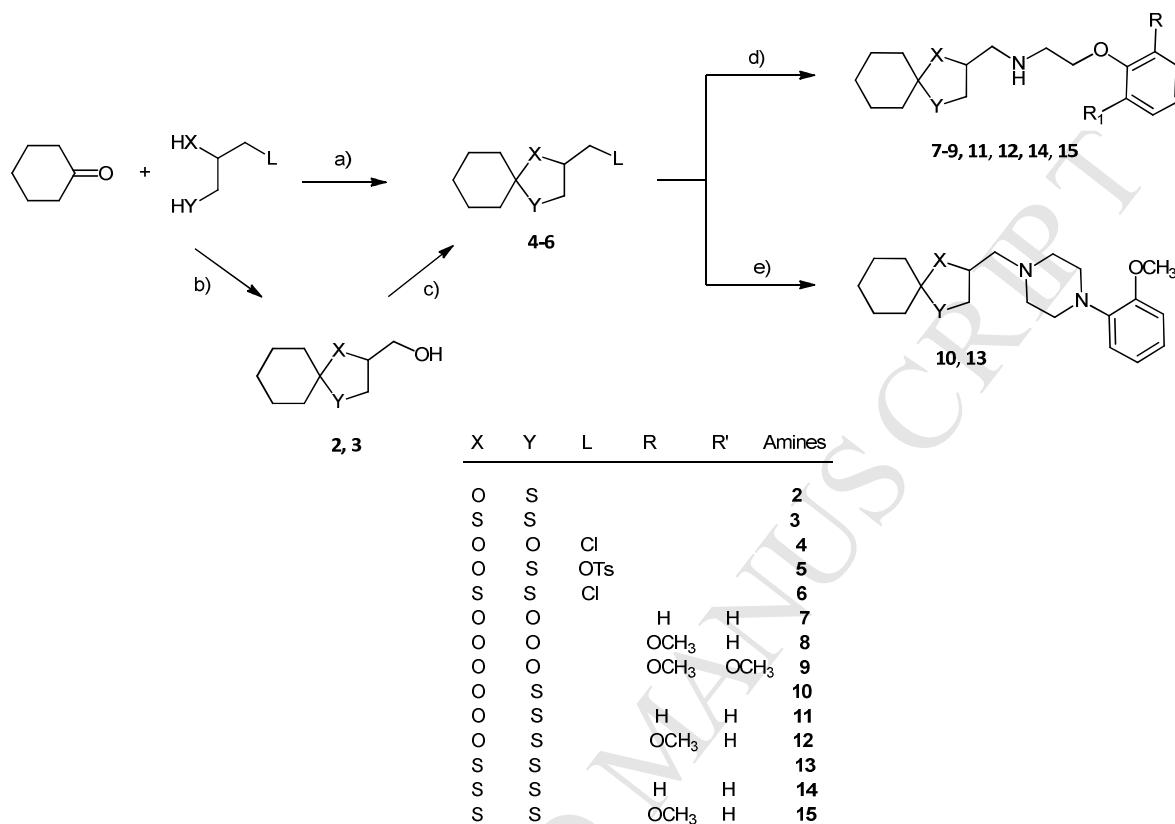
Figure 2. a) Pharmacophoric model of 5-HT_{1A} agonist; b) Chemical structure of compound **1** (pD₂ 5-HT_{1A} = 8.61, 5-HT_{1A} α_1 = 18); c) Working hypothesis

Results and Discussion

Synthesis

The compounds **7-15** were obtained by alkylation of the commercially available 1-(2-methoxyphenyl)piperazine or substituted 2-phenoxyethanamines with the proper chloro- or

tosyl- derivative, under microwave irradiation (Scheme 1).



Scheme 1. Reagents and conditions: a) 3-Chloro-1,2-propanediol, pTsOH, Dean-Stark, 48 h, quantitative for **4**; b) 3-mercaptopropane-1,2-diol or 2,3-dimercaptopropan-1-ol, HClO₄/SiO₂, r.t., 6 h, 81% for **2**, 71% for **3**; c) TsCl, Et₃N, DCM, 0 °C to r.t., 12 h, 82% for **5** or SOCl₂, toluene, 0 °C to 80 °C, 12 h, 30% for **6**; d) 2-phenoxyethanamine or (2-methoxyphenoxy)ethanamine or (2,6-dimethoxyphenoxy)ethanamine, KI, 2-methoxyethanol, MW, 160 °C, 30 min., 51% for **7**, 54% for **8**, 54% for **9**, 25% for **11**, 19% for **12**, 15% for **14**, 47% for **15**; e) 1-(2-methoxyphenyl)piperazine, KI, 2-methoxyethanol, MW, 160 °C, 30 min, 67% for **10**, 53% for **13**.

The phenoxyethanamines were easily prepared by reacting the chloroacetamide with the appropriate phenates, followed by reduction of the amides, as previously reported [40]. The 1,3-dioxolane-, oxathiolane- and dithiolane-scaffolds were readily prepared by acetalization of the cyclohexanone with the proper glycerol derivatives. In the case of 3-mercaptopropane-1,2-diol or

2,3-dimercaptopropan-1-ol, perchloric acid adsorbed on silica-gel ($HClO_4-SiO_2$) was employed as a new, highly efficient, inexpensive and reusable catalyst for acetal formation, under solvent-free conditions. The hydroxylic group was then converted into a better leaving group (tosylate or chloride) for the subsequent coupling reaction.

Pharmacology

The pharmacological profile of the synthesized compounds **7-15**, the lead compound **1** and BMY-3748, as reference compounds, was evaluated by radioligand binding assays using [3H]prazosin to label cloned human α_1 adrenoceptors expressed in CHO cells [41], and [3H]8-OH-DPAT to label cloned human 5-HT_{1A} receptors expressed in HeLa cells [42].

Functional characterization of the most active and selective compounds (**1**, **13-15**, and BMY-7378) at the 5-HT_{1A}R was performed according to methods of Stanton and Beer [43] using [^{35}S]GTP γ S binding, in cell membranes from HeLa cells transfected with the human cloned 5-HT_{1A}R. Stimulation of [^{35}S]GTP γ S binding was expressed as the percent increase in binding above basal value; maximal stimulation observed with serotonin was established as 100%.

Cytotoxicity assays were carried out against human neuroblastoma cell line SH-SY5Y. Cells were cultured at 37 °C in a humidified incubator containing 5% CO₂ and feed with DMEM (Lonza) nutrient supplemented with 10% heat inactivated FBS, 2 mM L-glutamine, 100 U/mL penicillin and 100 μ g/mL streptomycin. Cytotoxicity of compounds is expressed as IC₅₀ values, the concentrations that cause 50% growth inhibition. The results were determined using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide [44].

The neuroprotective capacity of the compounds was tested, as reported by Benchekroun et al. [45] Briefly, the ability of the compounds to prevent the human neuroblastoma cell line SH-

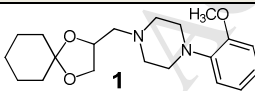
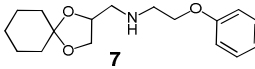
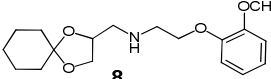
SY5Y from death was evaluated by using three toxicity models: 1) H_2O_2 , as a producer of exogenous free radicals, 2) oligomycin A, a mitochondrial respiratory chain blocker which produces mitochondrial ROS by inhibiting the mitochondrial electron-transport chain complex V, and 3) rotenone, showing the same effect of oligomycin A by inhibiting the mitochondrial electron-transport chain complex.

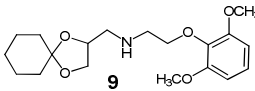
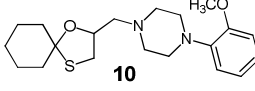
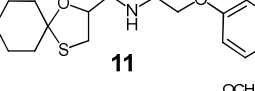
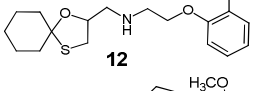
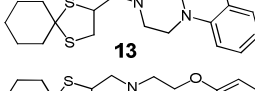
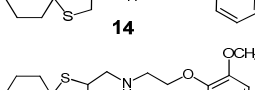
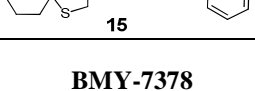
For the assessment of inflammatory pain, mice were subjected to the formalin test. Intraplantar administration of formalin (5%, 10 μl) produces a biphasic nocifensive behavioral response (i.e., licking or biting the injected hind paw). The acute nociceptive phase lasts for the first 10 min, whereas the second inflammatory phase occurs between 15 and 50 min and reflects the development of nociceptive sensitization in the dorsal horns of the spinal cord [46].

Structure–affinity and structure-activity relationships

All synthesized compounds were tested for binding affinity/activity at $5\text{-HT}_{1\text{A}}$ and α_1 receptors. Lead compound **1** and BMY-7378 were used as reference compounds on the basis of structure similarities and high affinity/activity for $5\text{-HT}_{1\text{A}}\text{R}$ and α_1 receptors.

Table 1. Affinity constants (pK_i)^a and selectivities^b of test and reference compounds for the human recombinant α_1 adrenoceptor subtypes and the $5\text{-HT}_{1\text{A}}\text{R}$.

Compound	α_{1a}	α_{1b}	α_{1d}	$5\text{-HT}_{1\text{A}}$	α_{1d}/α_{1a}	α_{1d}/α_{1b}	α_{1b}/α_{1a}	$5\text{-HT}_{1\text{A}}/\alpha_{1d}$
 1	7.04	6.90	<6	8.29	<0.1	<0.1	0.7	18
 7	<6	6.01	6.26	7.43	1.8	1.7	1	15
 8	6.64	6.41	7.02	8.52	2.4	4	0.6	31

	<6	<6	5.75	<6	0.5	0.5	1	1
	7.24	6.72	7.41	8.30	1.5	4.9	0.3	7.8
	<6	6.36	6.67	7.64	4.7	2	2.3	9.3
	6.57	6.66	7.49	8.65	8.3	6.8	1.2	14.4
	6.70	6.39	7.00	8.52	2	4	0.5	33
	<6	6.26	6.91	8.81	8	4.5	1.8	79
	6.73	6.71	7.46	9.03	5.4	5.6	1	37
BMX-7378	6.41	6.15	8.89	8.90	295	550	2	1

^a K_i values were derived from the Cheng–Prusoff equation [47] at one or two concentrations and agreed within 10%. ^b Antilog of the difference between the pK_i values for α_{1a} , α_{1b} and α_{1d} adrenoceptors. ^c Antilog of the difference between the pK_i values for α_1 adrenoceptors (higher value) and the 5-HT_{1A}R.

As shown in Table 1, the substitution of the 2-methoxy-phenylpiperazine moiety of **1** with the more flexible phenoxyethylamine chains, as for compounds **7–9**, caused substantial changes in binding affinity. In particular, the phenoxy derivative **7** showed a significant decrease in affinity, of about one order of magnitude, for both 5-HT_{1A} and α_1 receptors, with the exception for the α_{1D} subtype, leaving almost unchanged the 5-HT_{1A}/ α_1 selectivity. The 2-methoxy-phenoxy analogue **8**, showed a 10-fold increased affinity for 5HT_{1A}R, compared to **7**, thus restoring the affinity of the lead compound **1** and doubling its 5-HT_{1A}/ α_1 selectivity, as the increase also seen for α_1 subtypes is to a lesser extent. These results indicate the importance of the 2-methoxy group in stabilizing the binding process (see molecular modeling section). A similar trend was previously observed for the series of the 2,2-diphenyl-1,3-dioxolane derivatives [44]. On the contrary, the

introduction of a second methoxy group, as for **9**, caused a drastic loss in affinity at both receptor systems, indicating that a steric hindrance prevents a good anchoring to the binding site.

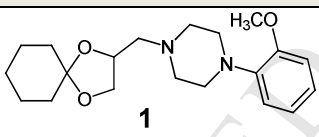
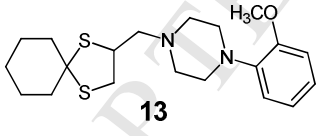
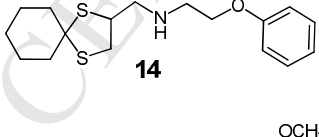
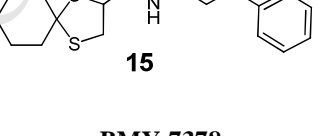
To investigate the effect of the replacement of the ring oxygen atom with a sulfur, 1,3-oxathiolane derivatives **10-12** were prepared. The binding data showed that at 5HT_{1A}R this substitution leaves unchanged the affinity, while it produces some effect on α_1 adrenoceptors. A significant increases of affinity was observed only for α_{1D} subtype in the 1,3-oxathiolane series (**10-12**) compared to the corresponding 1,3-dioxolanes (**1**, **7**, **8**), thus the 5-HT_{1A}/ α_1 selectivity ratio is halved. Again, the 2-methoxyphenoxy derivative **12** showed the highest 5-HT_{1A} affinity within this series (**12** > **10** >> **11**).

Surprisingly, when both oxygens atoms are replaced by sulfur, as for the 1,3-dithiolane derivatives **13-15**, the affinity at 5-HT_{1A}R increased, while, at α_1 subtypes, it is decreased or unchanged. The enhancement at 5-HT_{1A}R is remarkable only for **14**, whose affinity is 15- and 24-fold higher than the ones of the corresponding 1,3-oxathiolane **11** and 1,3-dioxolane **7**. In this case the role of the 2-methoxy group on the phenoxyethylamine chain is less relevant, as the variations in affinity, especially at 5-HT_{1A}R, is very limited and scarcely significant. This is probably due to an additional interactions of the 1,4-dithia-spiro[4.5]-decane moiety of **14** in stabilizing the binding mode of the protein-ligand complex, by several contacts with a deep receptor cavity, including V117, K191, Y195, T196, S199, T200, F361, F362, A365 (see molecular modeling section). Among all, **14** is outstanding in terms of selectivity with a 5-HT_{1A}/ α_1 ratio of 80. These findings support the previously published data and seem to confirm that moving from 1,3-dioxolanes or 1,3-oxathiolanes to 1,3-dithiolanes the affinity and/or selectivity at/for 5-HT_{1A}R progressively increases [44]. With the exception of compounds **7** and

11, the same improvement is observed by replacing the 2-methoxyphenylpiperazine with the more flexible 2-methoxy-phenoxethylamine.

Functional characterization at 5-HT_{1A}R was performed for the most promising compounds (**13-15**). The results are reported in Table 2. In stimulation experiments, compounds **13-15** increased the binding of [³⁵S]GTPγS with pD₂ values of 8.61, 8.27 and 9.58, respectively. The E_{max} values were of 37, 85 and 74, defining the three compounds as partial agonists. Notably, the most active compound **15** exhibited an improved potency (10-fold) and efficacy (1.5 fold) compared to the lead compound **1**, with a pD₂ value higher (10-fold) than that of the reference full agonist 8-OH-DPAT (pD₂= 8.49, Figure 3).

Table 2. Agonist potency (pD₂), relative effectiveness (E_{max}) in the agonist-induced [³⁵S]GTPγS-binding assay at the human 5-HT_{1A} receptor.

Compound	pD ₂	E _{max} ^a
 1	8.61	48
 13	8.61	37
 14	8.27	85
 15	9.58	74
BMX-7378	9.27	26
8-OH DPAT	8.49	100

^a Maximal stimulation expressed as a percentage of the maximal 5-HT response.

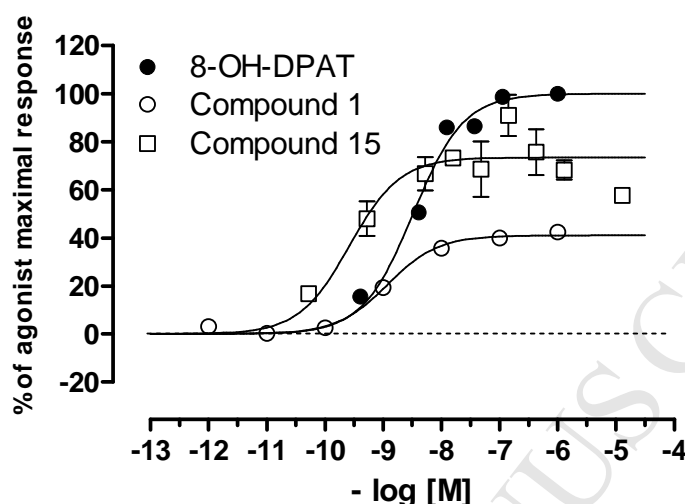


Figure 3: Stimulation of [35 S]GTP γ S binding in HeLa cells expressing human recombinant 5-HT $_1$ A R by compound **15** and the reference full agonist 8-OH-DPAT.

Prediction of ADMET Properties

The computational prediction of descriptors related to absorption, distribution, metabolism, excretion and toxicity properties (ADMET) represents a useful *in silico* strategy accelerating the lead compound discovery process [48]

In this work, for compounds **1**, **7-15** extent of blood-brain barrier permeation (LogBBB), rate of passive diffusion-permeability (Log PS), human intestinal absorption (HIA), volume of distribution (Vd), median lethal dose (LD $_{50}$) related to oral administration and the logarithmic ratio of the octanol-water partitioning coefficient (cLogP) were calculated.

As shown in Table 5, all the compounds are characterized by a favourable profile in terms of lipophilicity, being the calculated LogP below 5 (Lipinski rules) and also display the ability to fully be adsorbed at the human intestinal membrane. Notably, compounds **1**, **10**, **13-15** show

higher blood-brain barrier permeation with respect to the other compounds, being in any case all of them able to pass at the central nervous system. Finally, all the compounds exhibit a favourable toxicity profile, being the estimated LD₅₀ in the range of 600-1200 mg/kg for mouse after oral administration.

Table 3. Calculated ADMET properties for compounds **1**, **7-15**.

Comp.	LogBBB ^a	LogPS ^b	HIA (%) ^c	Vd (l/kg) ^d	LD ₅₀ (mg/kg) ^e	cLogP
1	0.76	-1.2	100	4.1	640	3.24
7	0.26	-1.6	100	3.3	1200	2.88
8	0.21	-1.9	100	3	1200	2.52
9	0.26	-1.9	100	2.8	1200	2.83
10	0.82	-1.1	100	3.5	680	3.96
11	0.42	-1.3	100	6	1100	3.80
12	0.32	-1.6	100	5.6	1000	3.30
13	0.89	-1.1	100	3.4	600	4.58
14	0.75	-1.2	100	6.2	890	4.49
15	0.61	-1.3	100	5.7	870	4.15

^a Extent of brain penetration based on ratio of total drug concentrations in tissue and plasma at steady-state conditions; ^b Rate of brain penetration. PS represents Permeability-Surface area product and is derived from the kinetic equation of capillary transport; ^c HIA represents the human intestinal absorption, expressed as percentage of the molecule able to pass through the intestinal membrane; ^d prediction of Volume of Distribution (Vd) of the compound in the body; ^e Acute toxicity (LD₅₀) for mouse after oral administration (RI ≥ 30)

Neuroprotective capacity of compound **15**

The neuroprotective capacity of **15** was tested for its ability to prevent the human neuroblastoma cell line SH-SY5Y from cell death induced by three toxicity models: 1) hydrogen peroxide for the generation of exogenous free radicals, 2) oligomycin A, a mitochondrial

respiratory chain blocker which produces mitochondrial ROS by inhibiting the mitochondrial electron-transport chain complex V, and 3) rotenone, showing the same effect of oligomycin A by inhibiting the mitochondrial electron-transport chain complex I [45]. Before the assessment of the neuroprotective capacity, the direct cytotoxicity of compound **15** was investigated and cell viability after 24 h of exposure was measured by MTT assay across a wide concentration range (0.1-100 μM). The IC_{50} values were 195 ± 1.7 , 29 ± 3.4 , 74.1 ± 4.5 and 51 ± 5 μM for H_2O_2 , oligomycin A, rotenone and compound **15**, respectively. Therefore, in the neuroprotective test, compound **15** was used at a concentration of 0.1 and 1 μM . In particular, as reported in Table 3, compound **15** showed good neuroprotective effect against the insult caused by oligomycin A and H_2O_2 (only at concentration of 0.1 μM) whereas it exhibited a minimum neuroprotection against rotenone, at the two tested concentrations.

Table 4. Protective effect of compound **15** on SH-SY5Y cell death induced by H_2O_2 or oligomycin A or rotenone. Data are expressed as percent neuroprotection \pm SD of three independent experiments.

Compd [μM]	H_2O_2 (195 μM)	Oligomycin A(30 μM)	Rotenone (75 μM)
15 (1 μM)	66 ± 4	86 ± 5	66 ± 2
15 (0.1 μM)	83 ± 6	81 ± 4	62 ± 2

Bi-directional transport studies on MDCKII-MDR1 monolayers

Many cell-based *in-vitro* methods have been developed to determine the BBB permeation of compounds under investigation. Among them, MDCK-MDR1 cell line represents a well-established *in-vitro* method mimicking the BBB [49, 50, 51]. It is well known that MDCKII-MDR1 cells form tight monolayers and express P-glycoprotein (P-gp), which is specifically involved in the efflux transport of drugs from the BBB. In particular, we were interested in

assessing whether compound **15** is able to permeate MDCK-MDR1 monolayers and to interact with P-gp. Thus, transport studies were conducted both in AP-to-BL and BL-to-AP direction and the results are reported in Table 4. Compound **15** showed not significant differences in P_{app} values between AP-to-BL and BL-to-AP direction and the efflux ratio (ER) calculated by the equation $ER = P_{app, BL-AP} / P_{app, AP-BL}$ was found to be less than 2, indicating that this compound is not likely to be considered substrate for P-gp transport. These results suggest that **15** is able to permeate the monolayer by passive diffusion with permeability values comparable to diazepam. The results for the controls were within the expected values.

Table 5. Bi-directional Transport Across MDCKII-MDR1 cells of compound **15**.

Compd	P_{app} AP(cm/sec)	P_{app} BL(cm/sec)	ER ^a $P_{app}BL/P_{app}AP$
15	1.11×10^{-5}	1.09×10^{-5}	0.98
diazepam	1.46×10^{-5}	1.23×10^{-5}	0.84
FD-4	1.03×10^{-6}	2.08×10^{-7}	0.20

^a Efflux ratio (ER) was calculated using the following equation: $ER = P_{app, BL-AP} / P_{app, AP-BL}$, where $P_{app, BL-AP}$ is the apparent permeability of basal-to-apical transport, and $P_{app, AP-BL}$ is the apparent permeability of apical-to-basal transport. An efflux ratio greater than 2 indicates that a test compound is likely to be a substrate for P-gp transport.

Antinociceptive activity

Compound **15**, endowed with high affinity and high agonist potency, was chosen for determining potential analgesic activity *in-vivo*. For the assessment of which, mice were subjected to the formalin test. As shown in Figure 4, compound **15** was administered 15 min before formalin at the dose of 3, 5 and 10 mg/kg i.p. The dose of 10 mg/kg was able to induce a significant analgesic effect in the second phase of formalin test (* $p < 0.05$). Pretreatment with the selective 5-

HT_{1A}R antagonist WAY-100635 (3 mg/kg i.p.), 30 min before of compound **15** (10 mg/kg i.p.), prevented its analgesic effect ($\#p < 0.05$). WAY-100635 (3 mg/kg i.p.), *per se* at least at the used dose, did not modify the licking response after formalin injection (Figure 5).

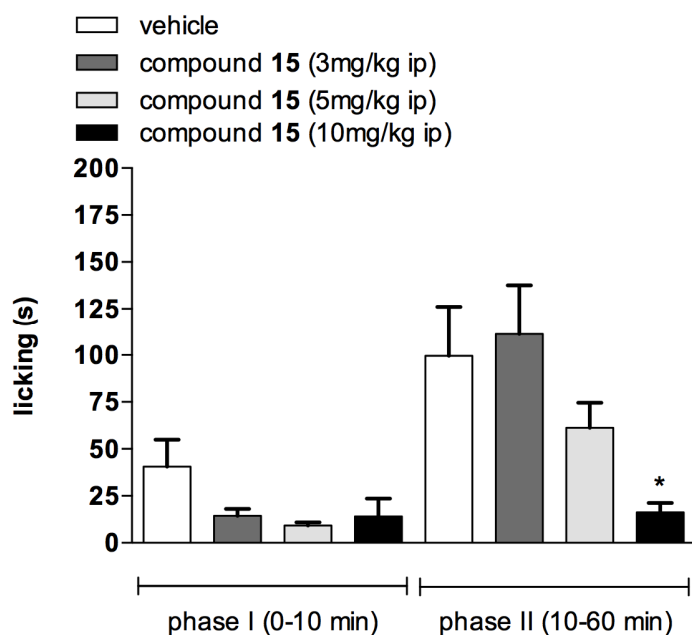


Figure 4. Effect of intraperitoneal (i.p.) injection of **15** (3, 5 or 10 mg/kg) on the first (0-10 min) and second (10-60 min) phase of the formalin test. Test compound or vehicle were injected 15 min prior to the intraplantar injection of formalin. Data are means + S.E.M. of 8-10 mice per group. * $p < 0.05$ vs. the respective groups of mice injected with vehicle.

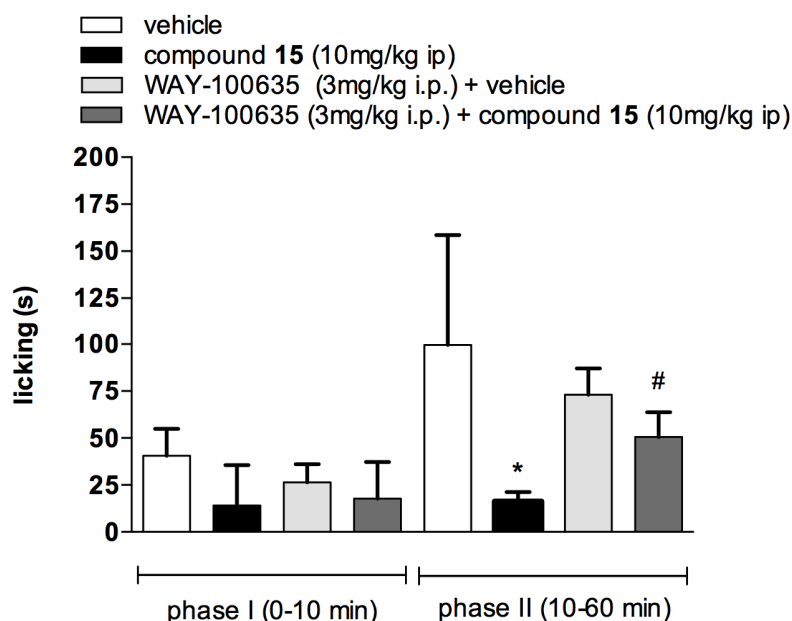


Figure 5 Effect of WAY-100635 (3 mg/kg i.p.) on analgesia induced by **15** (10 mg/kg i.p.) during the first (0-10 min) and second (10-60 min) phase of the formalin test. Test compound or vehicle were injected 15 min prior to the intraplantar injection of formalin. WAY-100635 was injected 30 min prior to **15** or vehicle. Data are means + S.E.M. of 8-10 mice per group. * $p < 0.05$ vs. the respective groups of mice injected with vehicle. # $p < 0.05$ vs mice treated with **15** (10 mg/kg i.p.).

Molecular Modeling

Molecular modelling studies have been undertaken in order to better elucidate the affinity and selectivity profiles of the newly synthesized analogues of compound **1**. To gain a perspective of the most relevant patterns of key contacts involved in the ligand binding mode, we performed our work based on 5-HT_{1A}R and α_{1D} R homology models. In the first case, we relied on the previously built 5-HT_{1A}R theoretical model employed in docking studies of several *in-house* series of 5-HT_{1A} ligands [39,52,53]

For the human α_{1D} receptor, a specific model has been built and is here discussed, focusing our attention on α_{1D} R subtype, because of the high affinity values shown by all compounds.

α_{1D} Homology Modeling

The 3D structure of the human α_{1d} receptor has been generated following the ligand-based homology modeling strategy proposed by Moro [54]. This computational option is very useful to build an homology model in the presence of a ligand docked into the primary template and has been widely and fruitfully used to build GPCR so as enzyme models [55,56,57]. In this case we selected a structurally-related analogue of compound **1** (compound **A**, as shown in Figure 6), exhibiting high degree of affinity and selectivity toward the α_{1D} receptor ($pK_i \alpha_{1D} = 9.09$, $\alpha_{1D}/5-HT_{1A} = 28$) [38].

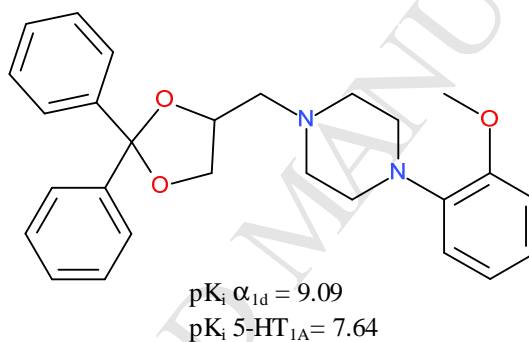


Figure 6. Structure of compound **A**

Accordingly, **A** was docked into the β_2 -adrenoreceptor X-ray coordinates (pdb: 2RH1) and employed in the building and refinement of the derived α_{1D} model. This kind of approach allowed us to set up a much more suitable receptor model, to be used for efficiently exploring the putative binding mode of analogues **7-15**. The sequence alignment of the α_{1D} receptor (P25100) with respect to the human β_2 -adrenoreceptor (pdb: 2RH1) coordinates is shown in Figure 7. The reliability of the alignment was verified by the high value of the pairwise percentage residue identity (PPRI = 42%). Accordingly, a consistent number of α_{1D} residues resulted to be conserved in comparison with those of the β_2 -adrenoreceptor TM helices: (i) V97,

G98, L108, V111, G113, N114, L116, V117, I118, A122 in TM1, (ii) V130, T131, N132, Y133, F134, I135, L138, A139, A141, D142, L143, V149, P151, F152, A154 in TM2, (iii) G165, C168, W172, D176, V177, L178, C179, T181, A182, S183, F184, L187, C188, I190, V192, D193, R194, Y195 in TM3 (the DRY motif; 193-195 residues), (iv) K212, A213, I216, W221, V223, S228 P231 in TM4, (v) Y254, A255, S258, S259, S262, F263, Y264, P226, V269, I270, V272, Y274, R276, V277 in TM5, (vi) E343, K345, A346, K348, T349, L350, I352, G355, F357, L359, C360, W361, P363, F364, F365, V367 in TM6 (CWXP motif; 360-363 residues); (vii) E381, V383, W389, G391, Y392, N394, S395, N398, P399, L400, I401, Y402, C404 in TM7 (NPXIY motif; 398-402 residues).

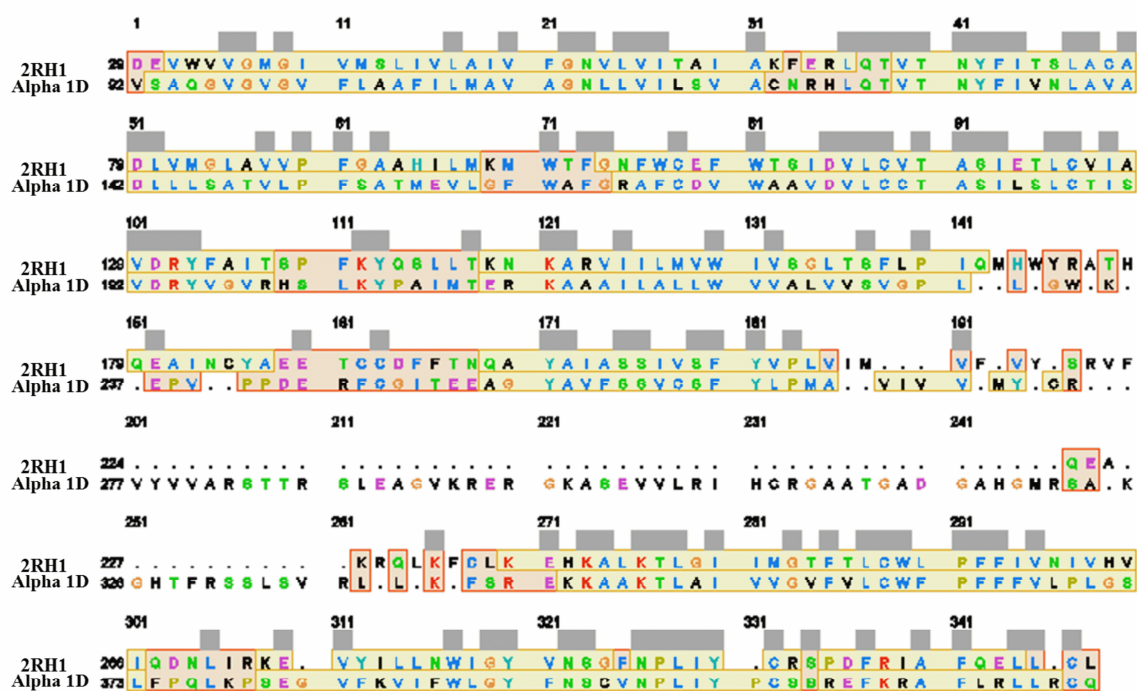


Figure 7. Sequence alignment of the α_1D on the human β_2 -adrenoreceptor (pdb: 2RH1) coordinates. Any conserved region is displayed by grey histograms. All the residues included in α -helix and loop domains are highlighted in light yellow and pink, respectively.

The derived backbone conformation was inspected by Ramachandran plot (showing absence of outliers) and superimposed to the coordinates of the template structure (RMSD = 0.747 Å; Figure 8). See Materials and Methods Section for further computational details.

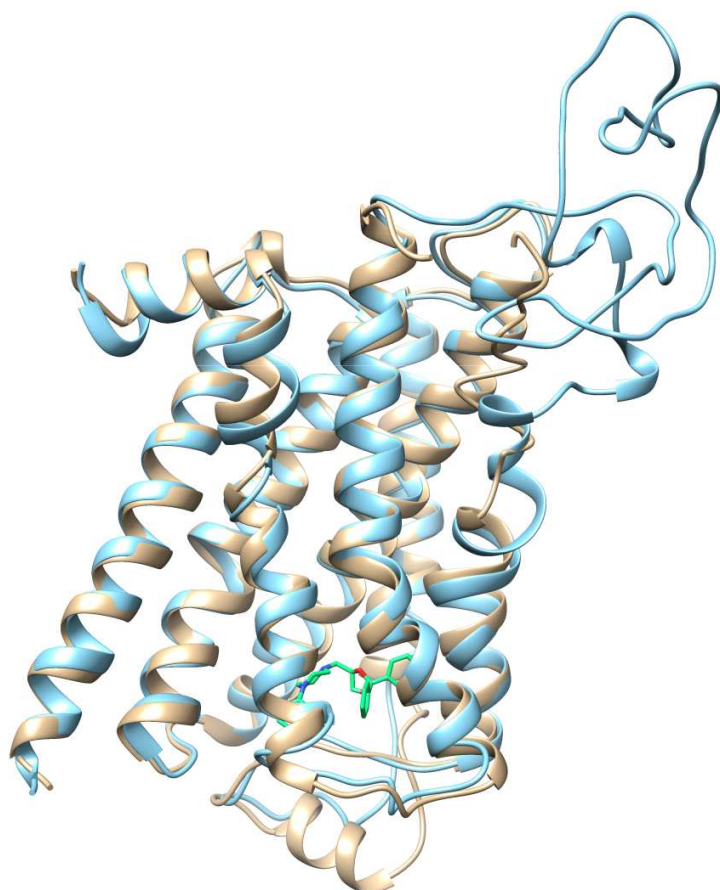


Figure 8. The superimposition of the final α_{1D} model (backbone in cyan) on the human β_2 -adrenoreceptor 2RH1 coordinates (backbone in khaki) is depicted. Ligand **A** structure is also depicted in stick (C atom: light green)

Molecular docking of compound **A**

In a second computational step, ligand **A** was docked into the two in-house GPCRs models in order to examine and compare the corresponding binding modes at the α_{1D} and 5-HT_{1A} receptors. According to our calculation, **A** (the *R* enantiomer proved to be the most probable)

was highly stabilized into the α_{1D} binding site through a salt-bridge interaction between the piperazine protonated nitrogen atom and the conserved D176, and by two H-bonds between: (i) one oxygen atom of the 1,3-dioxolane core and Y254 side-chain; (ii) the methoxy group and K385 ϵ -amino group. Furthermore, the 2-methoxyphenyl ring and the diphenyl substituents were also engaged in π - π stacking with W172, F388 and Y254, F364 and F365, respectively (Figure 9).

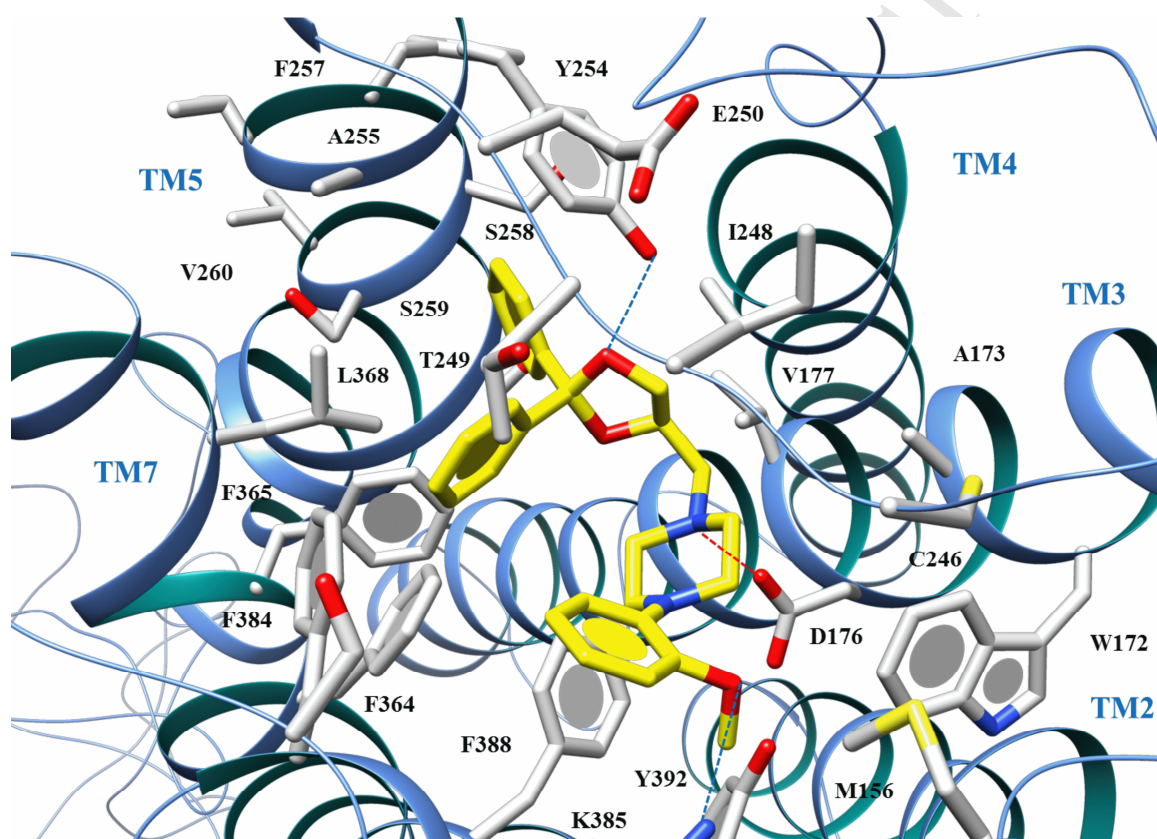


Figure 9. The ligand **A** docking pose into the final α_{1D} binding site is reported. The ligand is coloured by atom-type (C atom: yellow). Salt-bridge and H-bond contacts are displayed by line and coloured in red and light blue, respectively.

Differently, at 5HT_{1A}R, ligand **A** (the *R* enantiomer proved to be the most probable; pK_i 5-HT_{1A} = 7.64) displayed a salt-bridge between the piperazine protonated nitrogen atom and the

key residue D116, while only one H-bond was detected between the oxygen atom of the dioxolane ring and Y390. The lack of an additional H-bond interaction of the methoxy group into the 5HT_{1A} binding site turned around the docking pose of **A** with respect to α_{1D} adrenoceptor. In this case, the methoxyphenyl ring and the diphenyl portion were oriented towards Y195, F362 and Y96, F112, Y390 respectively, determining π - π stacking interactions (Figure 10).

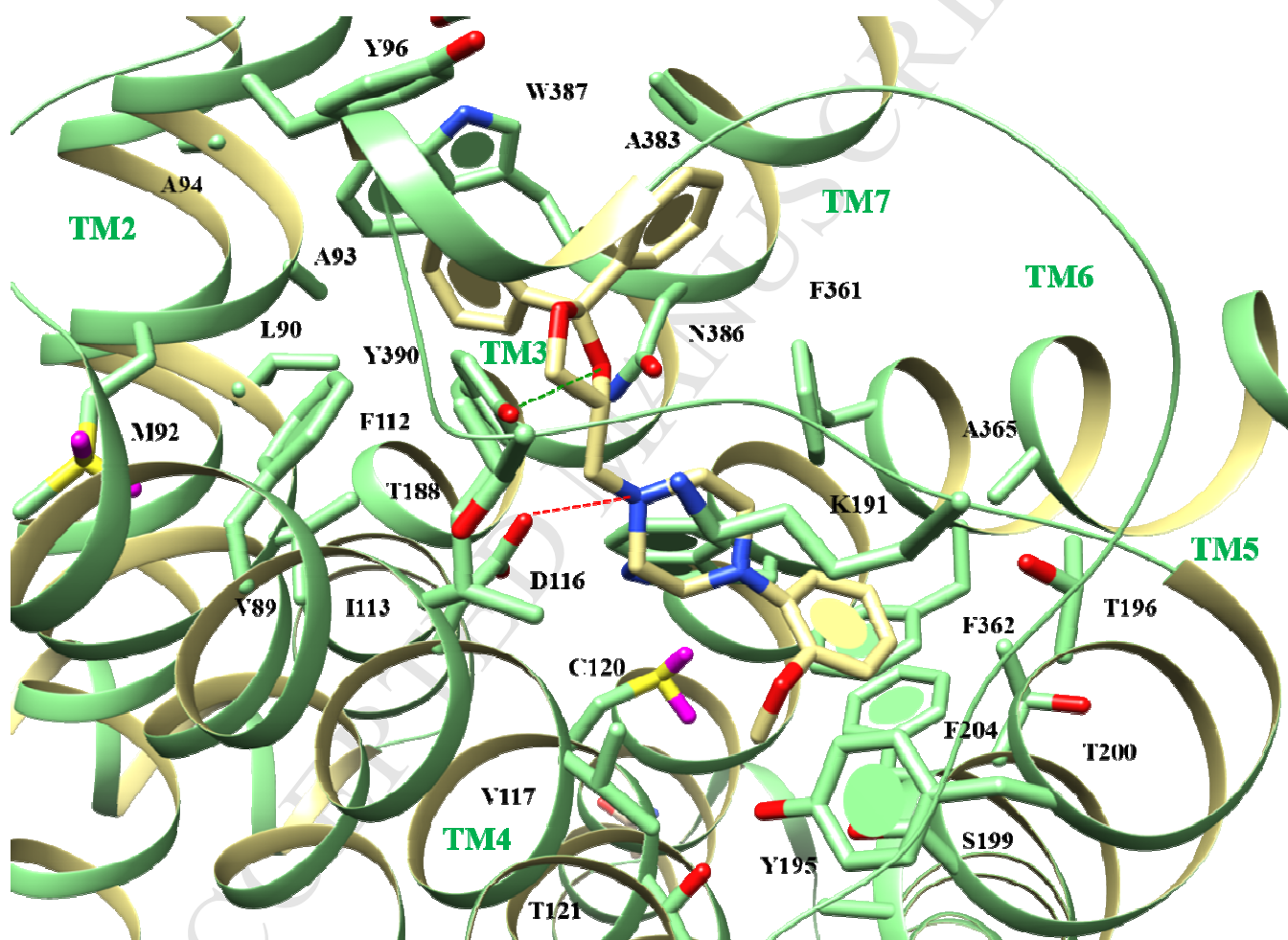


Figure 10. The ligand **A** docking pose into the 5HT_{1A} binding site is reported. The ligand is coloured by atom-type (C atom: khaki). Salt-bridge and H-bond contacts are displayed by line and coloured in red and light green, respectively.

However, this orientation was detrimental for the binding with the 5-HT_{1A}R, as confirmed by the biological data showing a lower affinity at 5-HT_{1A} ($pK_i = 7.64$) with respect to α_{1D} ($pK_i = 9.09$) (Figure 6).

Notably, these results highlighted a number of amino acids interacting with the ligands which are conserved within 5-HT_{1A}R and α_{1D} , in particular a key aspartic acid residue, showing that the two protein binding sites share a quite common hydrophobic/hydrophilic trend. In particular, the amino acids contributing to the ligand binding were classified by us into two sets:

- a first set consisting of specific residues present in both receptors and implicated in strong interactions. For example the acidic D176 in α_{1D} R or D116 in 5-HT_{1A}R, which represents the common anchoring points through the formation of a salt-bridge interaction plus some polar residues like Y254 for α_{1D} R and Y195 for 5-HT_{1A}R, which further stabilize the ligand binding through an H-bond interaction;
- a second set referring to specific amino acids present only in 5-HT_{1A}R or α_{1D} R, allowing the formation of H-bonds responsible of the ligand specificity towards the receptor. For example K385 in α_{1D} R and N386 in 5-HT_{1A}R seem to be fundamental in conferring specificity. Thus, any focused H-bond with K385 or N386 residue, might allow the discovery of more selective ligands towards α_{1D} or 5-HT_{1A}R, respectively.

5-HT_{1A}R Docking-studies

In our previous works we deeply investigated through docking studies 1,3-dioxolane-, 1,3-oxathiolane-, 1,3-dithiolane-, spiro-dioxolane-tetrahydrofuran-, cyclopentanone- and cyclopentanol-based derivatives as 5-HT_{1A} ligands, whose affinity profile proved to be due to the presence of a proper basic feature interacting with the aspartic acid D116. Moreover, agonists

and antagonists could also exhibit additional contacts with N386 and Y390, and with K191, respectively [52,53].

Notably, a number of following docking studies reported in literature described a unique receptor cavity involved in the 5-HT_{1A} full agonists, partial agonists and antagonists binding [58-60], giving a further validation to our previous computational findings. Indeed, H-bond interactions between agonists and D116 and N386 were reported in literature, falling in a crevice delimited by F112, I113, D116, K191, while partial agonists as well as antagonists were H-bonded at least with D116.

Docking studies on compounds **7-15** here performed within the previously built 5-HT_{1A}R homology model [39,52,53] allowed us to further explore their structure-activity relationships (SAR) and to gain more insight about the pattern of substitutions involved in their affinity. According to our calculations, the 1,4-dioxaspirodecane derivatives **7** and **8** shared the same docking mode of the previously described compound **1** [39]. Indeed, both compounds (the *R* enantiomers proved to be the most probable; pK_i 5-HT_{1A} = 7.43, 8.53), were properly stabilized into the 5-HT_{1A} binding site through the key salt-bridge between the protonated nitrogen atom and D116. Moreover, one H-bond interaction between the secondary amine and Y390 side chain was detected, as for **1**, while the 1,4-dioxaspirodecane moiety occupied a hydrophobic cavity delimited by A93, Y96, Q97, F112, A383, I384 and Y390. Notably, unlike **1**, compound **8** was able to establish an additional H-bond between the 2-methoxy oxygen atom and Y195 (Figure 11).

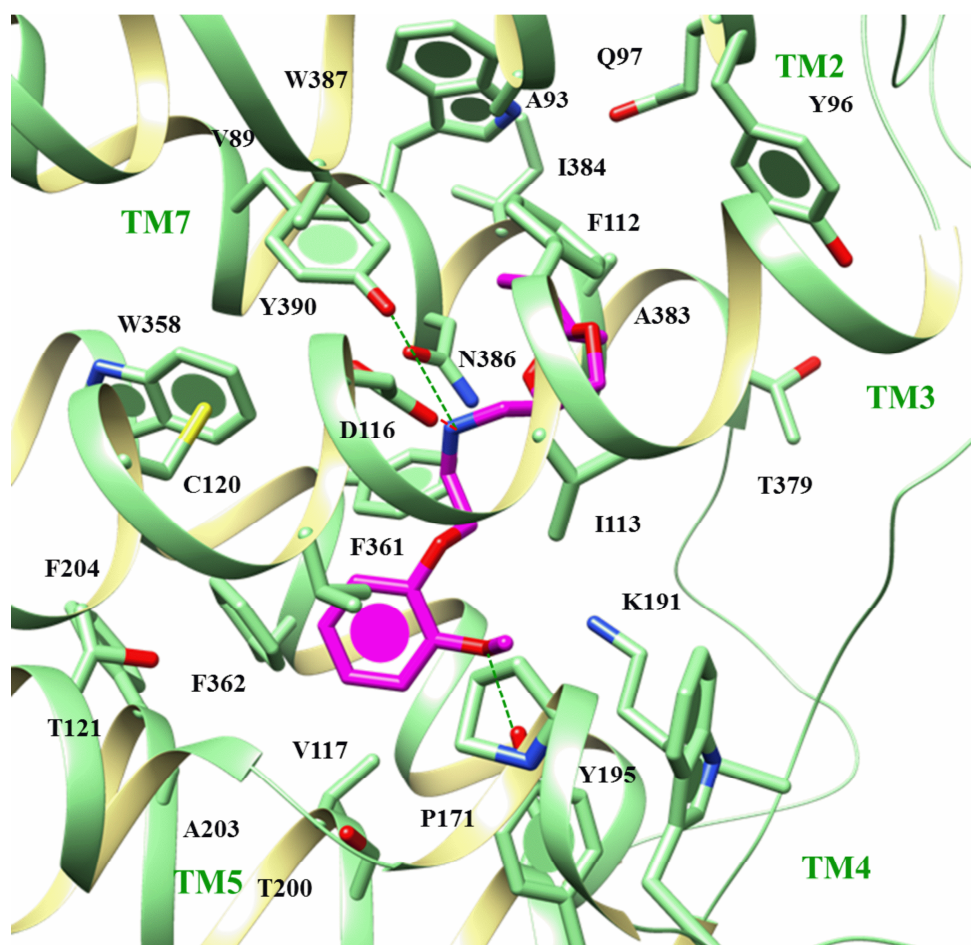


Figure 11 . The **8** docking pose into the 5HT_{1A} binding site is reported. The ligand is coloured by atom-type (C atom: magenta). Salt-bridge and H-bond contacts are displayed by line and coloured in red and light green, respectively.

This kind of positioning could be due to the presence of a flexible linker, as observed within all the members of the series (dioxo-, oxathia- dithiaspiro-decane derivatives), being in agreement with the higher affinity values of **8**, **12** and **15** with respect to **1**, **10** and **13**. Conversely, compound **9**, bearing a slightly bulkier amine chain, was unable to occupy the ligand binding site.

Interestingly, the members of the oxathia- (**10**, and **11**, **12** the *S* and the *R* enantiomers proved to be the most probable) and dithiaspiro-decane series (**13**, and **14**, **15** the *S* and the *R*

enantiomers proved to be the most probable) displayed a switched ligand binding mode, maintaining in any case a salt-bridge with D116, while the methoxy-analogues (**10**, **12** and **13**, **15**) also exhibited H-bonds with Y390 (in Figure 12 the docking mode of **12** and **15**, taken as reference compounds for the two series, are depicted).

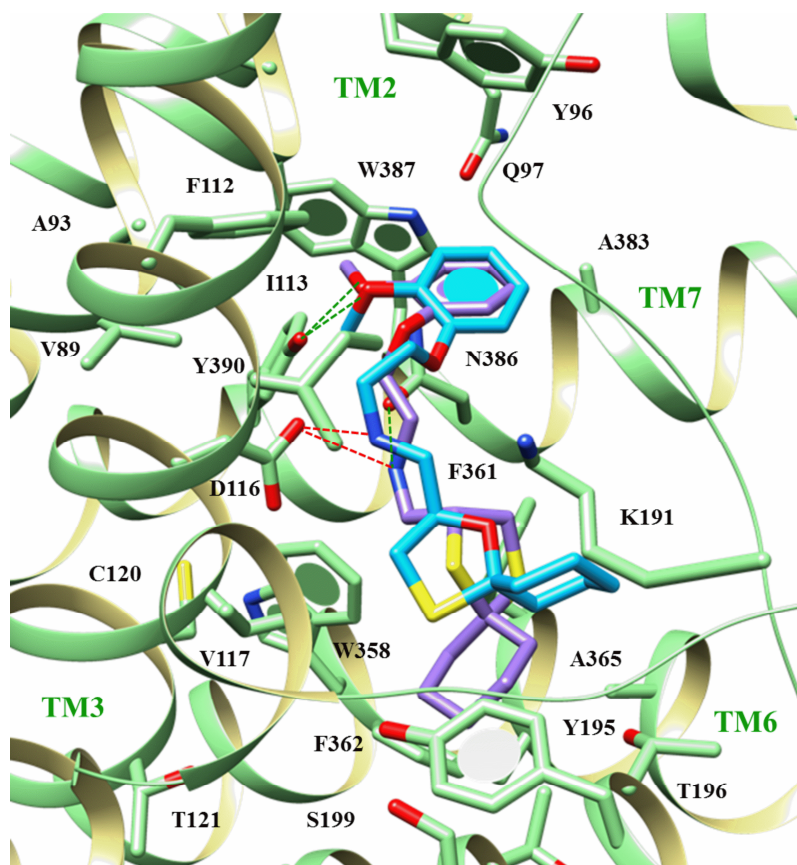


Figure 12. The compound **12** and **15** docking poses into the 5HT_{1A} binding site are reported. The ligands are coloured by atom-type (**12** C atom: cyan; **15** C atom: purple). Salt-bridge and H-bond contacts are displayed by line and coloured in red and light green, respectively.

The replacement of the dioxaspiro-decane of the prototype **1** with the much more bulkier oxathia- and dithiaspiro-decane moved the ligand cyclohexyl ring towards a deeper cavity including residues V117, K191, Y195, T196, S199, T200, W358, F362, A365, partially constraining the phenoxyethylamine chain flexibility. Consequentially, the dithiaspiro decane

derivatives **14** and **15** resulted highly stabilized into the 5-HT_{1A}R binding site also by H-bonds with N386, which proved to be a key residue for 5-HT_{1A}R agonism and selectivity. Accordingly, among all the compounds studied in this work, **14** and **15** showed the highest 5HT_{1A}/ α_{1D} selectivity ratio (79 and 37, respectively).

α_{1D} R docking-studies

In order to rationalize the low affinity values observed at α_{1D} R, the compounds were docked into the corresponding homology model. Briefly, all the ligands displayed a salt-bridge interaction between the protonated nitrogen atom and D176 anchoring residue. Moreover, when a methoxy group is present, as for compounds **8**, **10**, **12**, **13** and **15**, an H-bond interaction with Y254 side-chain is detected (in Figure 13 the docking mode of **12** and **15** are depicted).

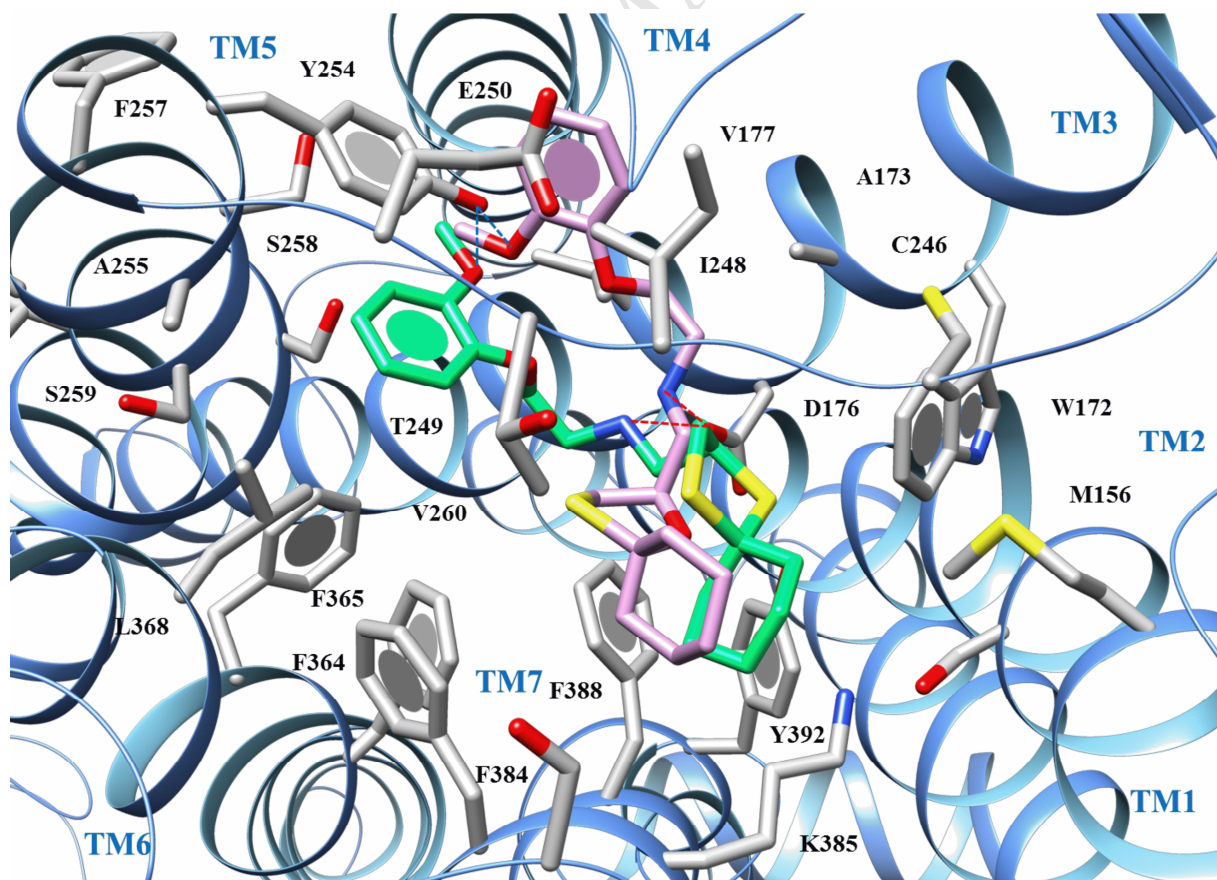


Figure 13. The compound **12** and **15** docking pose into the final α_{1D} binding site are reported. The ligand is coloured by atom-type (**12** C atom: light purple; **15** C atom: light green). Salt-bridge and H-bond contacts are displayed by line and coloured in red and light blue, respectively.

Again, as reported for 5-HT_{1A}R, the introduction of a flexible linker between the spiro-cyclic portion and the phenyl ring allow ligands to better occupy the receptor binding site. Thus, compounds which combine these two features displayed the higher affinity values of the series (**8**, **12** and **15**). However, a drastic drop of α_1 affinity is observed ($pK_i\alpha_{1D} < 7.5$) compared to the reference ligand **A** ($pK_i\alpha_{1D} = 9.09$). According to our results, this is probably related to the introduction of the spiro-decane moiety in place of the diphenyl-substituted ring. In particular, all these compounds oriented the spiro-cyclic portion towards a deep hydrophobic cavity including M156, W172, C246, F384, K385, F388, by detecting Van der Waals contacts. Thus, the phenyl or 2-methoxyphenyl group of the amine chain partially occupies the receptor region delimited by V177, Y254, F364, F365, L368. On this basis, the absence of an aromatic substituent on the spiro-cyclic moiety could cause a reversed ligand binding mode with respect to **A**, guiding the phenyl or methoxy-phenyl group towards Y254, F364, F365 residues. In this way, although a number of π - π stacking contacts were conserved, as for **A**, the relevant H-bond interaction with K385 is lost, resulting in lower affinity values.

Conclusion

In the present work a series of derivatives of **1** were synthesized and tested for binding affinity and activity at 5-HT_{1A}R and α_1 adrenoceptors and SAR studies were drawn (Figure 14). In particular we observed that:

- isosteric substitution of the ring oxygen atoms with sulphur favours 5-HT_{1A}R affinity, potency and efficacy especially in the presence of a more flexible amine chain;

- compounds **14** and **15** emerged as novel 5-HT_{1A}R partial agonists, the first being outstanding for selectivity (5-HT_{1A}/ α_{1d} = 80), the latter for potency (pD₂ = 9.58) and efficacy (E_{max} = 74%). Compared to the lead compound **1**, **15** exhibited a 10-fold improved potency and about 50% enhanced efficacy.
- compound **15** demonstrated to permeate the BBB by passive diffusion and showed a promising neuroprotective activity *in vitro*.
- In formalin test compound **15** reduces significantly the linking time in Phase II at a dose of 10mg/Kg i.p. indicating a potent analgesic activity and suggesting an additional and new strategy for pain control.

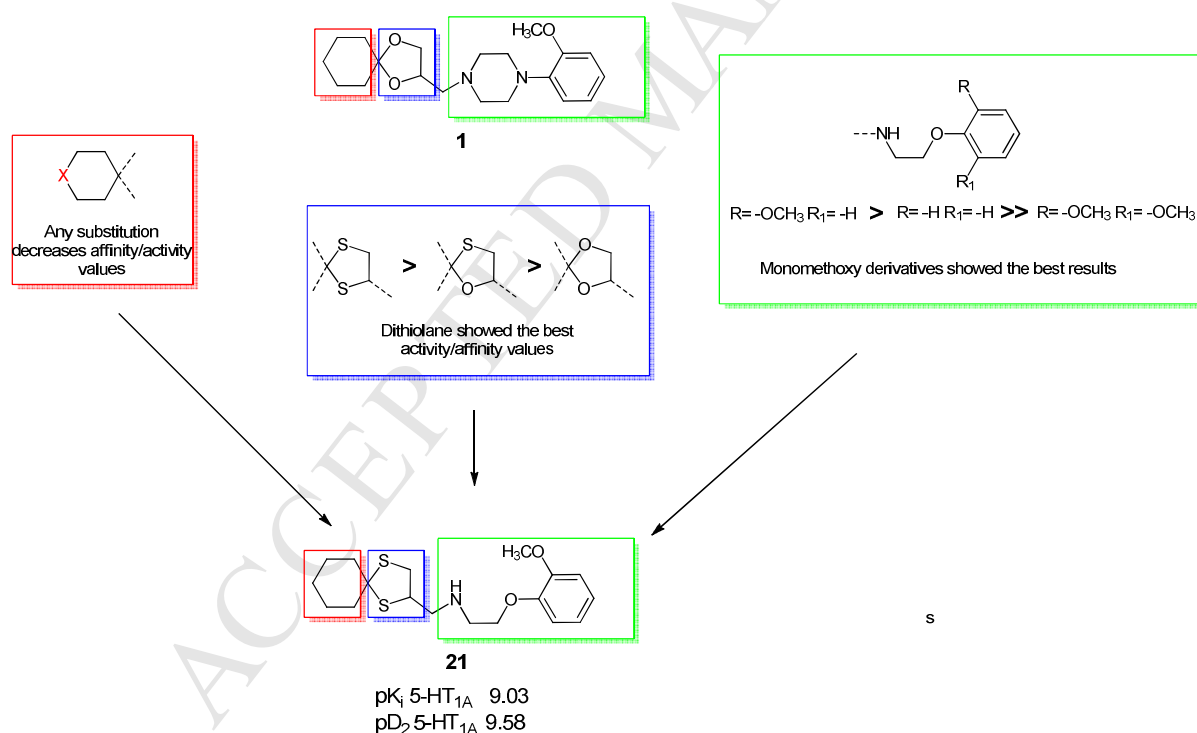


Figure 14. SAR milestones of compound **1**

Experimental Section

Materials and methods

All reagents, solvents and other chemicals were used as purchased from Sigma-Aldrich without further purification unless otherwise specified. Air- or moisture-sensitive reactants and solvents were employed in reactions carried out under nitrogen atmosphere unless otherwise noted. Flash column chromatography purifications (medium pressure liquid chromatography) were carried out using Merck silica gel 60 (230-400 mesh, ASTM). The purity of compounds was determined by elemental analysis (C,H,N) on a Carlo Erba 1106 Analyzer in the Microanalysis Laboratory of the Life Sciences Department of Università degli Studi di Modena e Reggio Emilia and the results are within $\pm 0.4\%$ of the theoretical values. Melting points were determined with a Stuart SMP3 in open capillaries and they are uncorrected. The structures of all isolated compounds were confirmed by Nuclear magnetic resonance (NMR) and Mass spectrometry. ^1H and ^{13}C NMR (1D and 2D experiments) spectra were recorded on a DPX-200/400 Avance (Bruker) spectrometer at 200 MHz and 400 MHz respectively and on AVANCE III (Bruker Biospin) at 600 MHz. Chemical shifts are expressed in δ (ppm). ^1H NMR chemical shifts are relative to tetramethylsilane (TMS) as internal standard. ^{13}C NMR chemical shifts are relative to TMS at δ 0.0 or to the ^{13}C signal of the solvent: CDCl_3 δ 77.04, CD_3OD δ 49.8, DMSO-d_6 δ 39.5. NMR data are reported as follows: chemical shift, number of protons/carbons, multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broadened), coupling constants (Hz) and assignment (Dosd= 1,4-dioxaspiro[4.5]decane; Ph= phenyl; Otsd= 1-oxa-4-thiaspiro[4.5]decane, Ts= tosyl; Arom= aromatic; Piperaz= piperazine; Dtsd= 1,4-dithiaspiro[4.5]decane). ^1H - ^1H correlation spectroscopy (COSY), ^1H - ^{13}C heteronuclear multiple quantum coherence (HMQC) and heteronuclear multiple bond connectivity (HMBC)

experiments were recorded for determination of ^1H - ^1H and ^1H - ^{13}C correlations respectively. Mass spectra were obtained on a hybrid QTOF mass spectrometer (PE SCIEX-QSTAR) using electrospray ionization mode (HR-ESI-MS, ion voltage of 4800 V). The HPLC experimental conditions of the HPLC-MS system are: flow rate 5 ml/min, sample solution (10 pmol/ml) of the selected compound with 0.1% acetic acid, mobile phase consisting of methanol (50%) and water (50%). The yields reported are based on a single experiment and are not optimized. The oxalate salts of all tested compounds were used for the pharmacological evaluations.

The compounds **4** and **7** were obtained as previously reported [40].

1-oxa-4-thiaspiro[4.5]decan-2-ylmethanol (2)

A round-bottom flask was charged with cyclohexanone (2 g, 20.0 mmol), 3-mercaptopropane-1,2-diol (2.60 g, 24.0 mmol) and (0.5 mmol/g) $\text{HClO}_4/\text{SiO}_2$ (0.4 g). The reaction mixture was stirred at room temperature, under nitrogen, for 6 hours. Then the reaction mixture was diluted with EtOAc and then filtered and evaporated under reduced pressure. Purification by flash chromatography (85/15 Cy/EtAc) afforded the title compound as brown oil [47].

Yield 3.05 g (16.0 mmol, 81%). ^1H NMR (400 MHz, CDCl_3): δ 1.18-1.33 (m, 1H, CHa-8 Otsd), 1.36-1.57 (m, 3H, CHb-8, CHa-7, CHa-9 Otsd), 1.65-1.99 (m, 6H, CHb-7, CHb-9, CH_2 -6, CH_2 -10 Otsd), 2.92 (dd, J = 9.1, 10.3 Hz, 1H, CHa-3 Otsd), 2.98 (dd, J = 5.4, 10.3 Hz, 1H, CHb-3 Otsd), 3.68 (dd, J = 5.3, 11.6 Hz, 1H, CHaOH), 3.81 (dd, J = 3.5, 11.6 Hz, 1H, CHbOH), 4.28-4.39 (m, 1H, CH-2 Otsd). ^{13}C NMR (100 MHz, CDCl_3): δ 23.9 (CH_2 , C-8 Otsd), 24.7 (CH_2 , C-7/C-9 Otsd), 25.0 (CH_2 , C-7/C-9 Otsd), 33.1 (CH_2 , C-3 Otsd), 39.7 (CH_2 , C-6/C-10 Otsd), 40.1 (CH_2 , C-6/C-10 Otsd), 63.3 (CH_2 , CH_2OH), 81.3 (CH, C-2 Otsd), 96.4 (C, C-5 Otsd).

1,4-dithiaspiro[4.5]decan-2-ylmethanol (3)

The title compound [47] was obtained from cyclohexanone and 2,3-dimercaptopropan-1-ol following the procedure described for **2**.

Yield 2.90 g (14.0 mmol, 71%). ¹H NMR (400 MHz, CDCl₃) δ 1.22-1.39 (m, 2H, CH₂-8 Dtsd), 1.33-1.79 (m, 4H, CH₂-7, CH₂-9 Dtsd), 1.81-2.14 (m, 4H, CH₂-6, CH₂-10 Dtsd), 3.26-3.41 (m, 2H, CH₂-3 Dtsd), 3.62 (dd, J = 5.1, 11.7 Hz, 1H, CHaOH), 3.77 (dd, J = 3.6, 11.7 Hz, 1H, CHbOH), 3.84-3.96 (m, 1H, CH-2 Dtsd). ¹³C NMR (100 MHz, CDCl₃): δ 22.6 (CH₂, C-8 Dtsd), 25.3 (CH₂, C-7/C-9 Dtsd), 25.9 (CH₂, C-7/C-9 Dtsd), 35.3 (CH₂, C-3 Dtsd), 37.2 (CH₂, C-6/C-10 Dtsd), 37.6 (CH₂, C-6/C-10 Dtsd), 52.3 (CH, C-2 Dtsd), 58.6 (C, C-5 Dtsd), 61.9 (CH₂, CH₂OH).

1-oxa-4-thiaspiro[4.5]decan-2-ylmethyl 4-methylbenzenesulfonate (5)

To a solution of **2** (1.5 g, 7.98 mmol) and Et₃N (1.11 ml, 7.98 mmol) in CH₂Cl₂ (10 mL) tosyl chloride (2.43 g, 18.0 mmol) was added at 0°C. The resulting mixture was stirred at room temperature, under nitrogen for 6 h and then the solvent was evaporated. The residue was taken up with EtOAc and washed with saturated solution sodium bicarbonate and brine. The organic layer was dried over anhydrous sodium sulfate, filtered and concentrated in vacuum. Purification by flash chromatography (gradient from 99/1 to 70/30 Cy/EtAc) afforded the title compound as yellow oil.

Yield 2.25 g, (6.57 mmol, 82%). ¹H NMR (400 MHz, CDCl₃): δ 1.22-1.36 (m, 1H, CHa-8 Otsd), 1.36-1.57 (m, 3H, CHb-8, CHa-7, CHa-9 Otsd), 1.63-1.94 (m, 6H, CHb-7, CHb-9, CH₂-6, CH₂-10 Otsd), 2.49 (s, 3H, CH₃ Ts), 2.83 (dd, J = 8.0, 10.7 Hz, 1H, CHa-3 Otsd), 3.09 (dd, J = 5.3, 10.7 Hz, 1H, CHb-3 Otsd), 4.09-4.22 (m, 2H, CH₂OH), 4.39-4.51 (m, 1H, CH-2 Otsd), 7.39 (d, J = 8.1 Hz, 2H, CH-3, CH-5 Ts), 7.84 (d, J = 8.1 Hz, 2H, CH-2, CH-6 Ts). ¹³C NMR (100 MHz, CDCl₃):

δ 21.4 (CH₃, CH₃ Ts), 24.0 (CH₂, C-8 Otsd), 24.7 (CH₂, C-7/C-9 Otsd), 25.0 (CH₂, C-7/C-9 Otsd), 33.9 (CH₂, C-3 Otsd), 39.7 (CH₂, C-6/C-10 Otsd), 40.1 (CH₂, C-6/C-10 Otsd), 69.6 (CH₂, CH₂OH), 78.0 (CH, C-2 Otsd), 96.9 (C, C-5 Otsd), 127.7 (2 CH, C-2, C-6 Ts), 129.6 (2 CH, C-3, C-5 Ts), 132.5 (C, C-1 Ts), 147.7 (C, C-4 Ts). HRMS-APCI m/z [M+H]⁺ calcd for C₁₆H₂₃O₄S₂⁺: 343.1032; found 343.1032.

2-(chloromethyl)-1,4-dithiaspiro[4.5]decane (6)

To a solution of **3** (2.9 g, 14.2 mmol), in toluene (15 mL) thionyl chloride (1.35 mL, 18.5 mmol) was added at 0°C. The resulting mixture was stirred at 80 °C for 12 h and then the solvent was evaporated. The residue solubilized in EtOAc was washed with saturated solution of sodium bicarbonate and brine. The organic phase was dried over anhydrous sodium sulfate, filtered and evaporated. Purification by flash chromatography (99/1 Cy/EtAc) afforded the title compound as dark oil.

Yield 0.93 g (4.19 mmol, 30%). ¹H NMR (400 MHz, CDCl₃): δ 1.31-1.52 (m, 2H, CH₂-8 Dtsd), 1.57-1.84 (m, 4H, CH₂-7, CH₂-9 Dtsd), 1.94-2.07 (m, 4H, CH₂-6, CH₂-10 Dtsd), 3.39 (dd, J = 4.6, 12.5 Hz, 1H, CHa-3 Dtsd), 3.47 (dd, J = 2.6, 12.5 Hz, 1H, CHb-3 Dtsd), 3.54 (dd, J = 3.7, 10.3 Hz, 1H, CHaCl), 3.84 (dd, J = 10.3, 10.9 Hz, 1H, CHbCl), 3.88-3.97 (m, 1H, CH-2 Dtsd). ¹³C NMR (100 MHz, CDCl₃): δ 24.6 (CH₂, C-8 Dtsd), 25.2 (CH₂, C-7/C-9 Dtsd), 26.5 (CH₂, C-7/C-9 Dtsd), 38.9 (CH₂, C-3 Dtsd), 42.0 (CH₂, C-6/C-10 Dtsd), 43.0 (CH₂, C-6/C-10 Dtsd), 45.1 (CH₂, CH₂Cl), 55.0 (CH, C-2 Dtsd), 69.1 (C, C-5 Dtsd). HRMS-APCI m/z [M+H]⁺ calcd for C₉H₁₆ClS₂⁺: 223.0376; found 223.0378.

General procedure for the synthesis of the amines 8-15.

A 10 mL microwave vial was charged with **4** or **5** or **6** (1.0 mmol), a small excess (1.2 mmol) of 2-phenoxy- or (2-methoxyphenoxy)- or (2,6-dimethoxyphenoxy)-ethanamine and a catalytic amount of potassium iodide in 1 mL of 2-methoxyethanol. The reaction was stirred under microwave irradiation at 160 °C (pressure 100 PSI, power 50 W) for 30 min. Then the solvent was evaporated under reduced pressure. The residue was taken up with EtOAc, basified with 5% NaOH. The organic layer was dried over anhydrous sodium sulfate, filtered and evaporated under reduced pressure. The residue was purified by flash column chromatography.

N-(1,4-dioxaspiro[4.5]decan-2-ylmethyl)-2-(2-methoxyphenoxy)ethanamine (8)

The title compound was purified by flash chromatography on silica gel cartridge (80/20/5 Cy/EtAc/MeOH + 2 NH₄OH) to afford 0.111 g (0.34 mmol, 54%) of **8** as yellow oil.

¹H NMR (400 MHz, CDCl₃): δ 1.31-1.52 (m, 2H, CH₂-8 Dosd), 1.49-1.79 (m, 8H, CH₂-7, CH₂-9, CH₂-6, CH₂-10 Dosd), 2.66-2.95 (m, 2H, CH₂N), 3.07 (t, *J* = 5.5 Hz, 2H, CH₂CH₂O), 3.69 (dd, *J* = 7.4, 7.9 Hz, 1H, CHa-3 Dosd), 3.86 (s, 3H, OCH₃), 4.01-4.19 (m, 3H, CHb-3 Dosd, CH₂O), 4.19-4.36 (m, 1H, CH-2 Dosd), 6.73-7.08 (m, 4H, CH-3, CH-4, CH-5, CH-6 Ph). ¹³C NMR (100 MHz, CDCl₃): δ 23.6 (CH₂, C-8 Dosd), 24.1 (CH₂, C-7/C-9 Dosd), 25.2 (CH₂, C-7/C-9 Dosd), 34.3 (CH₂, C-6/C-10 Dosd), 36.4 (CH₂, C-6/C-10 Dosd), 48.1 (CH₂, CH₂CH₂O), 51.7 (CH₂, CHCH₂N), 67.0 (CH₂, C-3 Dosd), 73.0 (CH₂, CH₂O), 74.6 (CH, C-2 Dosd), 109.5 (C, C-5 Dosd), 111.9 (CH, C-3 Ph), 114.4 (CH, C-5 Ph), 120.6 (CH, C-6 Ph), 121.7 (CH, C-4 Ph), 148.1 (C, C-1 Ph), 149.7 (C, C-2 Ph).

The free amine (0.100 g, 0.31 mmol) was dissolved in Et₂O and treated with 1.2 eq. of oxalic acid to give 0.083 g (0.20 mmol, 64% yield) of the corresponding oxalate salt.

Mp: 201-203°C. ^1H NMR (600 MHz, $\text{DMSO}-d_6$): δ 1.25-1.44 (m, 2H, CH_2 -8), 1.45-1.68 (m, 8H, CH_2 -7, CH_2 -9 CH_2 -6, CH_2 -10 Dosd), 3.13 (dd, J = 2.8, 4.2 Hz, 1H, CHCH_aN) 3.18-3.29 (m, 3H, CHCH_bN , $\text{CH}_2\text{CH}_2\text{O}$), 3.74 (dd, J = 1.8, 2.8 Hz, 1H, CH_a -3 Dosd), 3.80 (s, 3H, OCH_3), 4.05-4.14 (m, 3H, CH_2O , CH_b -3 Dosd), 4.37-4.44 (m, 1H, CH -2 Dosd), 6.87-7.05 (m, 4H, CH -3, CH -4, CH -5, CH -6 Ph). HRMS-ESI m/z $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{18}\text{H}_{28}\text{NO}_4^+$: 322.2013; found 322.2016. Anal. calcd for $\text{C}_{20}\text{H}_{29}\text{NO}_8$: C 58.38, H 7.10, N 3.40; found: C 58.62, H 7.27, N 3.49.

N-(1,4-dioxaspiro[4.5]decan-2-ylmethyl)-2-(2,6-dimethoxyphenoxy)ethanamine (9)

The title compound was purified by flash column chromatography on silica gel (gradient from 30/70 to 100 Cy/EtAc) to afford 0.165 g (0.47 mmol, 54%) of **9** as oil.

^1H NMR (400 MHz, CDCl_3): δ 1.34-1.50 (m, 2H, CH_2 -8 Dosd), 1.52-1.82 (m, 8H, CH_2 -7, CH_2 -9, CH_2 -6, CH_2 -10 Dosd), 2.91 (d, J = 5.9 Hz, 2H, CHCH_2N), 3.02 (t, J = 5.4 Hz, 2H, $\text{CH}_2\text{CH}_2\text{O}$), 3.74 (dd, J = 6.3, 8.1 Hz, CH_a -3 Dosd), 3.90 (s, 6H, 2 x OCH_3), 4.06-4.19 (m, 3H, CH_b -3 Dosd, CH_2O), 4.35-4.47 (m, 1H, CH -2 Dosd), 6.61 (d, J = 7.4 Hz, 2H, CH -3, CH -5 Ph), 7.04 (t, J = 7.4 Hz, CH -4 Ph). ^{13}C NMR (100 MHz, CDCl_3): δ = 23.5 (CH_2 , C-8 Dosd), 23.7 (CH_2 , C-7/C-9 Dosd), 24.9 (CH_2 , C-7/C-9 Dosd), 34.7 (CH_2 , C-6/C-10 Dosd), 36.4 (CH_2 , C-6/C-10 Dosd), 49.3 (CH_2 , $\text{CH}_2\text{CH}_2\text{O}$), 52.1 (CH_2 , CHCH_2N), 55.8 (2 CH_3 , OCH_3), 67.0 (CH_2 , C-3 Dosd), 71.4 (CH_2 , CH_2O), 74.3 (CH , C-2 Dosd), 104.9 (2 CH , C-3, C-5 Ph), 109.7 (C, C-5 Dosd), 123.7 (CH , C-4 Ph), 136.3 (C, C-1 Ph), 153.3 (2 C, C-2, C-6 Ph).

The free amine (0.150 g, 0.42 mmol) was dissolved in Et_2O and treated with 1.2 eq. of oxalic acid to give 0.108 g (0.24 mmol, 58% yield) of the corresponding oxalate salt.

Mp: 182-184°C. ^1H NMR (600 MHz, $\text{DMSO}-d_6$): δ 1.29-1.42 (m, 2H, CH_2 -8), 1.47-1.63 (m, 8H, CH_2 -7, CH_2 -9 CH_2 -6, CH_2 -10 Dosd), 3.10 (dd, J = 2.9, 4.3 Hz, 1H, CHCH_aN) 3.20-3.30 (m, 3H,

CHCHbN, CH₂CH₂O), 3.74 (dd, J = 1.9, 2.9 Hz, 1H, CHa-3 Dosd), 3.79 (s, 6H, OCH₃), 4.06-4.13 (m, 3H, CH₂O, CHb-3 Dosd), 4.41 (m, 1H, CH-2 Dosd), 6.71 (d, 2H, J = 8.5 Hz, CH-3, CH-5 Ph), 7.06 (t, J = 8.3 Hz, 1H, CH-4Ph). HRMS-ESI m/z [M+H]⁺ calcd for C₁₉H₃₀NO₅⁺: 352.2118; found 352.2126. Anal. calcd for C₂₁H₃₁NO₉: C 57.13, H 7.08, N 3.17; found: C 57.42, H 7.31, N 3.44.

1-(1-oxa-4-thiaspiro[4.5]decan-2-ylmethyl)-4-(2-methoxyphenyl)piperazine (10)

The title compound was purified by flash chromatography on silica gel cartridge (80/20 Cy/EtAc) to afford 0.178 g (0.49 mmol, 67% of **10** as oil.

¹H NMR (400 MHz, CDCl₃): δ 1.23-1.41 (m, 1H, CHa-8 Otsd), 1.42-1.60 (m, 3H, CHb-8, CHa-7, CHa-9 Otsd), 1.67-2.04 (m, 6H, CHb-7, CHb-9, CH₂-6, CH₂-10 Otsd), 2.67-2.97 (m, 7H, CH₂-N, CH₂-2, CH₂-6 Piperaz, CHa-3 Otsd), 3.08-3.27 (m, 5H, CH₂-3, CH₂-5 piperaz, CHb-3 Otsd), 3.90 (s, 3H, OCH₃), 4.43-4.56 (m, 1H, CH-2 Otsd), 6.90 (d, J = 7.9 Hz, 1H, CH-3 Arom), 6.93-7.02 (m, 2H, CH-5, CH-6 Arom), 7.04-7.16 (m, 1H, CH-4 Arom). ¹³C NMR (100 MHz, CDCl₃): δ 24.1 (CH₂, C-8 Otsd), 24.8 (CH₂, C-7/C-9 Otsd), 25.1 (CH₂, C-7/C-9 Otsd), 36.4 (CH₂, C-3 Otsd), 39.9 (CH₂, C-6/C-10 Otsd), 40.5 (CH₂, C-6/C-10 Otsd), 50.2 (2 CH₂, C-3, C-5 Piperaz), 53.8 (2 CH₂, C-2, C-6 Piperaz), 55.1 (CH₃, OCH₃), 61.4 (CH₂, CH₂N), 79.2 (CH, C-2 Otsd), 96.4 (C, C-5 Otsd), 110.9 (CH, C-3 Arom), 117.9 (CH, C-5 Arom), 120.7 (CH, C-6 Arom), 122.7 (CH, C-4 Arom), 141.0 (C, C-1 Arom), 152.0 (C, C-2 Arom).

The free amine (0.178 g, 0.49 mmol) was dissolved in Et₂O and treated with 1.2 eq. of oxalic acid to give 0.174 g (0.38 mmol, 79% yield) of the corresponding oxalate salt.

Mp: 217-218°C. ¹H NMR (600 MHz, DMSO-*d*₆): δ 1.25-1.48 (m, 4H, CH₂-8, CHa-7, CHa-9 Otsd), 1.62-1.94 (m, 6H, CHb-7, CHb-9, CH₂-6, CH₂-10 Otsd), 2.81 (dd, J = 9.4, 10.3 Hz, 1H,

CHa-3 Otsd), 3.11-3.32 (m, 11H, CH₂-2, CH₂-3, CH₂-5, CH₂-6 Piperaz, CHb-3 Otsd, CH₂N), 3.79 (s, 3H, OCH₃), 4.55-4.62 (m, 1H, CH-2), 6.87-7.02 (m, 4H, Arom). HRMS-ESI *m/z* [M+H]⁺ calcd for C₂₀H₃₁N₂O₂S⁺: 363.2106; found 363.2103. Anal. calcd for C₂₂H₃₂N₂O₆S: C 58.39, H 7.13, N 6.19; found: C 58.44, H 7.27, N 6.24.

{1-oxa-4-thiaspiro[4.5]decan-2-ylmethyl}(2-phenoxyethyl)amine (11)

The title compound was purified by flash chromatography on silica gel cartridge (80/20 Cy/EtAc) to afford 0.056 g (0.18 mmol, 25%) of **11** as yellow oil.

¹H NMR (400 MHz, CDCl₃): δ 1.21-1.39 (m, 1H, CHa-8 Otsd), 1.43-1.61 (m, 3H, CHb-8, CHa-7, CHa-9 Otsd), 1.73-2.01 (m, 6H, CHb-7, CHb-9, CH₂-6, CH₂-10 Otsd), 2.85 (dd, *J* = 9.2, 10.0 Hz, 1H, CHa-3 Otsd), 2.91-3.06 (m, 2H, CHCH₂NH), 3.09 (dd, *J* = 5.0, 10.0 Hz, 1H, CHb-3 Otsd), 3.14 (d, *J* = 5.3 Hz, 2H, NHCH₂CH₂), 4.09 (t, *J* = 5.3 Hz, 2H, CH₂O), 4.42-4.51 (m, 1H, CH-2 Otsd), 6.82-7.12 (m, 3H, CH-2, CH-4, CH-6 Ph), 7.21-7.48 (m, 2H, CH-3, CH-5 Ph). ¹³C NMR (100 MHz, CDCl₃): δ 24.0 (CH₂, C-8 Otsd), 24.8 (CH₂, C-7/C-9 Otsd), 25.1 (CH₂, C-7/C-9 Otsd), 35.3 (CH₂, C-3 Otsd), 39.9 (CH₂, C-6/C-10 Otsd), 40.4 (CH₂, C-6/C-10 Otsd), 48.7 (CH₂, NCH₂CH₂), 52.6 (CH₂, CHCH₂N), 66.8 (CH₂, CH₂O), 80.4 (CH, C-2 Otsd), 96.3 (C, C-5 Otsd), 114.3 (2 CH, C-2, C-6 Ph), 120.6 (CH, C-4 Ph), 129.2 (2 CH, C-3, C-5 Ph), 158.5 (C, C-1 Ph).

The free amine (0.057 g, 0.186 mmol) was dissolved in Et₂O and treated with 1.2 eq. of oxalic acid to give 0.067 g (0.169 mmol, 91% yield) of the corresponding oxalate salt.

Mp: 210-211°C. ¹H NMR (600 MHz, DMSO-*d*₆) δ 1.20-1.52 (m, 4H, CH₂-8, CHa-7, CHa-9 Otsd), 1.64-1.93 (m, 6H, CHb-7, CHb-9, CH₂-6, CH₂-10 Otsd), 2.83 (dd, *J* = 8.5, 10.6 Hz, 1H, CHa-3 Otsd), 3.14-3.22 (m, 2H, CHb-3 Otsd, CHCH₂N), 3.31-3.42 (m, 3H, CHCH₂N,

NCH_2CH_2), 4.25 (t, $J=4.9$ Hz, 2H, CH_2O), 4.52 (m, 1H, CH-2 Otsd), 6.99 (m, 3H, CH-2, CH-4, CH-6 Ph), 7.32 (m, 2H, CH-3, CH-5 Ph). HRMS-ESI m/z $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{17}\text{H}_{26}\text{NO}_2\text{S}^+$: 308.1679; found 308.1675. Anal. calcd for $\text{C}_{19}\text{H}_{27}\text{NO}_6\text{S}$: C 57.41, H 6.85, N 3.52, found: C 57.62, H 6.94, N 3.66.

N-(1-oxa-4-thiaspiro[4.5]decan-2-ylmethyl)-2-(2-methoxyphenoxy)ethanamine (12)

The title compound was purified by flash chromatography on silica gel cartridge (70/30/10 Cy/EtAc/MeOH + 1 NH_4OH) to afford 0.046 g (0.14 mmol, 19%) of **12** as yellow oil.

^1H NMR (400 MHz, CDCl_3): δ 1.23-1.38 (m, 1H, CHa-8 Otsd), 1.41-1.58 (m, 3H, CHb-8, CHa-7, CHa-9 Otsd), 1.69-2.00 (m, 6H, CHb-7, CHb-9, CH_2 -6, CH_2 -10 Otsd), 2.86 (dd, $J=9.1, 9.9$ Hz, 1H, CHa-3 Otsd), 3.01 (d, $J=5.4$ Hz, 2H, CHCH_2N), 3.09 (dd, $J=5.0, 9.9$ Hz, 1H, CHb-3), 3.15 (dt, $J=1.3, 5.2$ Hz, 2H, NCH_2CH_2), 3.90 (s, 3H, OCH_3), 4.19 (t, $J=5.2$ Hz, 2H, CH_2O), 4.39-4.50 (m, 1H, CH-2 Otsd), 6.72-7.04 (m, 4H, Ph). ^{13}C NMR (100 MHz, CDCl_3): δ 24.0 (CH_2 , C-8 Otsd), 24.8 (CH_2 , C-7/C-9 Otsd), 25.1 (CH_2 , C-7/C-9 Otsd), 35.3 (CH_2 , C-3 Otsd), 39.9 (CH_2 , C-6/C-10 Otsd), 40.4 (CH_2 , C-6/C-10 Otsd), 48.5 (CH_2 , NCH_2CH_2), 52.5 (CH_2 , CHCH_2N), 55.6 (CH_3 , OCH_3), 68.4 (CH_2 , CH_2O), 80.2 (CH, C-2 Otsd), 96.3 (C, C-5 Otsd), 111.8 (CH, C-3 Ph), 114.3 (CH, C-5 Ph), 120.7 (CH, C-6 Ph), 121.5 (CH, C-4 Ph), 148.0 (C, C-1 Ph), 149.6 (C, C-2 Ph).

The free amine (0.046 g, 0.137 mmol) was dissolved in Et_2O and treated with 1.2 eq. of oxalic acid to give 0.048 g (0.159 mmol, 83% yield) of the corresponding oxalate salt.

Mp: 202-203°C. ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 1.23-1.48 (m, 4H, CH_2 -8, CHa-7, CHa-9 Otsd), 1.66-1.89 (m, 6H, CHb-7, CHb-9, CH_2 -6, CH_2 -10 Otsd), 2.82 (dd, $J=9.2, 10.0$ Hz, 1H, CHa-3 Otsd), 3.17 (dd, $J=4.5, 10.0$ Hz, 1H, CHb-3 Otsd), 3.24 (dd, $J=8.1, 12.4$ Hz, 1H,

CHCH_aN), 3.31-3.40 (m, 3H, CHCH_bN, NCH₂CH₂), 3.78 (s, 3H, OCH₃), 4.23 (t, J = 5.2 Hz, 2H, CH₂O), 4.52 (m, 1H, CH-2 Otsd), 6.88-7.05 (m, 4H, Ph). HRMS-ESI m/z [M+H]⁺ calcd for C₁₈H₂₈NO₃S⁺: 338.1784; found 338, 1797. Anal. calcd for C₂₀H₂₉NO₇S: C 56.19, H 6.84, N 3.28; found: C 56.28, H 6.99, N 3.37.

1-{1,4-dithiaspiro[4.5]decan-2-ylmethyl}-4-(2-methoxyphenyl)piperazine (13)

The title compound was purified by flash chromatography on silica gel cartridge (90/10 Cy/EtAc) to afford 0.136 g (0.36 mmol, 53%) of **13** as dark oil.

¹H NMR (400 MHz, CDCl₃) δ = 1.33-1.51 (m, 2H, CH₂-8 Dtsd), 1.53-1.81 (m, 4H, CH₂-7, CH₂-9 Dtsd), 1.92-2.09 (m, 4H, CH₂-6, CH₂-10 Dtsd), 2.47-2.77 (m, 7H, CH₂-N, CH₂-2, CH₂-6 Piperaz, CH_a-3 Dtsd), 3.01-3.25 (m, 5H, CH₂-3, CH₂-5 Piperaz, CH_b-3 Dtsd), 3.25-3.47 (m, 2H, CH₂-3 Dtsd), 3.85 (m, 3H, OCH₃), 3.87-3.99 (m, 1H, CH-2 Dtsd), 6.90 (d, J = 8.0 Hz, 1H, CH-3 Arom), 6.91-7.05 (m, 2H, CH-5, CH-6 Arom), 7.06-7.15 (m, 1H, CH-4 Arom). ¹³C NMR (100 MHz, CDCl₃): δ 24.1 (CH₂, C-8 Dtsd), 25.0 (CH₂, C-7/C-9 Dtsd), 26.3 (CH₂, C-7/C-9 Dtsd), 38.1 (CH₂, C-3 Dtsd), 41.9 (CH₂, C-6/C-10 Dtsd), 42.5 (CH₂, C-6/C-10 Dtsd), 50.1 (2 CH₂, C-3, C-5 Piperaz), 53.7 (2 CH₂, C-2, C-6 Piperaz), 55.1 (CH₃, OCH₃), 55.2 (CH, C-2 Dtsd), 61.2 (CH₂, CH₂N), 69.3 (C, C-5 Dtsd), 111.0 (CH, C-3 Arom), 117.8 (CH, C-5 Arom), 120.6 (CH, C-6 Arom), 122.6 (CH, C-4 Arom), 141.0 (C, C-1 Arom), 151.9 (C, C-2 Arom).

The free amine (0.136 g, 0.36 mmol) was dissolved in Et₂O and treated with 1.2 eq. of oxalic acid to give 0.128 g (0.27 mmol, 76% yield) of the corresponding oxalate salt.

Mp: 170-172°C. ¹H NMR (600 MHz, DMSO-*d*₆) δ 1.31-1.43 (m, 2H, CH₂-8 Dtsd), 1.48-1.66 (m, 4H, CH₂-7, CH₂-9 Dtsd), 1.86-2.07 (m, 4H, CH₂-6, CH₂-10 Dtsd), 2.64-3.42 (m, 12H, CH₂-2, CH₂-3, CH₂-5, CH₂-6 Piperaz, CH₂N, CH₂-3 Dtsd), 3.78 (s, 3H, OCH₃), 4.09 (m, 1H, CH-2

Dtsd), 6.83-7.03 (m, 4H, Arom). HRMS-ESI m/z $[M+H]^+$ calcd for $C_{20}H_{31}N_2OS_2^+$: 379.1872; found 379.1872. Anal. calcd for $C_{22}H_{32}N_2O_5S_2$: C 56.38, H 6.88, N 5.98; found: C 56.48, H 7.02, N 6.11.

{1,4-dithiaspiro[4.5]decan-2-ylmethyl}(2-phenoxyethyl)amine (14)

The title compound was purified by flash chromatography on silica gel cartridge (70/30 Cy/EtAc) to afford 0.032 g (0.1 mmol, 15%) of **14** as yellow oil.

1H NMR (400 MHz, $CDCl_3$) δ 1.18-1.49 (m, 6H, CH_2 -7, CH_2 -8, CH_2 -9 Dtsd), 1.64 (m, 4H, CH_2 -6, CH_2 -10 Dtsd), 2.95 (dd, J = 7.4, 12.2 Hz, 1H, CHaNH), 3.02 (dd, J = 6.9, 12.2 Hz, 1H, CHbNH), 3.05 (t, J = 5.2 Hz, 2H, $NHCH_2CH_2$), 3.19 (dd, J = 5.6, 12.0 Hz, 1H, CHa-3 Dtsd), 3.35 (dd, J =12.0, 5.1 Hz, 1H, CHb-3 Dtsd), 3.83-3.95 (m, 1H, CH-2 Dtsd), 4.08 (t, J = 5.3 Hz, 2H, CH_2O), 6.81-7.08 (m, 3H, CH-2, CH-4, CH-6 Ph), 7.21-7.39 (m, 2H, CH-3, CH-5 Ph). ^{13}C NMR (100 MHz, $CDCl_3$): δ 24.3 (CH_2 , C-8 Dtsd), 24.9 (CH_2 , C-7/C-9 Dtsd), 25.1 (CH_2 , C-7/C-9 Dtsd), 38.4 (CH_2 , C-3 Dtsd), 40.9 (CH_2 , C-6/C-10 Dtsd), 41.6 (CH_2 , C-6/C-10 Dtsd), 48.4 (CH_2 , NCH_2CH_2), 50.5 (CH, C-2 Dtsd), 52.4 (CH_2 , $CHCH_2N$), 68.2 (CH_2 , CH_2O), 69.0 (C, C-5 Dtsd), 114.2 (2 CH, C-2, C-6 Ph), 120.7 (CH, C-4 Ph), 129.2 (2 CH, C-3, C-5 Ph), 158.6 (C, C-1 Ph).

The free amine (0.032 g, 0.099 mmol) was dissolved in acetone and treated with 1.2 eq. of oxalic acid to give 0.038 g (0.092 mmol, 92% yield) of the corresponding oxalate salt.

Mp: 211-214°C. 1H NMR (600 MHz, $DMSO-d_6$) δ 1.34-1.41 (m, 2H, CH_2 -8 Dtsd), 1.50-1.65 (m, 4H, CH_2 -7, CH_2 -9 Dtsd), 1.89-2.02 (m, 4H, CH_2 -6, CH_2 -10 Dtsd), 3.16 (dd, J = 5.3, 12.3 Hz, 1H, CHa-3 Dtsd), 3.27-3.45 (m, 5H, CH_2N , CH_2CH_2O , CHb-3 Dtsd), 4.07 (m, 1H, CH-2 Dtsd), 4.20 (t, J = 5.2 Hz, 2H, CH_2O), 6.93-7.01 (m, 3H, CH-2, CH-4, CH-6 Ph), 7.21-7.33 (m, 2H, CH-3,

CH-5 Ph). HRMS-ESI m/z $[M+H]^+$ calcd for $C_{17}H_{26}NOS_2^+$: 324.1449; found 324.1450. Anal. calcd for $C_{19}H_{27}NO_5S_2$: C 55.18, H 6.58, N 3.39; found: C 55.33, H 6.71, N 3.57.

{1,4-dithiaspiro[4.5]decan-2-ylmethyl}[2-(2-methoxyphenoxy)ethyl]amine (15)

The title compound was purified by flash chromatography on silica gel cartridge (50/50 Cy/EtAc) to afford 0.183 g (0.52 mmol, 47%) of **15** as dark oil.

1H NMR (400 MHz, $CDCl_3$) δ 1.21-1.40 (m, 2H, CH_2 -8 Dtsd), 1.49-1.69 (m, 4H, CH_2 -7, CH_2 -9 Dtsd), 1.83-2.14 (m, 4H, CH_2 -6, CH_2 -10 Dtsd), 2.89 (dd, J = 7.0, 12.1 Hz, 1H, $CHaNH$), 2.93-3.16 (m, 3H, $CHaNH$, $NHCH_2CH_2$), 3.19 (dd, J = 5.7, 12.0 Hz, 1H, CHa -3 Dtsd), 3.35 (dd, J = 5.1, 12.0 Hz, 1H, CHb -3 Dtsd), 3.81-3.92 (m, 4H, OCH_3 , CH -2 Dtsd), 4.16 (t, J = 5.3 Hz, 2H, CH_2O), 6.73-7.06 (m, 4H, Ph). ^{13}C NMR (100 MHz, $CDCl_3$): δ 24.2 (CH_2 , C-8 Dtsd), 24.8 (CH_2 , C-7/C-9 Dtsd), 25.4 (CH_2 , C-7/C-9 Dtsd), 37.9 (CH_2 , C-3 Dtsd), 40.8 (CH_2 , C-6/C-10 Dtsd), 41.4 (CH_2 , C-6/C-10 Dtsd), 48.5 (CH_2 , NCH_2CH_2), 50.3 (CH , C-2 Dtsd), 52.5 (CH_2 , $CHCH_2N$), 55.8 (CH_3 , OCH_3), 68.6 (CH_2 , CH_2O), 69.0 (C, C-5 Dtsd), 111.9 (CH, C-3 Ph), 114.3 (CH, C-5 Ph), 120.8 (CH, C-6 Ph), 121.5 (CH, C-4 Ph), 147.9 (C, C-1 Ph), 149.7 (C, C-2 Ph).

The free amine (0.183 g, 0.52 mmol) was dissolved in Et_2O and treated with 1.2 eq. of oxalic acid to give 0.216 g (0.486 mmol, 94% yield) of the corresponding oxalate salt.

Mp: 206-207°C. 1H NMR (600 MHz, $DMSO-d_6$) δ 1.34-1.44 (m, 2H, CH_2 -8 Dtsd), 1.48-1.65 (m, 4H, CH_2 -7, CH_2 -9 Dtsd), 1.86-2.01 (m, 4H, CH_2 -6, CH_2 -10 Dtsd), 3.18 (dd, J = 5.1, 12.5 Hz, 1H, CHa -3 Dtsd), 3.30-3.44 (m, 5H, CH_2N , CH_2CH_2O , CHb -3 Dtsd), 3.78 (s, 3H, OCH_3), 4.11 (m, 1H, CH -2 Dtsd), 4.21 (t, J = 5.4 Hz, 2H, CH_2O), 6.86-7.04 (m, 4H, Ph). HRMS-ESI m/z $[M+H]^+$ calcd for $C_{18}H_{28}NO_2S_2^+$: 354.116; found 354.1556. Anal. calcd for $C_{20}H_{29}NO_6S_2$: C 54.15, H 6.59, N 3.16; found: C 54.17, H 6.762, N 3.19.

Biological assays

Radioligand Binding Assay at Human Recombinant 5-HT_{1A}R and α_1 Adrenoceptor Subtypes.

A human cell line (HeLa) stably transfected with genomic clone G-21 coding for the human 5-HT_{1A} serotonergic receptor was used. Cells were grown as monolayers in Dulbecco's modified Eagle's medium supplemented with 10% fetal calf serum and gentamycin (100 μ g/mL) under 5% CO₂ at 37 °C. Cells were detached from the growth flask at 95% confluence by a cell scraper and were lysed in ice-cold Tris (5 mM) and EDTA buffer (5 mM, pH 7.4). Homogenates were centrifuged for 20 min at 40000g, and pellets were resuspended in a small volume of ice-cold Tris/EDTA buffer (above) and immediately frozen and stored at 70 °C until use. On the day of experiment, cell membranes (80-90 μ g of protein) were resuspended in binding buffer (50 mM Tris, 2.5 mM MgCl₂, and 10 mM pargiline, pH 7.4). Membranes were incubated in a final volume of 0.32 mL for 30 min at 30 °C with 1 nM [³H]8-OH-DPAT, in the absence or presence of various concentrations of the competing drugs (1 pM to 1 μ M); each experimental condition was performed in triplicate. Nonspecific binding was determined in the presence of 10 μ M 5-HT [42]. Binding to recombinant human α_1 adrenoceptor subtypes was performed in membranes from Chinese hamster ovary (CHO) cells transfected by electroporation with DNA expressing the gene encoding each α_1 adrenoceptor subtype. Cloning and stable expression of the human α_1 adrenoceptor genes were performed as described [41]. CHO cell membranes (70 μ g of protein) were incubated in 50 mM Tris (pH 7.4) with 0.1 – 0.4 nM [³H]prazosin, in a final volume of 0.32 mL for 30 min at 25 °C, in the absence or presence of competing drugs (1 pM to 1 μ M). Nonspecific binding was determined in the presence of 10 μ M Tamsulosin. The incubation was stopped by addition of ice-cold Tris buffer and rapid filtration through Unifilter B

filters (Perkin-Elmer) using a Filtermate cell harvester (Packard), and the radioactivity retained on the filters was determined by TopCount, Perkin-Elmer liquid scintillation counting at 90% efficiency.

[³⁵S]GTP γ S Binding Assay.

The effects of the various compounds tested on [³⁵S]GTP γ S binding in HeLa cells expressing the recombinant human 5-HT_{1A} receptor were evaluated according to the method of Stanton and Beer [43] with minor modifications [44,52]. Stimulation experiments: Cell membranes (50-70 μ g of protein) were resuspended in buffer containing 20 mM HEPES, 3 mM MgSO₄, and 120 mM NaCl (pH 7.4). The membranes were incubated with 30 μ M GDP, and various concentrations (from 0.01 nM to 10 μ M) of test drugs or 8-OH-DPAT (reference curve) for 20 min at 30 °C in a final volume of 0.5 mL. Samples were transferred to ice, [³⁵S]GTP γ S (200 pM) was added, and samples were incubated for another 30 min at 30 °C. The pre-incubation with both agonist and antagonist, before initiating the [³⁵S]GTP γ S binding, ensures that agonist and antagonist are at equilibrium. Nonspecific binding was determined in the presence of 10 μ M GTP γ S. Incubation was stopped by the addition of ice-cold HEPES buffer and rapid filtration on Unifilter B filters (Perkin Elmer) using a Filtermate cell harvester (Packard). The filters were washed with ice-cold Hepes buffer, and the radioactivity retained on the filters was determined by TopCount, Perkin Elmer liquid scintillation counting at 90% efficiency.

Data Analysis

Binding data were analyzed using the nonlinear curve-fitting program GraphPad (Prism for windows, version 5.04). Scatchard plots were linear for all preparations. None of the pseudo-Hill

coefficients (nH) were significantly different from unity ($p > 0.05$). Equilibrium dissociation constants (K_i) were derived from the Cheng–Prusoff equation $K_i = IC_{50}/(L/K_d)$, where L and K_d are the concentration and the equilibrium dissociation constant of the radioligand. pK_i values are the mean of 2–3 separate experiments performed in duplicate [61]. Stimulation of [35 S]GTP γ S binding induced by the compounds tested was expressed as the percent increase in binding above basal value, with the maximal stimulation observed with 8-OH-DPAT taken as 100%. The concentration–response curves of the agonistic activity were analyzed by GraphPad as reported above [62]. The maximum percentage of stimulation of [35 S]GTP γ S binding (E_{max}) achieved for each drug, and the concentration required to obtain 50% of E_{max} ($pD_2 = -\log_{10} [EC_{50}]$), were evaluated.

Cytotoxicity Assays

Cytotoxicity assays were carried out against human neuroblastoma cell line SH-SY5Y. Cells were cultured at 37 °C in a humidified incubator containing 5% CO₂ and feed with DMEM (Lonza) nutrient supplemented with 10% heat inactivated FBS, 2 mM L-glutamine, 100 U/mL penicillin and 100 µg/mL streptomycin. Cytotoxicity of compounds is expressed as IC_{50} values, the concentrations that cause 50% growth inhibition. The results were determined using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide [51]. Cells were dispensed into 96-well microtiter plates at a density of 10,000 cells/well. Following overnight incubation, cells were treated with the tested compounds, oligomycin A and rotenone in the range concentration 0.1–100 µM, and with the range 1–500 µM for H₂O₂. Then the plates were incubated at 37 °C for 24 h. An amount of 10 µL of 0.5% w/v MTT was further added to each well and the plates were incubated for an additional 3 h at 37 °C. Finally the cells were lysed by addition of 100 µL of

DMSO/EtOH 1:1 (v/v) solution. The absorbance at 570 nm was determined using a Perkin Elmer 2030 multilabel reader Victor TM X3.

Neuroprotective capacity

The neuroprotective capacity of the compounds was tested, as reported by Bencheekroun et al. [45] Briefly, the ability of the compounds to prevent the human neuroblastoma cell line SH-SY5Y from death was evaluated by using three toxicity models: 1) H_2O_2 , as a producer of exogenous free radicals, 2) oligomycin A, a mitochondrial respiratory chain blocker which produces mitochondrial ROS by inhibiting the mitochondrial electron-transport chain complex V, and 3) rotenone, showing the same effect of oligomycin A by inhibiting the mitochondrial electron-transport chain complex I. In this experiment, the toxic insults were used at the concentrations equal to their IC_{50} after 24 h of incubation: 195, 30 and 75 μM for H_2O_2 , oligomycin A and rotenone, respectively. The tested compounds were used at non-cytotoxic concentrations after 24h of incubation. Compounds that are able of inhibiting the effect of the toxic insults may be considered neuroprotectants [63]. For the assay, SH-SY5Y cells were plated in 96-well plates at a seeding density of 10,000 cells /well. After 24 h of incubation at 37 °C in a humidified incubator containing 5% CO_2 , cells were co-incubated with H_2O_2 (195 μM), or oligomycin A (30 μM), or rotenone (75 μM) and tested compound at several concentration for further 24 h. In particular, compound **15** was tested at concentrations 0.1 and 1 μM . The cell viability was determined by MTT assay and analysed as previously described.

Bi-directional Transport Studies on MDCKII-MDR1 Monolayers

Apical to basolateral (Papp, AP) and basolateral to apical (Papp, BL) permeability of the tested compounds was measured using Madin-Darby Canine Kidney (i.e., MDCK) cells, retrovirally transfected with the human MDR1 cDNA (MDCKII-MDR1), as previously reported [49,50]. The cells were cultured in DMEM medium and seeded at a density of 100,000 cell/cm² onto polyester 12 well Transwell inserts (pore size 0.4 µm, 12 mm diameter, apical volume 0.5 mL, basolateral volume 1.5 mL). At first, MDCKII-MDR1 cell barrier function was verified by means of trans-epithelial electrical resistance (TEER) using an EVOM apparatus and by the measurement of the flux of the paracellular standard fluorescein isothiocyanate-dextran (FD4, Sigma) (200 µg/mL) and the transcellular standard diazepam (75 µM). The TEER was measured in growth media at room temperature and calculated as the measured resistance minus the resistance of an empty Transwell (blank without cells). Cell monolayers with TEER values 800 Ohm cm² were used for the successive transport experiments. The cells were equilibrated in transport medium in both the apical and basolateral chambers for 30 minutes at 37 °C. The composition of transport medium was: 0.4 mM K₂HPO₄, 25 mM NaHCO₃, 3 mM KCl, 122 mM NaCl, 10 mM glucose with final pH of 7.4, and the osmolarity was 300 mOsm as determined by a freeze point based osmometer. At time 0, culture medium was aspirated from both the apical (AP) and basolateral (BL) chambers of each insert, and cell monolayers were washed three times (10 min per wash) with Dulbecco's Phosphate Buffered Saline (DPBS) pH = 7.4. Finally, a solution of compounds diluted in transport medium was added to the apical or basolateral chamber. For AP-to-BL or BL-to-AP flux studies, the drug solution was added in the AP chamber or in the BL chamber, respectively. Except for FD4, which was solubilized directly in the assay medium at a concentration of 200 µg/mL, the other compounds were first dissolved in DMSO and then diluted with the assay medium to a final concentration of 75 µM. Next, the

tested solutions were added to the donor side (0.5 mL for the AP chamber and 1.5 mL for the BL chamber) and fresh assay medium was placed in the receiver compartment. The percentage of DMSO never exceeded 1% (v/v) in the samples. The transport experiments were carried out under cell culture conditions (37 °C, 5% CO₂, 95% humidity). After incubation time of 120 min, samples were removed from the apical and basolateral side of the monolayer and then stored until further analysis. Quantitative analysis of the tested compounds and diazepam, was performed through UV–visible (Vis) spectroscopy using a PerkinElmer double-beam UV–visible spectrophotometer Lambda Bio 20 (Milan, Italy), equipped with 10 mm path-length-matched quartz cells. Standard calibration curves were prepared at maximum absorption wavelength of each compound using PBS as solvent and were linear ($r^2 = 0.999$) over the range of tested concentration (from 5 to 100 μ M). The FD4 samples were analyzed with a Victor3 fluorometer (Wallac Victor3, 1420 Multilabel Counter, Perkin-Elmer) at excitation and emission wavelengths of 485 and 535 nm, respectively. Each compound was tested in triplicate, and the experiments were repeated three times.

The apparent permeability, in units of cm/sec, was calculated using the following equation:

$$P_{app} = \left(\frac{V_A}{\text{area} \times \text{time}} \right) \times \left(\frac{[\text{drug}]_{\text{acceptor}}}{[\text{drug}]_{\text{initial}}} \right)$$

where “V_A” is the volume in the acceptor well, “area” is the surface area of the membrane, “time” is the total transport time, “[drug]_{acceptor}” is the concentration of the drug measured by UV-spectroscopy and “[drug]_{initial}” is the initial drug concentration in the AP or BL chamber. Efflux ratio (ER) was calculated using the following equation: $ER = P_{app, BL-AP} / P_{app, AP-BL}$, where $P_{app, BL-AP}$ is the apparent permeability of basal-to-apical transport, and $P_{app, AP-BL}$

BL is the apparent permeability of apical-to-basal transport. An efflux ratio greater than 2 indicates that a test compound is likely to be a substrate for P-gp transport.

Antinociceptive activity in *in-vivo* model

For the assessment of antinociceptive activity of inflammatory pain, mice were subjected to the formalin test. Male Swiss CB1 mice (Envigo, S.Pietro al Natisone (UD)) weighing 25-30 g were used. Animals were kept at a constant room temperature (25 ± 1 °C) under a 12:12 h light and dark cycle with free access to food and water. Each mouse was used for only one experiment. Experimental procedures were approved by the Local Ethical Committee (IACUC) and conducted in accordance with international guidelines as well as European Communities Council Directive and National Regulations (CEE Council 86/609 and DL 116/92). All tests were performed blind to treatment.

Formalin (5%, 10 μ l; Sigma-Aldrich) was injected subcutaneously into the plantar side of the right hind paw [64]. After the injection, mice were immediately placed in a plexiglas box: the total time (in seconds) spent on licking or biting the injected hind paw was recorded for each five minute in selected intervals, 0–10 (phase I) and 10–60 (phase II) min, in the different experimental group as indicator of nociceptive behavior. Formalin scores were separated in two phases: phase I (0–10 min) and phase II (10–60 min). A mean response was then calculated for each phase. Compound **15** and WAY-100635 (Sigma-Aldrich) were dissolved in normal saline solution containing 10% dimethyl sulfoxide (DMSO, Sigma-Aldrich). A vehicle solution containing 10 % DMSO was given as control. Compound **15** and vehicle were intraplantar (i.p.) administered (5ml/kg) 15 minutes before formalin. WAY-100635 (3mg/kg i.p.) was injected 30 minutes before test compound or vehicle. Data are expressed as mean values (SEM). Analysis of variance (two-way repeated measures ANOVA followed by post hoc Bonferroni test) were

performed to assess significance using the InStat 3.0 software (GraphPad Software, San Diego, CA). $p < 0.05$ was considered significant.

Molecular Modeling

Ligand preparation

Compounds **A** and **9-11**, **14-16**, **19-21** were built, parameterised (Gasteiger-Huckel method) and energy minimised within MOE using MMFF94 force field [65]. For all compounds, the protonated form was considered for the *in silico* analyses.

Alpha_{1D} homology modeling

The alpha_{1D} theoretical model has been built starting from the X-ray structure of the β_2 -adrenoreceptor (PDB code: 2RH1; resolution = 2.40 Å) [66], by applying the ligand-based homology modeling strategy. In particular, compound **A** was docked into the β_2 -adrenoreceptor binding site and employed in the alpha_{1D} model building and refinement, by taking into account the ligand specific steric and chemical features. The amino acid sequence of α_{1D} -adrenoreceptor (P25100) was retrieved from the SWISSPROT database [67] while the three-dimensional structure co-ordinates file of the GPCR template was obtained from the Protein Data Bank [68]. The amino acid sequences of α_{1D} TM helices were aligned with the corresponding residues of 2RH1, on the basis of the Blosom62 matrix (MOE software). The connecting loops were constructed by the loop search method implemented in MOE. The MOE output file included a series of ten models which were independently built on the basis of a Boltzmann-weighted randomized procedure [69], combined with specialized logic for the handling of sequence insertions and deletions [70]. Among the derived models, there were no significant main chain deviations. The model with the best packing quality function was selected for full energy

minimization. The retained structure was minimized with MOE using the AMBER94 force field [71]. The energy minimization was carried out by the 1000 steps of steepest descent followed by conjugate gradient minimization until the rms gradient of the potential energy was less than 0.1 kcal mol⁻¹ Å⁻¹. The assessment of the final obtained model was performed using Ramachandran plots, generated within MOE.

Docking studies

Docking studies were performed according to the following protocol. The binding site of the ligand in the 5HT_{1A} receptor model (previously built by us) was determined starting from the fact that, for the ligand activity, formation of the salt bridge between the protonated piperazine nitrogen on the ligand and Asp116 is necessary [72-74].

On the other hand, the α_{1d} -adrenoreceptor binding site has been determined taking into account the conserved residues highlighted by superimposition on the 2RH1 X-ray β 2-adrenoreceptor binding site. For all compounds, each isomer was docked into the putative ligand binding site by means of the Surflex docking module implemented in Sybyl-X1.0 [75].

Surflex-Dock uses an empirically derived scoring function based on the binding affinities of X-ray protein-ligand complexes. The Surflex-Dock scoring function is a weighted sum of non-linear functions involving van der Waals surface distances between the appropriate pairs of exposed protein and ligand atoms, including hydrophobic, polar, repulsive, entropic and solvation and crash terms represented in terms of a total score conferred to any calculated conformer.

Then, for all compounds, the best docking geometries (selected on the basis of the SurFlex scoring functions) were refined by ligand/receptor complex energy minimization (CHARMM27) by means of the MOE software. To verify the reliability of the derived docking poses, the

obtained ligand/receptor complexes were further investigated by docking calculations (10 run), using MOE-Dock (Genetic algorithm; applied on the poses already located into the putative 5-HT_{1A} and alpha_{1D} receptors). The conformers showing lower energy scoring functions and rmsd values (respect to the starting poses) were selected as the most stable and allowed us to identify the most probable conformers interacting with the two GPCRs.

Prediction of ADMET Properties

The prediction of ADMET properties were performed using the Advanced Chemistry Developmente (ACD) Percepta platform. The aforementioned descriptors blood-brain barrier permeation (LogBBB), rate of passive diffusion-permeability (Log PS), human intestinal absorption (HIA), volume of distribution (Vd), median lethal dose (LD₅₀) related to oral administration and the logarithmic ratio of the octanol-water partitioning coefficient (cLogP) were calculated. All of them were derived and evaluated by Percepta on the basis of training libraries, implemente in the software, which include a consistent number of molecules whose pharmacokinetic and toxicity profile are known.

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Highlights

- 1,4-dioxaspiro[4,5]dec-2-ylmethyl-amines were synthesized and tested.
- Oxygen/Sulphur substitution favors 5-HT_{1A}R affinity, potency and efficacy.
- **14** and **15** behave as selective and potent 5-HT_{1A}R partial agonists.
- **15** shows a promising neuroprotective activity *in-vitro*.
- **15** reduces significantly the linking time in Phase II at a dose of 10mg/Kg i.p.