

This is the peer reviewed version of the following article:

Bone marrow-specific loss of ABL1 induces myeloproliferative neoplasm with features resembling human myelofibrosis / Chorzalska, Anna; Morgan, John; Ahsan, Nagib; Treaba, Diana O.; Olszewski, Adam J.; Petersen, Max; Kingston, Nathan; Cheng, Yan; Lombardo, Kara; Schorl, Christoph; Yu, Xiaoqing; Zini, Roberta; Pacilli, Annalisa; Tepper, Alexander; Coburn, Jillian; Hryniewicz-Jankowska, Anita; Zhao, Ting C.; Oancea, Elena; Reagan, John L.; Liang, Olin; Kotula, Leszek; Quesenberry, Peter J.; Gruppuso, Philip A.; Manfredini, Rossella; Vannucchi, Alessandro Maria; Dubielecka, Patrycja M.. - In: BLOOD. - ISSN 0006-4971. - 132:19(2018), pp. 2053-2066. [10.1182/blood-2018-05-848408]

*Terms of use:*

The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

16/05/2026 19:38

(Article begins on next page)

16/05/2026 19:38

## **Bone marrow-specific loss of *ABI1* induces myeloproliferative neoplasm with features resembling human myelofibrosis**

Running title: Loss of Abi-1 induces myeloproliferative neoplasm

Anna Chorzalska<sup>1</sup>, John Morgan<sup>2</sup>, Nagib Ahsan<sup>3,4</sup>, Diana O. Treaba<sup>5</sup>, Adam J. Olszewski<sup>6</sup>, Max Petersen<sup>1</sup>, Nathan Kingston<sup>1</sup>, Yan Cheng<sup>6</sup>, Kara Lombardo<sup>5</sup>, Christoph Schorl<sup>7</sup>, Xiaoqing Yu<sup>8</sup>, Roberta Zini<sup>9</sup>, Annalisa Pacilli<sup>10</sup>, Alexander Tepper<sup>1</sup>, Jillian Coburn<sup>3</sup>, Anita Hryniewicz-Jankowska<sup>11,14</sup>, Ting C. Zhao<sup>12</sup>, Elena Oancea<sup>13</sup>, John L. Reagan<sup>6</sup>, Olin Liang<sup>6</sup>, Leszek Kotula<sup>14</sup>, Peter J. Quesenberry<sup>6</sup>, Philip A. Gruppuso<sup>15</sup>, Rossella Manfredini<sup>9</sup>, Alessandro Maria Vannucchi<sup>10</sup> and Patrycja M. Dubielecka<sup>1\*</sup>

<sup>1</sup> Signal Transduction Lab, Division of Hematology/Oncology at Rhode Island Hospital and Warren Alpert Medical School at Brown University, Providence, RI 02903, USA

<sup>2</sup> Flow Cytometry and Cell Sorting Core Facility, Roger Williams Medical Center, Providence, RI 02906, USA

<sup>3</sup> COBRE Center for Cancer Research Development, Proteomics Core Facility. Rhode Island Hospital, Providence, RI 02903, USA

<sup>4</sup> Division of Biology and Medicine, Brown University, Providence, RI 02903, USA

<sup>5</sup> Department of Pathology and Laboratory Medicine at Rhode Island Hospital and Warren Alpert Medical School at Brown University, Providence, RI 02903, USA

<sup>6</sup> Division of Hematology/Oncology at Rhode Island Hospital and Warren Alpert Medical School at Brown University, Providence, RI 02903, USA

<sup>7</sup> Genomics Core Facility, Brown University, Providence, RI 02903, USA

<sup>8</sup> Department of Biostatistics, Yale School of Public Health, New Haven, CT 06510, USA

<sup>9</sup> Center for Regenerative Medicine “Stefano Ferrari”, Department of Life Sciences, University of Modena and Reggio Emilia, Modena, Italy

<sup>10</sup> CRIMM, Center for Research and Innovation for Myeloproliferative Neoplasms, AOU Careggi, and Department of Experimental and Clinical Medicine, University of Florence, Excellence Center Denothe, Florence, Italy

<sup>11</sup> Cytobiochemistry Lab, Department of Biotechnology, University of Wroclaw, 50-383 Wroclaw, Poland

<sup>12</sup> Cardiovascular Lab, Department of Surgery, Roger Williams Medical Center, Boston University School of Medicine, Providence, RI 02906, USA

<sup>13</sup> Department of Molecular Pharmacology, Physiology, and Biotechnology, Brown University, Providence, RI 02912, USA

<sup>14</sup> Department of Urology, SUNY Upstate Medical University, Syracuse, New York 13210

<sup>15</sup> Department of Pediatrics, Brown University and Rhode Island Hospital, Providence, RI 02903

\*Corresponding author:

Patrycja M Dubielecka, Warren Alpert Medical School of Brown University, Signal Transduction Lab, Rhode Island Hospital, 1 Hoppin St, Coro West, Suite 5.01, Providence, RI 02903, USA, patrycja\_dubielecka-szczerba@brown.edu (P) 401 444-8391, (F) 401 444-4661.

## KEY POINTS

- Bone marrow-specific deletion of *Abi1* in mice results in MPN-like phenotype and is linked to hyperactive SFKs/STAT3/NF- $\kappa$ B signaling
- *ABI1* is downregulated in hematopoietic stem/progenitor cells and granulocytes from patients with PMF

## ABSTRACT

Although the pathogenesis of primary myelofibrosis (PMF) and other myeloproliferative neoplasms (MPNs) is linked to constitutive activation of the JAK-STAT pathway, JAK inhibitors neither have curative nor MPN-stem cell-eradicating potential, indicating that other targetable mechanisms are contributing to pathophysiology of MPNs. We previously demonstrated that Abelson interactor 1 (*Abi-1*), a negative regulator of Abelson kinase 1, functions as a tumor suppressor. Here we present data showing that bone marrow-specific deletion of *Abi1* in a novel mouse model leads to development of an MPN-like phenotype resembling human PMF. *Abi1* loss resulted in a significant increase in the activity of the Src Family Kinases (SFKs), STAT3 and NF- $\kappa$ B signaling. We also observed impairment of hematopoietic stem cells self-renewal and fitness, as evidenced in non-competitive and competitive bone marrow transplant experiments. CD34+ hematopoietic progenitors and granulocytes from patients with PMF showed decreased levels of *ABI1* transcript as well as increased activity of SFKs, STAT3 and NF- $\kappa$ B. In aggregate, our data link the loss of *Abi-1* function to hyperactive SFKs/STAT3/NF- $\kappa$ B signaling and suggest that this signaling axis may represent a regulatory module involved in the molecular pathophysiology of PMF.

## INTRODUCTION

The phenotype of primary myelofibrosis (PMF) is characterized by progressive bone marrow fibrosis, organomegaly, extramedullary hematopoiesis, thromboembolism, and, ultimately, marrow failure or transformation to acute myeloid leukemia (AML).<sup>1-3</sup> Median survival in PMF varies between 1 and 15 years depending on risk factors, and treatment options are limited.<sup>3,4</sup> Identification of *JAK2*-activating mutations as major drivers in myeloproliferative neoplasms (MPNs) prompted clinical development of JAK2 inhibitors.<sup>5,6</sup> Ruxolitinib, an ATP-mimetic JAK1/2 inhibitor, induces symptomatic improvement in PMF, but exacerbates associated cytopenias, does not have curative potential, and responses occur regardless of presence of *JAK2* mutations.<sup>7-11</sup> Therefore, a major need remains to identify other targetable mechanisms contributing to the pathogenesis of PMF and related MPNs, polycythemia vera (PV) and essential thrombocythemia (ET).

Abelson Interactor 1 (Abi-1) is a negative regulator of Abl1 kinase,<sup>12-15</sup> involved in regulation of cell proliferation.<sup>16,17</sup> By forming a complex with Wiskott-Aldrich syndrome protein family member 2 (WAVE2),<sup>18,19</sup> Wiskott–Aldrich Syndrome protein (WASP), or Diaphanous (Dia) formin<sup>16,18-23</sup> Abi-1 affects actin remodeling, cell adhesion and migration. Abi-1 also interacts with integrin  $\alpha 4$  and is involved in integrin  $\beta 1$  signaling.<sup>24-26</sup> Abi-1-deficient mice uniformly die *in utero* with lethal defects of the heart and placenta.<sup>19,24</sup> The role of Abi-1 in carcinogenesis is controversial, as both loss or overexpression were implicated in cancer.<sup>27-30</sup> Its involvement in malignant hematopoiesis, while reported by us and others, remains unclear.<sup>31-35</sup>

Here, we present evidence for direct involvement of Abi-1 in homeostasis of hematopoietic system. We found that conditional deletion of *Abi1* in murine bone marrow results in impairment of hematopoietic stem cell self-renewal, progressive anemia, megakaryocytosis and myeloid hyperplasia, with resulting PMF-like phenotype characterized by marrow fibrosis and splenomegaly. Furthermore, Abi-1 protein and mRNA levels are decreased in hematopoietic progenitors from patients with PMF, but not from those with ET or PV. Mechanistically, loss of Abi-1 leads to upregulation of SFKs, STAT3, and

NF- $\kappa$ B signaling, suggesting that the Abi-1/SFKs/STAT3/NF- $\kappa$ B axis may represent a new regulatory module involved in the pathophysiology of MPNs.

## **PATIENTS, MATERIALS AND METHODS**

### **Patient samples**

CD34<sup>+</sup> cells were isolated from bone marrow of PMF patients or from healthy marrow purchased from AllCells (Alameda, CA). Granulocytes were isolated from peripheral blood (PB) of patients with EV, PV, primary or secondary (PV- or ET-derived) myelofibrosis, and healthy donors (Supplementary Table 1). CD34 MicroBeads (Miltenyi Biotec, San Diego, CA) and gradient centrifugation were used for CD34<sup>+</sup> and granulocyte isolation, respectively. Human subject participation was conducted with informed consent, and approved by local ethics committees.

### **Transgenic mice**

Conditional *Abi1*(fl/fl) mice<sup>19</sup> were crossed with B6.Cg-Tg(*Mx1-cre*+1)Cgn/J strain (#003556, JAX, Bar Harbor, ME)<sup>36</sup> to generate *Abi1*(fl/fl);Tg(*Mx1-cre*(+/-)) mice. These animals were back-crossed to B6.SJL-*Ptprc*<sup>A</sup>*Pepec*<sup>B</sup>/BoyJ(#002014, JAX)(CD45.1) background. *Abi1*(fl/fl);Tg(*Mx1-cre*(-)), *Abi1*(fl/wt);Tg(*Mx1-cre*(+)), or *Abi1*(fl/fl);Tg(*Mx1-cre*(+)) mice were subjected to polyinosinic:polycytidylic acid [poly(I:C)](Invivo Gen, San Diego, CA)-induced activation of the Cre recombinase under control of *Mx1* promoter to obtain animals with an *Abi1*(fl/fl);Tg(*Mx1-cre*(-)), *Abi1*(-/wt);Tg(*Mx1-cre*(-)), or *Abi1*(-/-);Tg(*Mx1-cre*(+)) genotypes, designated as Abi-1<sup>WT</sup>, Abi-1<sup>HET</sup>, or Abi-1<sup>KO</sup>, respectively. Recombination of *Abi1*<sup>fl<sup>oxed</sup></sup> allele was confirmed by PCR. We evaluated 76 Abi-1<sup>WT</sup>, 41 Abi-1<sup>HET</sup>, and 85 Abi-1<sup>KO</sup> animals. Animal experiments were approved by the Institutional Animal Care and Use Committee.

### **Murine hematopoietic stem/progenitor cells isolation**

A biotin-conjugated antibody cocktail containing anti-TER119, CD127, CD8a, Ly-6G, CD11b, CD4, and CD45R was used to stain lineage-committed cells. BUV395-Streptavidin, anti-CD34-FITC, CD117/c-Kit-APC, Ly-6A/E/Sca-1-PE-Cy7(or -BV605), CD135/Flt3-PE, and CD16/CD32-PE were used to stain hematopoietic stem/progenitors. Cells were sorted using Legacy MoFlo High-Speed cell sorter.

### **Bone marrow transplantation**

For non-competitive bone marrow transplantation,  $5 \times 10^6$  marrow cells isolated by flushing from poly(I:C)-uninduced *Abi1*(fl/fl);Tg(*Mx1-cre*(+)) or *Abi1*(fl/fl);Tg(*Mx1-cre*(-)) (CD45.1) mice, were injected via tail vein into lethally irradiated (2x475cGy) C57BL/6 wild-type mice recipients (CD45.2, (#000664, JAX)). Four weeks post-transplant, *Abi1* inactivation was performed by poly(I:C)-induction. Donor chimerism was evaluated in PB every 4 weeks for 24 weeks. After 24 weeks, marrow was harvested from primary recipients and  $5 \times 10^6$  cells were transplanted into CD45.2 secondary recipients conditioned as above. Donor chimerism in PB was evaluated as above. For competitive repopulation assays, marrow cells were isolated via flushing from poly(I:C)-induced *Abi-1*<sup>KO</sup> or *Abi-1*<sup>WT</sup> (CD45.1) mice. After confirming *Abi1* inactivation, donor cells ( $1 \times 10^6$  *Abi-1*<sup>KO</sup> or *Abi-1*<sup>WT</sup>) were mixed with competitor cells ( $1 \times 10^6$ ) (1:1; CD45.1:CD45.2) and injected via tail vein into lethally irradiated CD45.2 recipients. Donor chimerism in PB was evaluated as above.

### **Cytokine levels assay**

Levels of IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, IL-10, IL-12p70, IL-17A, IL-23, IL-27, MCP-1, IFN- $\beta$ , IFN- $\gamma$ , TNF- $\alpha$ , and GM-CSF were detected in plasma of *Abi-1*<sup>WT</sup>(n=11) or *Abi-1*<sup>KO</sup>(n=11) 14-week-old animals using LEGENDplex™ Mouse Inflammation Panel (Biolegend, San Diego, CA).

### ***ABI1* silencing in human CD34+ cells**

Healthy CD34+ cells (n=3) were expanded for 48h in the presence of TPO, Flt3, SCF, IL-3, IL-6, and GM-CSF (StemCell Technologies, Cambridge, MA) and incubated for additional 48h with 15  $\mu$ M of *ABI1*-silencing antisense 2'-deoxy-2'-fluoro-beta-D-arabinonucleotides (FANA-*ABI1*-ASO) (AUM BioTech, Philadelphia, PA). After 48h culture with FANA-*ABI1*-ASO, *Abi-1* protein levels were evaluated by immunoblotting, and cell cycle status determined by EdU incorporation assay.

### **Liquid Chromatography–Tandem Mass Spectrometry**

Bone marrow was isolated from 20-weeks-old *Abi-1*<sup>WT</sup>(n=3) or *Abi-1*<sup>KO</sup>(n=3) animals by flushing. Cell pellets were lysed and 100 $\mu$ g of protein per sample was subjected to tryptic digestion. Tryptic peptides were subjected to LC-MS/MS using automated proteomic technology platform.<sup>37,38</sup>

## Gene microarrays

Gene array analysis of LSK-enriched cells from 14-week-old *Abi-1*<sup>KO</sup> (n=4) or *Abi-1*<sup>WT</sup> (n=5) mice was done using the Affymetrix-WT Pico Expression Kit (Affymetrix, Louisville, KY) and Affymetrix-3000 7G gene scanner. Partek Genomics Suite v6.6 was used for quality control and data analysis.

Microarray data are deposited in GEO as GSE83288.

## ***Abi-1*<sup>HET</sup> bone marrow transplant model of *MPL*<sup>W515L</sup>-mediated MPN**

MSCV-*MPL*<sup>WT</sup>-IRES-GFP and MSCV-*MPL*<sup>W515L</sup>-IRES-GFP retroviral vectors were generously provided by Ross Levine (MSKCC). Marrow was isolated from 14-week-old *Abi-1*<sup>WT</sup> or *Abi-1*<sup>HET</sup> animals, and *Abi1* recombination was verified by PCR. Retroviral transduction and murine marrow transplant assay were performed as previously described.<sup>39,40</sup> *Abi-1*<sup>WT</sup>/*MPL*<sup>WT</sup>, *Abi-1*<sup>HET</sup>/*MPL*<sup>WT</sup>, *Abi-1*<sup>WT</sup>/*MPL*<sup>W515L</sup>, and *Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup> transplantation groups were used.

## Statistics

Two-tailed unpaired t-tests or log-rank tests were used for between-group comparisons, using Bonferroni correction where appropriate. P-values <0.05 were considered statistically significant. Throughout, \* indicates p<0.05, \*\* indicates p<0.01, and \*\*\* indicates p<0.001. Two-tailed unpaired t-test and q-values for multiple hypothesis tests, using the R package QVALUE<sup>41</sup> were used to select peptides with significant change in paired analyses.

Additional materials and methods are described in Supplementary Information.

## RESULTS

### Downregulation of *ABI1* transcripts in human PMF samples

*ABI1* transcript levels in CD34+ cells isolated from bone marrow were decreased by approximately 40% in PMF (n=5) compared to controls (n=5, Figure 1A). Gene expression profiles (GEO/GSE53482)<sup>42</sup> of CD34+ cells isolated from PB also showed significant downregulation of *ABI1* in PMF (n=42) with mutations in *JAK2* or *CALR* relative to controls (n=31, Supplementary Figure 1A-C). Furthermore, granulocytes from patients with PMF (n=36), or secondary myelofibrosis post-PV (n=9) showed a 40-60% decrease in *ABI1* mRNA relative to controls (n=16; Figure 1B). Downregulation of *ABI1* in PMF granulocytes was observed regardless of *JAK2* or *CALR* mutation status (Figure 1C). Notably, no significant changes in granulocyte *ABI1* transcript levels were noted in ET (n=15), PV (n=20) or post-ET myelofibrosis (n=4, Figure 1B). Consistent with these findings, examination of gene expression profiles of CD34+ cells from bone marrow of ET (n=24) and PV (n=26) patients (GEO/GSE103176)<sup>43</sup> showed no downregulation of *ABI1* (Supplementary Figure 1D), indicating that reduced *ABI1* expression may be specific to PMF.

### Inactivation of *Abi1* in bone marrow induces leukocytosis, thrombocytosis, anemia, splenomegaly and decreased survival

To determine the role of *Abi-1* in the homeostasis of the hematopoietic system, we phenotyped transgenic *Abi-1*<sup>KO</sup> mice carrying a marrow-selective knockout of *Abi1*. We first confirmed both inducible inactivation of the *Abi1*<sup>fl<sup>ox</sup></sup> allele and loss of *Abi-1* protein in the marrow (Figure 2A-C, Supplementary Figure 2A-B). *Abi-1* levels were also reduced in spleen and liver, but not in heart, lung, or kidney (Supplementary Figure 2C). *Abi-1*<sup>KO</sup> mice showed femur pallor and splenomegaly that progressed with age, but no change in liver size (Figure 2D-E). Blood count analysis at 14, 35, and 56 weeks indicated progressive leukocytosis, mild anemia and thrombocytosis in the *Abi-1*<sup>KO</sup>, which were absent in *Abi-1*<sup>WT</sup> animals (Figure 2F and Supplementary Table 2). Survival of *Abi-1*<sup>KO</sup> mice decreased 20 weeks post-poly(I:C) injection, with significant weight loss after 60 weeks, and death in all animals by 66 weeks of age (Figure 2G, Supplementary Figure 2D) likely due to progressive disease. We observed no changes

in gross pathology, blood counts or survival in 56-week-old *Abi-1*<sup>HET</sup> animals. Therefore, in subsequent analyses we focused on characterization of *Abi-1*<sup>KO</sup> mice. Our findings suggest that marrow-specific inactivation of *Abi1* disrupts normal hematopoiesis.

### ***Abi-1* loss results in myeloid hyperplasia, megakaryocytosis and fibrosis**

As *Abi-1*<sup>KO</sup> animals aged, the fraction of myeloid (CD11b+/Gr-1+) cells in their PB increased at the expense of lymphoid B220 cells, while the CD3+ lymphoid fraction remained unchanged (Figure 3A). *Abi-1*<sup>KO</sup> erythrocytes demonstrated polychromasia, anisopoikilocytosis, and teardrop-shaped forms (Figure 3B, Supplementary Figure 3A). Giant platelets were observed in 2 of 10 14-week-old, 4 of 10 36-week-old, and 5 of 10 56-week-old or older *Abi-1*<sup>KO</sup> mice. Bone marrow showed hypercellularity with myeloid hyperplasia, erythroid hypoplasia, and megakaryocytosis (Figure 3C-E, Supplementary Figure 3B-C). Bone marrow cellularity at the distal aspect of femur in the medial and lateral condyles was 95% for *Abi-1*<sup>WT</sup> and 100% for 56-week-old *Abi-1*<sup>KO</sup> mice (Supplementary Figure 3D). Spleen histology showed expansion of red pulp, with numerous islands of megakaryocytic, myeloid, and erythroid infiltration (Figure 3C-D, Supplementary Figure 3B-C). Clusters of megakaryocytes were commonly observed in marrow and spleen of 56-week-old *Abi-1*<sup>KO</sup> mice (Supplementary Figure 3C). *Abi-1*<sup>KO</sup> marrow samples and spleens showed progressive deficits of stainable iron on Prussian blue staining (Figure 3D and 3F, Supplementary Figure 4A-B), possibly linked to observed anemia (Supplementary Table 2, Figure 2F). Silver stain and semiquantitative grading<sup>44</sup> showed progressive increase in reticulin fibrosis in *Abi-1*<sup>KO</sup> marrow, from grade 1 (out of 3) at 14 weeks to grade 2-3 at 56 weeks (Figure 3G, Supplementary Figure 4C). Increased thickness and density of reticulin fibers was present in *Abi-1*<sup>KO</sup> spleen, from grade 1 at 14 weeks to grade 2 at 56 weeks (Figure 3G, Supplementary Figure 4C). Progressive splenomegaly, megakaryocytosis, and marrow and spleen fibrosis in *Abi-1*<sup>KO</sup> animals meet Mouse Models of Human Cancers Consortium criteria for MPN.<sup>45</sup>

### ***Abi-1* deficiency results in increased frequency and cell cycle activity of hematopoietic stem/progenitor cells**

We assessed the frequency of hematopoietic stem and progenitor cells in the marrow of 14-week-old *Abi-1*<sup>WT</sup> or *Abi-1*<sup>KO</sup> mice (Supplementary Figure 5A). Deletion of *Abi1* resulted in no change in Lin- fractions. However, there were increased frequencies of LSKs (from 2.3% $\pm$ 0.4] to 4.6% $\pm$ 0.8]) and LK progenitors (from 4.9% $\pm$ 0.9] to 17.6% $\pm$ 1.1], Figure 4A-B), a 2.6-fold increase in LT-HSCs, no change in ST-HSCs, a 47% reduction in CMPs, and a 25% increase in GMPs (Figure 4C, Supplementary Figure 5B). Evaluation of the absolute number of stem/progenitor cells confirmed these trends with exception of CMPs, which showed an increased number in *Abi-1*<sup>KO</sup> mice (Supplementary Figure 5C).

Given this evidence for expansion of HSCs, we evaluated the cell cycle activity of LSK and LT-HSC cells. *Abi-1*<sup>KO</sup> mice at 14 weeks showed a higher proportion of LSK cells in S-phase, and 46% more LT-HSCs in the S/G2/M phases relative to *Abi-1*<sup>WT</sup> animals (Figure 4D-E, Supplementary Figure 5D-E). Transcript expression analysis of genes regulating cell cycle progression in LT-HSCs confirmed these observations (Supplementary Figure 5F).

To determine the effect of *Abi-1* loss on hematopoietic progenitors, we performed colony-forming unit (CFU) assays. Results showed a significant decrease in BFU-E and increase in CFU-GM colonies derived from *Abi-1*<sup>KO</sup> marrow isolated from 14-weeks-old mice, consistent with anemia and leukocytosis (Supplementary Figure 5G). We further observed a 3-fold increase in CFU-Mk colonies, consistent with the observed megakaryocytosis in *Abi-1*<sup>KO</sup> mice (Supplementary Figure 5H). We found no indication of growth factor independence or hypersensitivity.

To examine the effect of *Abi-1* loss on long-term engraftment and self-renewal of HSCs, we transplanted bone marrow cells isolated from *Abi-1*<sup>WT</sup> or *Abi-1*<sup>KO</sup> mice (before poly(I:C) injection), into lethally irradiated recipient C57BL/6 wild-type mice in the absence of competitor cells. Four weeks after the transplantation, *Abi-1* loss was induced by poly(I:C)-induction. Whereas initial engraftment remained comparable (~95%) for primary recipients of *Abi-1*<sup>WT</sup> or *Abi-1*<sup>KO</sup> transplants, 12 weeks-post poly(I:C)-induction *Abi-1*<sup>KO</sup>-transplanted mice showed progressive loss of chimerism (Figure 4F). These mice also exhibited significant weight loss at 24 weeks post-transplant, with an average weight of

22.2g( $\pm$ 1.8) compared with 28.6g( $\pm$ 3.1) for *Abi-1*<sup>WT</sup> marrow recipients (p=0.04). Bone marrow cells were harvested from primary recipients 24 weeks post-transplant and transplanted into secondary recipients. A further decrease in donor chimerism and significantly reduced initial engraftment of *Abi-1*<sup>KO</sup> bone marrow cells were seen in secondary recipients (Figure 4G).

In competitive repopulation assays, C57BL/6 wild-type mice transplanted with the whole bone marrow isolated from *Abi-1*<sup>KO</sup> mice 4-weeks post-poly(I:C)-induction (after PCR confirmed recombination) showed progressive decrease in donor chimerism relative to *Abi-1*<sup>WT</sup> marrow recipients, reaching significance 4 months after the transplant (Supplementary Figure 5I). Of note, initial engraftment at 4 weeks post-transplant was comparable between recipients of *Abi-1*<sup>KO</sup> or *Abi-1*<sup>WT</sup> marrow (46% and 45% chimerism, respectively).

To confirm a direct link between *Abi-1* loss and cell cycle activity in hematopoietic stem/progenitor cells, we performed EdU incorporation assays using CD34+ cells isolated from the bone marrow of healthy donors (n=3) and exposed to *ABI1*-silencing FANA antisense oligonucleotides (FANA-*ABI1*-ASO). More than 50% efficiency of *ABI1* silencing after 48h incubation with FANA-*ABI1*-ASO was noted for CD34+ cells (Figure 4H, Supplementary Figure 6A). Cellular penetrance of FANA oligos was confirmed by confocal microscopy and FACS (Supplementary Figure 6B-C). EdU incorporation assays showed that 48h exposure to FANA-*ABI1*-ASO resulted in a nearly 2-fold increase in CD34+ cells in S-phase (Figure 4I and Supplementary Figure 6D). Overall, our data suggest that deletion of *Abi-1* leads to abnormal cell cycle activity, impaired HSCs self-renewal and defective reconstitution of hematopoiesis.

### **Activation of NF- $\kappa$ B pathway and pro-inflammatory signaling in *Abi-1*<sup>KO</sup> mice**

To determine factors contributing to the abnormal expansion of hematopoietic stem/progenitor cells in *Abi-1*-deficient animals, we performed a genome-wide expression analysis of LSK cells from 14-week-old *Abi-1*<sup>KO</sup> and *Abi-1*<sup>WT</sup> mice. We found significant overexpression of genes regulated by or involved in regulation of the NF- $\kappa$ B pathway (Figure 4J and Supplementary Table 3). MetaCore Pathway Analysis indicated that the NF- $\kappa$ B pathway was one of the top 10 dysregulated pathways in

the Abi-1<sup>KO</sup> LSK cells (Supplementary Figure 6E). Ingenuity Pathway Analysis identified three overlapping networks in which NF-κB activation was predicted based on increased expression of known NF-κB targets (*Cd69, Cxcl10, Cxcl2, Hp*, and *Nfkbia*) and regulators (*Myd88, Chuk*, and *Ikbkb*, Supplementary Figure 6F).<sup>46,47</sup> Of note, gene expression profiling of CD34+ cells from PMF patients (GEO/GSE53482) showed upregulation of the same NF-κB target genes *CXCL10, CD69*, and *HP* (Supplementary Figure 7A), as well as genes involved in leukocyte migration and recruitment (*FCER1A, FCER1G, FCGR2A, TPSAB1* and *TPSB2*) (Supplementary Figure 7B).<sup>42</sup>

We also assessed levels of 13 pro-inflammatory cytokines, finding a nearly 2-fold increase in IL-1B, IL-12, IL-17, IL-23, IL-27, and MCP-1, a roughly 10-fold increase in INFγ, and no significant changes in IL-1A, IL-6, IL-10, TNFα or IFNβ in Abi-1<sup>KO</sup> plasma (Figure 4K). Overall, these data link upregulation of NF-κB pathway and inflammatory signaling to Abi-1 deficiency in hematopoietic system.

#### **Abi-1 deficient bone marrow cells exhibit increased activity of SFKs, STAT3 and NF-κB.**

Based on our gene expression data indicating active NF-κB signaling in Abi-1<sup>KO</sup> marrow, we evaluated major components of this pathway by immunoblotting. We found increased levels of IκB accompanied by increased phosphorylation of NF-κB (Figure 5A, Supplementary Figure 8A). Surprisingly, in the Abi-1<sup>KO</sup> bone marrow showed only modest increase in JAK2 and STAT5 activities, while the phosphorylation of STAT3, Akt and Erk1/2 increased significantly (Figure 5B, Supplementary Figure 8B).

Based on our previous work showing high-affinity binding between Abi-1 and the Src homology domains 2 (SH2) of Src Family Kinases (SFKs), we assessed the activity of SFKs.<sup>48,49</sup> We found increased phosphorylation of the Src auto-inhibitory Tyr-527 site. However, pan-phospho-Src family antibody showed increased phosphorylation on Tyr-416, consistent with activation of other SFKs (Figure 5C, Supplementary Figure 8C). We did not detect phosphorylation of c-Abl on Tyr-412 in Abi-1<sup>KO</sup> bone marrow cells (Supplementary Figure 8D). Immunoblotting showed an Abi-1-loss-associated decrease in the stability of WAVE2 complex components (WAVE2, Nap1, and Sra-1) (Figure 5D, Supplementary Figure 9A), consistent with our previous findings.<sup>19</sup>

To obtain unbiased insight into the mechanistic effects of Abi-1 loss, we performed label-free, intensity-based quantitative proteomic analysis of bone marrow from 20-week-old Abi-1<sup>KO</sup> and Abi-1<sup>WT</sup> mice. The analysis yielded 12,103 peptides derived from 2,464 unique proteins. When quantified, there were significant changes in the abundance of 318 peptides representing 226 unique proteins in the Abi-1<sup>KO</sup> samples (Supplementary Table 4, Supplementary Figure 9B). A volcano plot showed that among peptides with a more than two-fold difference, 25.1% showed lower and 74.9% higher abundance (Figure 5E). Among peptides showing >2-fold increase in abundance in Abi-1<sup>KO</sup> samples, we detected peptides derived from calreticulin, CD177, CD97, haptoglobin, Mac-1, myeloperoxidase, STAT1, STAT3, and SFKs Hck and Fgr. We interpreted these data as confirming not only activation of SFKs and STAT3 signaling, but also an increase in proteins previously associated with MPN. Within the group of peptides showing >2-fold decrease in abundance, we detected peptides derived from WAVE2, Sra-1, and Nap1 (Figure 5F).

Finally, immunoblotting analysis performed on CD34+ or CD34- cells isolated from PMF patients showed decreases of 80% and 50% in Abi-1 protein levels, respectively. Similar to Abi-1-depleted murine marrow, we also found significant upregulation of SFKs and NF- $\kappa$ B activities (Figure 5G, Supplementary Figure 9C-D). These data indicate that SFKs/STAT3/NF- $\kappa$ B signaling operating in Abi-1-deficient marrow may be involved in pathogenesis of the observed PMF-like phenotype. A model of Abi-1/SFKs/STAT3/NF- $\kappa$ B crosstalk is presented in Figure 5H.

### **Inactivation of one copy of Abi-1 accelerates development of MPL<sup>W515L</sup>-mediated MPN-like disease**

To further explore the role of Abi-1 in the pathogenesis of MPNs, we assessed involvement of Abi-1 in one of the established models of the disease. Such models are based on expression of mutated *JAK2*, *CALR*, or *MPL*. Because of less pronounced *ABI1* loss in granulocytes bearing *CALR* mutation, and no significant overactivity of JAK2-STAT5 in Abi-1<sup>KO</sup> marrow, we decided to use the transplant model of MPL<sup>W515L</sup>-mediated MPN.<sup>39</sup> As lethality and penetrance of the MPL<sup>W515L</sup> model is variable in C57/BL6 wild-type mice recipients (in contrast to full penetrance in tumor growth-permissive

Balb/C background),<sup>39,50</sup> and homozygous loss of *Abi1* allele is sufficient to produce MPN phenotype, we decided to use *Abi-1*<sup>HET</sup> animals to achieve controllable manifestation of the disease in double-mutant (*Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup>) animals.

Bone marrow isolated from *Abi-1*<sup>WT</sup> (n=3) and *Abi-1*<sup>HET</sup> (n=3) animals was transduced by retrovirus encoding *MPL*<sup>WT</sup> or *MPL*<sup>W515L</sup>. Transduction efficiency was >60%. *Abi-1*<sup>WT</sup> marrow cells expressing *MPL*<sup>WT</sup> or *MPL*<sup>W515L</sup> (*Abi-1*<sup>WT</sup>/*MPL*<sup>WT</sup> or *Abi-1*<sup>WT</sup>/*MPL*<sup>W515L</sup>), or *Abi-1*<sup>HET</sup> marrow cells expressing *MPL*<sup>WT</sup> or *MPL*<sup>W515L</sup> (*Abi-1*<sup>HET</sup>/*MPL*<sup>WT</sup> or *Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup>) were transplanted into lethally irradiated C57/BL6 mice (n=6 per group) (Figure 6A). Analyses performed 60 days post-transplant showed no femur pallor or significant changes in spleen or liver sizes in the *Abi-1*<sup>WT</sup>/*MPL*<sup>WT</sup>, *Abi-1*<sup>WT</sup>/*MPL*<sup>W515L</sup> or *Abi-1*<sup>HET</sup>/*MPL*<sup>WT</sup> groups. Conversely, *Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup> animals showed 3-fold and 1.7-fold increases in spleen and liver size, respectively, and femur pallor (Figure 6B-C). Furthermore, we noted pronounced leukocytosis, thrombocytosis, and polycythemia in *Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup> animals compared with *Abi-1*<sup>WT</sup>/*MPL*<sup>W515L</sup> (Figure 6D). We also observed a significant increase in Mac-1+/Gr-1+, CD41, TER119+/CD71+, TER119+/CD71- and B220+ cells in both marrow and blood, and a decrease in CD3+ cells in the marrow of the *Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup> animals (Figure 6E and Supplementary Figure 10A-B). Staining for reticulin fibers in the *Abi-1*<sup>HET</sup>/*MPL*<sup>W515L</sup> mice showed significant (3+) thickening in both marrow and spleen (Figure 6F). Overall, these data suggest that loss of one copy of *Abi1* markedly accelerates the *MPL*<sup>W515L</sup>-mediated MPN in mice.

## DISCUSSION

Extensive evidence implicates dysregulated JAK/STAT signaling in the pathophysiology of MPNs. Nonetheless, JAK2 inhibitors do not have salutary disease-modifying effects at the stem cell level.<sup>7-10</sup> The cross-talk between JAK/STAT, MAPK, and Akt pathways, and the potential of therapeutic targeting of these pathways has received much less attention.<sup>39,51-53</sup> Inflammation signaling and cytokine involvement in MPNs, particularly PMF, is widely acknowledged; however, the specific role of NF- $\kappa$ B signaling in MPNs was highlighted only recently.<sup>54</sup>

Abi-1 is an adapter protein involved in regulation of the Abl1, PI3K and Ras signaling, with a previously unknown role in hematopoiesis.<sup>12,17,49</sup> To address this gap, we generated bone marrow-specific murine knockout of *Abi1*. Loss of Abi-1 proved sufficient to induce MPN-like phenotype that replicates many features of human myelofibrosis. Myeloid hyperplasia, megakaryocytosis, progressive fibrosis, splenomegaly, and extramedullary hematopoiesis observed in Abi-1<sup>KO</sup> mice resemble other murine models of PMF.<sup>6,39,55-58</sup> Notably, we observed no transformation to acute leukemia, yet Abi-1<sup>KO</sup> animals had shorter lifespan and evidence of cytokine dysregulation. While there was no significant phenotypic effect of heterozygous Abi-1 loss, given that PMF typically develops in patients older than 55 (equivalent to 18 months in mice), we continue observation of the effect of heterozygous *Abi1* loss in mice beyond 72 weeks of age.

Molecular analyses of the MPN-like phenotype in Abi-1<sup>KO</sup> mice showed unexpectedly only modest activation of JAK2-STAT5 signaling, with hyperactivation of SFKs, STAT3 and NF- $\kappa$ B. Upregulation of Mpo, integrin  $\alpha$ M $\beta$ 2, Runx1, Hck, Fgr, STAT3 and NF- $\kappa$ B target- haptoglobin, seen in proteomic analysis, are consistent with increase in myeloid cells and upregulated SFKs, STAT3 and NF- $\kappa$ B signaling in Abi-1<sup>KO</sup> mice. Further work will be needed to elucidate how overactive SFKs/STAT3/NF- $\kappa$ B signaling relates to established MPN models. *JAK2*, *MPL*, or *CALR* mutations are frequent events in MPNs,<sup>5,6,39,59</sup> and murine models incorporate these variants.<sup>39,51,56,60-63</sup> In contrast to *JAK2*<sup>V617F</sup> models that progress to secondary myelofibrosis via PV-like stage,<sup>51,60,61,64</sup> our model resembles the TPO-driven models expressing MPL<sup>W515L</sup> or CALR<sup>del52</sup>, in which fibrosis develops after a

period of primary thrombocytosis.<sup>39,56,62,63</sup> To explore the association between Abi-1 deficiency and established pathogenetic mechanisms in MPNs, we assessed how heterozygous loss of Abi-1 affected the MPL<sup>W515L</sup>-mediated mouse model. Results showed accelerated MPN-like disease in Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> animals. These data support a contributory role of Abi-1 to the pathophysiology of MPNs. In addition to these findings, proteomics and immunoblotting (Supplementary Figure 9E) indicated upregulation of calreticulin in Abi-1<sup>KO</sup>. Calreticulin has an established role in MPN pathophysiology, and somatic *CALR* mutations occur in 30% of PMF.<sup>59</sup> At this point, the contribution of increased calreticulin to the Abi-1 loss-dependent phenotype remains unclear. Strikingly, Abi-1 deficiency in hematopoietic stem/progenitor cells results in upregulation of genes controlled, directly or indirectly, by NF-κB. Genome-wide expression profiling of Abi-1<sup>KO</sup> LSK cells highlighted overexpression of pro-inflammatory pathways associated with NF-κB activation. Upregulation of pro-inflammatory cytokines in PMF patients,<sup>65</sup> as well as TLR activation,<sup>66</sup> may contribute to marrow fibrosis. Our data support a mechanism in which Abi-1 loss results in upregulation of SFKs, which in turn activate STAT3, ultimately modulating the NF-κB pathway. Notably, STATs can be activated by SFKs,<sup>67,68</sup> and STAT3 and NF-κB pathways are linked.<sup>69 70</sup> Interestingly, Ser-32-phosphorylated IκB is not degraded and remains highly expressed in Abi-1<sup>KO</sup> marrow. IκB phosphorylation on Tyr allows NF-κB activation without IκB degradation<sup>71</sup>. Elucidation of how Abi-1 loss might lead to increased NF-κB signaling and the role of IκB phosphorylation in this process will require further research.

Abi-1 loss induced significant impairment of the hemostasis of HSCs. Increased cell cycle activity of Abi-1<sup>KO</sup> hematopoietic stem/progenitor cells is possibly leading to their expansion, displacement from the marrow, and progressive exhaustion, and is reflected in their impaired self-renewal capacities. Similarly, increased cycling and impaired self-renewal of HSCs was noted in animal models of hematopoietic tissue-specific overactivation of Akt or NF-κB.<sup>72,73</sup> Given the activity of *Mx1* promoter-driven Cre in mesenchymal stromal cells, it is reasonable to expect that changes in homeostasis of Abi-1<sup>KO</sup> HSCs may prove to be both intrinsic and non-autonomous in future experiments.

Analyses of *ABI1* transcripts in MPNs indicated that its loss may be specific to PMF.

Mechanisms responsible for loss of *ABI1* transcripts in PMF are however not clear. Notably, SNP-arrays performed on 151 MPN samples showed microdeletions in 10p12 region encompassing *ABI1* locus<sup>74</sup>. Sequencing of *ABI1* is warranted to provide mutational profile of the gene in MPNs.

In the aggregate, our data provide the first evidence for a role of Abi-1 in the pathogenesis of PMF, and a link to upregulation of the SFKs/STAT3/NF- $\kappa$ B signaling axis. To our knowledge, this is the first model of MPN mechanistically uncoupled from JAK2-STAT5 hyperactivity. Full elucidation of the cross-communication between JAK/SFKs/STAT3/NF- $\kappa$ B signaling may provide insight into the association between proliferative and pro-inflammatory features that characterize MPNs, particularly PMF. Targeting Abi-1 and associated pathways may offer a new therapeutic strategy for human PMF.

## **ACKNOWLEDGEMENTS**

This work was supported by Brown Biomed Division DEANS Award grant (PMD and EO), University of Medicine Foundation Integration Funding (PMD), Rhode Island Foundation grant (PMD), a pilot grant from the Lifespan COBRE Center for Cancer Research Development (NIH/NIGMS 1P30GM110759) (PMD), NIH/NIGMS 1P20GM119943 (PJQ, PMD) and Savit Foundation (PJQ, PMD). Work in Florence was supported by a grant from Associazione Italiana per la Ricerca sul Cancro (AIRC; Milan, Italy), Special Program Molecular Clinical Oncology 5x1000 to AIRC-Gruppo Italiano Malattie Myeloproliferative (AGIMM) project #1005. AHJ and LK were supported by NIH/NCI R01CA161018 (LK). AJO is supported by the American Society of Hematology Research Scholar award. We thank Mandy Pereira, Laura Goldberg, Mark Dooner, Elaine Pappa, Michael DeTatto, Joan Boylan, Aleksander F. Sikorski, Paola Guglielmelli and Loren Fast for helpful suggestions and technical assistance, and Eric Milner for his editorial help. We thank Matthew Adler and Veenu Aishwarya from AUM BioTech, LLC and Chris Gould from Biogen for technical assistance and helpful suggestions. We also thank John Crispino and Qiang Jeremy Wen (Northwestern University) for critical comments, discussion and technical assistance. We thank Ross Levine (MSKCC) for MSCV-MPL WT-IRES-GFP and MSCV-MPL W515L-IRES-GFP retroviral vectors.

## **AUTHORSHIP CONTRIBUTIONS**

A.Ch., J.M., M.P., N.K., Y.C., K.L., C.S., X.Y., R.Z., A.P., A.T., J.C., N.A., A.H.J. and O.L., performed experiments, acquired and analyzed data, and contributed to manuscript writing. N.A., D.O.T., A.J.O., T.C.Z., E.O., L.K., P.J.Q., J.R., P.G., R.M. and A.M.V. analyzed data and contributed to manuscript writing. P.M.D. designed research, analyzed data and wrote the manuscript. Authors declare no conflict of interest.

## REFERENCES

1. Tefferi A. Primary myelofibrosis: 2014 update on diagnosis, risk-stratification, and management. *Am J Hematol*. 2014;89(9):915-925.
2. Vardiman JW, Thiele J, Arber DA, et al. The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: rationale and important changes. *Blood*. 2009;114(5):937-951.
3. Barbui T, Thiele J, Gisslinger H, et al. The 2016 WHO classification and diagnostic criteria for myeloproliferative neoplasms: document summary and in-depth discussion. *Blood Cancer J*. 2018;8(2):15.
4. Gangat N, Caramazza D, Vaidya R, et al. DIPSS plus: a refined Dynamic International Prognostic Scoring System for primary myelofibrosis that incorporates prognostic information from karyotype, platelet count, and transfusion status. *J Clin Oncol*. 2011;29(4):392-397.
5. Kralovics R, Passamonti F, Buser AS, et al. A gain-of-function mutation of JAK2 in myeloproliferative disorders. *N Engl J Med*. 2005;352(17):1779-1790.
6. Scott LM, Tong W, Levine RL, et al. JAK2 exon 12 mutations in polycythemia vera and idiopathic erythrocytosis. *N Engl J Med*. 2007;356(5):459-468.
7. Tefferi A, Pardanani A. Serious adverse events during ruxolitinib treatment discontinuation in patients with myelofibrosis. *Mayo Clin Proc*. 2011;86(12):1188-1191.
8. Verstovsek S, Mesa RA, Gotlib J, et al. A double-blind, placebo-controlled trial of ruxolitinib for myelofibrosis. *N Engl J Med*. 2012;366(9):799-807.
9. Harrison C, Kiladjian JJ, Al-Ali HK, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. *N Engl J Med*. 2012;366(9):787-798.
10. Cervantes F, Vannucchi AM, Kiladjian JJ, et al. Three-year efficacy, safety, and survival findings from COMFORT-II, a phase 3 study comparing ruxolitinib with best available therapy for myelofibrosis. *Blood*. 2013;122(25):4047-4053.
11. Geyer HL, Mesa RA. Therapy for myeloproliferative neoplasms: when, which agent, and how? *Hematology Am Soc Hematol Educ Program*. 2014;2014(1):277-286.
12. Shi Y, Alin K, Goff SP. Abl-interactor-1, a novel SH3 protein binding to the carboxy-terminal portion of the Abl protein, suppresses v-abl transforming activity. *Genes Dev*. 1995;9(21):2583-2597.
13. Xiong X, Cui P, Hossain S, et al. Allosteric inhibition of the nonMyristoylated c-Abl tyrosine kinase by phosphopeptides derived from Abi1/Hsh3bp1. *Biochim Biophys Acta*. 2008;1783(5):737-747.
14. Brehme M, Hantschel O, Colinge J, et al. Charting the molecular network of the drug target Bcr-Abl. *Proc Natl Acad Sci U S A*. 2009;106(18):7414-7419.
15. Ikeguchi A, Yang HY, Gao G, Goff SP. Inhibition of v-Abl transformation in 3T3 cells overexpressing different forms of the Abelson interactor protein Abi-1. *Oncogene*. 2001;20(36):4926-4934.
16. Scita G, Nordstrom J, Carbone R, et al. EPS8 and E3B1 transduce signals from Ras to Rac. *Nature*. 1999;401(6750):290-293.
17. Innocenti M, Tenca P, Frittoli E, et al. Mechanisms through which Sos-1 coordinates the activation of Ras and Rac. *J Cell Biol*. 2002;156(1):125-136.
18. Innocenti M, Gerboth S, Rottner K, et al. Abi1 regulates the activity of N-WASP and WAVE in distinct actin-based processes. *Nat Cell Biol*. 2005;7(10):969-976.
19. Dubielecka PM, Ladwein KI, Xiong X, et al. Essential role for Abi1 in embryonic survival and WAVE2 complex integrity. *Proc Natl Acad Sci U S A*. 2011;108(17):7022-7027.
20. Sini P, Cannas A, Koleske AJ, Di Fiore PP, Scita G. Abl-dependent tyrosine phosphorylation of Sos-1 mediates growth-factor-induced Rac activation. *Nat Cell Biol*. 2004;6(3):268-274.
21. Campa F, Machuy N, Klein A, Rudel T. A new interaction between Abi-1 and betaPIX involved in PDGF-activated actin cytoskeleton reorganisation. *Cell Res*. 2006;16(9):759-770.

22. Beli P, Mascheroni D, Xu D, Innocenti M. WAVE and Arp2/3 jointly inhibit filopodium formation by entering into a complex with mDia2. *Nat Cell Biol.* 2008;10(7):849-857.
23. Innocenti M, Zucconi A, Disanza A, et al. Abi1 is essential for the formation and activation of a WAVE2 signalling complex. *Nat Cell Biol.* 2004;6(4):319-327.
24. Ring C, Ginsberg MH, Haling J, Pendergast AM. Abl-interactor-1 (Abi1) has a role in cardiovascular and placental development and is a binding partner of the alpha4 integrin. *Proceedings of the National Academy of Sciences of the United States of America.* 2011;108(1):149-154.
25. Elias JE, Gygi SP. Target-decoy search strategy for increased confidence in large-scale protein identifications by mass spectrometry. *Nat Methods.* 2007;4(3):207-214.
26. Sun X, Li Y, Yu W, Wang B, Tao Y, Dai Z. MT1-MMP as a downstream target of BCR-ABL/ABL interactor 1 signaling: polarized distribution and involvement in BCR-ABL-stimulated leukemic cell migration. *Leukemia.* 2008;22(5):1053-1056.
27. Xiong X, Chorzalska A, Dubielecka PM, et al. Disruption of Abi1/Hssh3bp1 expression induces prostatic intraepithelial neoplasia in the conditional Abi1/Hssh3bp1 KO mice. *Oncogenesis.* 2012;1:e26.
28. Sowalsky AG, Sager R, Schaefer RJ, et al. Loss of Wave1 gene defines a subtype of lethal prostate cancer. *Oncotarget.* 2015;6(14):12383-12391.
29. Tod J, Hanley CJ, Morgan MR, et al. Pro-migratory and TGF-beta-activating functions of alphavbeta6 integrin in pancreatic cancer are differentially regulated via an Eps8-dependent GTPase switch. *J Pathol.* 2017;243(1):37-50.
30. Steinestel K, Bruderlein S, Lennerz JK, et al. Expression and Y435-phosphorylation of Abelson interactor 1 (Abi1) promotes tumour cell adhesion, extracellular matrix degradation and invasion by colorectal carcinoma cells. *Mol Cancer.* 2014;13:145.
31. Juskevicius D, Lorber T, Gsponer J, et al. Distinct genetic evolution patterns of relapsing diffuse large B-cell lymphoma revealed by genome-wide copy number aberration and targeted sequencing analysis. *Leukemia.* 2016;30(12):2385-2395.
32. Chorzalska A, Salloum I, Shafqat H, et al. Low expression of Abelson interactor-1 is linked to acquired drug resistance in Bcr-Abl-induced leukemia. *Leukemia.* 2014;28(11):2165-2177.
33. Maruoka M, Sato M, Yuan Y, et al. Abl-1-bridged tyrosine phosphorylation of VASP by Abelson kinase impairs association of VASP to focal adhesions and regulates leukaemic cell adhesion. *Biochem J.* 2012;441(3):889-899.
34. Yu W, Sun X, Clough N, Cobos E, Tao Y, Dai Z. Abi1 gene silencing by short hairpin RNA impairs Bcr-Abl-induced cell adhesion and migration in vitro and leukemogenesis in vivo. *Carcinogenesis.* 2008;29(9):1717-1724.
35. Eto K, Nishikii H, Ogaeri T, et al. The WAVE2/Abi1 complex differentially regulates megakaryocyte development and spreading: implications for platelet biogenesis and spreading machinery. *Blood.* 2007;110(10):3637-3647.
36. Kuhn R, Schwenk F, Aguet M, Rajewsky K. Inducible gene targeting in mice. *Science.* 1995;269(5229):1427-1429.
37. Yu K, Salomon AR. HTAPP: high-throughput autonomous proteomic pipeline. *Proteomics.* 2010;10(11):2113-2122.
38. Ahsan N, Belmont J, Chen Z, Clifton JG, Salomon AR. Highly reproducible improved label-free quantitative analysis of cellular phosphoproteome by optimization of LC-MS/MS gradient and analytical column construction. *J Proteomics.* 2017;165:69-74.
39. Pikman Y, Lee BH, Mercher T, et al. MPLW515L is a novel somatic activating mutation in myelofibrosis with myeloid metaplasia. *PLoS Med.* 2006;3(7):e270.
40. Kelly LM, Liu Q, Kutok JL, Williams IR, Boulton CL, Gilliland DG. FLT3 internal tandem duplication mutations associated with human acute myeloid leukemias induce myeloproliferative disease in a murine bone marrow transplant model. *Blood.* 2002;99(1):310-318.

41. Storey JD, Tibshirani R. Statistical significance for genomewide studies. *Proc Natl Acad Sci U S A*. 2003;100(16):9440-9445.
42. Norfo R, Zini R, Pennucci V, et al. miRNA-mRNA integrative analysis in primary myelofibrosis CD34+ cells: role of miR-155/JARID2 axis in abnormal megakaryopoiesis. *Blood*. 2014;124(13):e21-32.
43. Zini R, Guglielmelli P, Pietra D, et al. CALR mutational status identifies different disease subtypes of essential thrombocythemia showing distinct expression profiles. *Blood Cancer J*. 2017;7(12):638.
44. Thiele J, Kvasnicka HM, Facchetti F, Franco V, van der Walt J, Orazi A. European consensus on grading bone marrow fibrosis and assessment of cellularity. *Haematologica*. 2005;90(8):1128-1132.
45. Kogan SC, Ward JM, Anver MR, et al. Bethesda proposals for classification of nonlymphoid hematopoietic neoplasms in mice. *Blood*. 2002;100(1):238-245.
46. Kfoury A, Virard F, Renno T, Coste I. Dual function of MyD88 in inflammation and oncogenesis: implications for therapeutic intervention. *Curr Opin Oncol*. 2014;26(1):86-91.
47. Viatour P, Merville MP, Bours V, Chariot A. Phosphorylation of NF-kappaB and I kappa B proteins: implications in cancer and inflammation. *Trends Biochem Sci*. 2005;30(1):43-52.
48. Machida K, Thompson CM, Dierck K, et al. High-throughput phosphotyrosine profiling using SH2 domains. *Mol Cell*. 2007;26(6):899-915.
49. Dubielecka PM, Machida K, Xiong X, et al. Abi1/Hssh3bp1 pY213 links Abl kinase signaling to p85 regulatory subunit of PI-3 kinase in regulation of macropinocytosis in LNCaP cells. *FEBS Lett*. 2010;584(15):3279-3286.
50. Kleppe M, Kwak M, Koppikar P, et al. JAK-STAT pathway activation in malignant and nonmalignant cells contributes to MPN pathogenesis and therapeutic response. *Cancer Discov*. 2015;5(3):316-331.
51. Akada H, Yan D, Zou H, Fiering S, Hutchison RE, Mohi MG. Conditional expression of heterozygous or homozygous Jak2V617F from its endogenous promoter induces a polycythemia vera-like disease. *Blood*. 2010;115(17):3589-3597.
52. Khan I, Huang Z, Wen Q, et al. AKT is a therapeutic target in myeloproliferative neoplasms. *Leukemia*. 2013;27(9):1882-1890.
53. Wisniewska-Chudy E, Szyberg L, Dworacki G, Mizera-Nyczak E, Marszalek A. pSTAT5 and ERK exhibit different expression in myeloproliferative neoplasms. *Oncol Rep*. 2017;37(4):2295-2307.
54. Fisher DAC, Malkova O, Engle EK, et al. Mass cytometry analysis reveals hyperactive NF Kappa B signaling in myelofibrosis and secondary acute myeloid leukemia. *Leukemia*. 2017;31(9):1962-1974.
55. Vannucchi AM, Bianchi L, Cellai C, et al. Development of myelofibrosis in mice genetically impaired for GATA-1 expression (GATA-1(low) mice). *Blood*. 2002;100(4):1123-1132.
56. Yan XQ, Lacey D, Hill D, et al. A model of myelofibrosis and osteosclerosis in mice induced by overexpressing thrombopoietin (mpl ligand): reversal of disease by bone marrow transplantation. *Blood*. 1996;88(2):402-409.
57. Marty C, Pecquet C, Nivarthi H, et al. Calreticulin mutants in mice induce an MPL-dependent thrombocytosis with frequent progression to myelofibrosis. *Blood*. 2015.
58. Wernig G, Mercher T, Okabe R, Levine RL, Lee BH, Gilliland DG. Expression of Jak2V617F causes a polycythemia vera-like disease with associated myelofibrosis in a murine bone marrow transplant model. *Blood*. 2006;107(11):4274-4281.
59. Nangalia J, Massie CE, Baxter EJ, et al. Somatic CALR mutations in myeloproliferative neoplasms with nonmutated JAK2. *N Engl J Med*. 2013;369(25):2391-2405.
60. James C, Ugo V, Le Couedic JP, et al. A unique clonal JAK2 mutation leading to constitutive signalling causes polycythaemia vera. *Nature*. 2005;434(7037):1144-1148.
61. Tiedt R, Hao-Shen H, Sobas MA, et al. Ratio of mutant JAK2-V617F to wild-type Jak2 determines the MPD phenotypes in transgenic mice. *Blood*. 2008;111(8):3931-3940.
62. Marty C, Pecquet C, Nivarthi H, et al. Calreticulin mutants in mice induce an MPL-dependent thrombocytosis with frequent progression to myelofibrosis. *Blood*. 2016;127(10):1317-1324.

63. Chachoua I, Pecquet C, El-Khoury M, et al. Thrombopoietin receptor activation by myeloproliferative neoplasm associated calreticulin mutants. *Blood*. 2016;127(10):1325-1335.
64. James C, Mazurier F, Dupont S, et al. The hematopoietic stem cell compartment of JAK2V617F-positive myeloproliferative disorders is a reflection of disease heterogeneity. *Blood*. 2008;112(6):2429-2438.
65. Hasselbalch HC. The role of cytokines in the initiation and progression of myelofibrosis. *Cytokine Growth Factor Rev*. 2013;24(2):133-145.
66. Micera A, Balzamino BO, Zazzo AD, Biamonte F, Sica G, Bonini S. Toll-Like Receptors and Tissue Remodeling: The Pro/Cons Recent Findings. *J Cell Physiol*. 2016;231(3):531-544.
67. Silva CM. Role of STATs as downstream signal transducers in Src family kinase-mediated tumorigenesis. *Oncogene*. 2004;23(48):8017-8023.
68. Kim LC, Song L, Haura EB. Src kinases as therapeutic targets for cancer. *Nat Rev Clin Oncol*. 2009;6(10):587-595.
69. Yu H, Pardoll D, Jove R. STATs in cancer inflammation and immunity: a leading role for STAT3. *Nat Rev Cancer*. 2009;9(11):798-809.
70. Grivennikov SI, Karin M. Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev*. 2010;21(1):11-19.
71. Imbert V, Rupec RA, Livolsi A, et al. Tyrosine phosphorylation of I kappa B-alpha activates NF-kappa B without proteolytic degradation of I kappa B-alpha. *Cell*. 1996;86(5):787-798.
72. Kharas MG, Okabe R, Ganis JJ, et al. Constitutively active AKT depletes hematopoietic stem cells and induces leukemia in mice. *Blood*. 2010;115(7):1406-1415.
73. Pietras EM. Inflammation: a key regulator of hematopoietic stem cell fate in health and disease. *Blood*. 2017;130(15):1693-1698.
74. Stegelmann F, Bullinger L, Griesshammer M, et al. High-resolution single-nucleotide polymorphism array-profiling in myeloproliferative neoplasms identifies novel genomic aberrations. *Haematologica*. 2010;95(4):666-669.

## FIGURE LEGENDS

**Figure 1. *ABI1* is downregulated in hematopoietic progenitor cells and granulocytes obtained from patients with MPNs.** (A) RT-PCR analysis of *ABI1* transcript levels in CD34+ cells isolated from bone marrow aspirates of patients with PMF (n=5) or healthy controls (n=5). *ABI1* gene expression was normalized to *RPP14* (RNaseP) and *ACTB*. (B) RT-PCR analysis of *ABI1* gene expression in peripheral blood granulocytes from patients with PMF (n=36), ET (n=15), PV (n=20), myelofibrosis post-ET (n=5) and post-PV (n=9) or from healthy controls (n=16), normalized to *RPP14*. (C) RT-PCR analysis of *ABI1* gene expression in peripheral blood granulocytes from PMF patients with *JAK2* (*JAK2* V617F) or *CALR* mutations (Del52 or Ins5), normalized to *RPP14*. Clinical details of the analyzed samples are presented in Supplemental Table 1. Boxes represent the interquartile range that contains 50% of the subjects, and the horizontal line in the box indicates the median; \*p<0.05.

**Figure 2. Transgenic *Abi-1*<sup>KO</sup> mice show splenomegaly, leukocytosis, thrombocytosis and decreased survival.** (A) Schematic representation of the experimental design of deleting *Abi1*<sup>fllox</sup> allele in *Abi1*(fl/fl);Tg(*Mx1-cre*)(+) mice by poly(I:C) induction. Analyses were conducted on animals at 14, 35, or 56 weeks of age (4, 25, or 46 weeks after recombination, respectively). (B) Genomic PCR analysis of *Abi1*<sup>fllox</sup> deletion efficiency in tail (top) and bone marrow (bottom) DNA. PCR amplification of the mutated, LoxP site containing *Abi1* allele produces a 359nt band, recombined *Abi1* allele produces no band, and non-mutated wild type *Abi-1* allele produces a 179nt band. A locus map is presented in Supplementary Figure 2A. Tissue from 3 different *Abi-1*<sup>WT</sup>, *Abi-1*<sup>HET</sup> and *Abi-1*<sup>KO</sup> animals was used. (C) Western blot analysis of *Abi-1* protein levels in the bone marrow of *Abi-1*<sup>WT</sup>, *Abi-1*<sup>HET</sup> and *Abi-1*<sup>KO</sup> mice. Tissue from 3 different animals per group was used. (D) Representative gross anatomy of femurs and spleens of 14, 35, and 56-week-old *Abi-1*<sup>WT</sup> or *Abi-1*<sup>KO</sup> animals. (E) Average spleen and liver sizes of *Abi-1*<sup>WT</sup> and *Abi-1*<sup>KO</sup> mice. Relative organ weight was calculated as an absolute organ weight(g)/body weight on sacrifice day (g) X100. Organs from twelve (n=12) gender-matched animals were evaluated per age group. (F) Average white blood cells count, hematocrit values and platelet count of the peripheral blood obtained from 14, 35, and 56-week-old *Abi-1*<sup>WT</sup> or *Abi-1*<sup>KO</sup> animals performed using

automated hematology analyzer. Peripheral blood from at least 12 gender-matched mice was analyzed per each age group. \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . (G) Survival of Abi-1<sup>WT</sup> (n=48) or Abi-1<sup>KO</sup> (n=60) animals monitored from birth for 67 weeks, (log-rank  $p = 0.003$ ).

**Figure 3. Abi-1 deficiency results in age progressive increase in circulating granulocytes, anisopoikilocytosis, megakaryocytosis, loss of stainable iron and fibrosis.** (A) FACS analysis of CD11b+/Gr-1+, B220+, and CD3+ populations in the peripheral blood of Abi-1<sup>WT</sup> and Abi-1<sup>KO</sup> mice, performed at 14, 35, or 56 weeks of age, 20 Abi-1<sup>WT</sup> and 20 Abi-1<sup>KO</sup> gender-matched mice were evaluated at each time point. (B) Wright staining of representative blood smears obtained from Abi-1<sup>KO</sup> and Abi-1<sup>WT</sup> mice at 56 weeks of age, teardrop cells are marked with arrows. Similar results were observed in blood smears of at least 12 gender-matched Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> mice. Magnification 100x, bars correspond to 20  $\mu\text{m}$ . Images were obtained using a Zeiss Axiophot microscope and Zeiss Pan-Apochromat 100x/1.40 oil lens. Representative (C) hematoxylin and eosin or (D) Prussian blue stains of bone marrow from femurs and spleen sections of 56-week-old Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> animals. Similar results were observed in at least 12 gender-matched Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> mice. Bars correspond to 100  $\mu\text{m}$ . Images were obtained using a Zeiss Axiophot microscope with Zeiss Pan-Apochromat 20x/1.0 lens. (E) Average number of megakaryocytes per high power field (hpf) and (F) percentage of macrophages with hemosiderin in the bone marrow and spleen/red pulp evaluated in 14, 35, and 56-week-old Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> animals, 6 gender-matched animals were evaluated per age group per genotype. Representative (G) Gomori reticulin staining of the bone marrow from femurs and spleen sections of 56-week-old Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> animals (black stain). Similar results were obtained for at least 12 gender-matched animals per group. Bars correspond to 100  $\mu\text{m}$ . Images were obtained using Zeiss Axiophot microscope with Zeiss Pan-Apochromat 20x/1.0 lens. \* $p < 0.05$ .

**Figure 4. Expansion, increased cell cycle activity, decreased engraftability and inflammatory signature in Abi-1 deficient hematopoietic stem/progenitor cells.** (A) Frequencies of Lin- cells in  $5 \times 10^5$  bone marrow (B) Frequencies of LSK or LK cells in lineage-negative fractions of Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> bone marrow. (C) Frequencies of long-term (LT)- and short-term (ST)-HSCs and MPPs

in LSK fractions were determined by sorting. Sorting was done using bone marrow obtained from 6 gender-matched 14-week-old Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> mice. Sorting strategy is presented in Supplementary Figure 5A. Average percentages of (D) EdU-positive LSK cells in G0/G1, S and G2/M phase and (E) EdU-positive LT-HSCs in G0/G1 and S/G2/M phase of the cell cycle. Data from four experiments each using bone marrow from 14-week-old gender-matched Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> animals are shown as mean. Non-competitive bone marrow primary (F) and secondary (G) transplant. Bone marrow cells isolated from Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> mice (before poly(I:C) injection) were transplanted into lethally irradiated recipient C57BL/6 wild-type mice in the absence of competitor cells. Four weeks after the transplantation, Abi-1 loss was induced by poly(I:C) administration. 24 weeks post-transplant bone marrow cells were harvested from primary recipients and transplanted into the secondary recipients. Average donor chimerism in primary and secondary recipients was monitored every 4 weeks for 24 weeks post-transplantation. n=4 Abi-1<sup>WT</sup> or n=4 Abi-1<sup>KO</sup> donor mice (CD45.1) were used for primary or secondary transplants, and 10 recipient lethally irradiated (CD45.2) mice per Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> transplant group were used. (H) Western blot analysis of Abi-1 protein levels and (I) average percentage of G0/G1, S or G2/M human CD34+ cells (n=3) exposed to FANA *ABI1* silencing antisense oligonucleotide (KD) or scrambled control (Ctrl) for 48h. (J) Heat map showing differently expressed transcripts in LSK-enriched cells isolated from the bone marrow of n=4 Abi-1<sup>KO</sup> and n=5 Abi-1<sup>WT</sup> 14-week-old animals. False discovery rate (FDR) was <0.05. (K) Cytokine levels detected in the plasma of 14-weeks old gender-matched Abi-1<sup>WT</sup> (n=12) and Abi-1<sup>KO</sup> (n=12) animals. \*p<0.05, \*\*p<0.01.

**Figure 5. Abi-1 loss in the bone marrow results in instability of WAVE2 complex, and is accompanied by increased phosphorylation of STAT3, SFKs, and NF-κB.** Western blot evaluation of the activity status of (A) NF-κB and its inhibitor IκB, (B) JAK2, STAT5, STAT3, Erk1/2, Akt and (C) SFKs using phospho-site-specific antibodies, and (D) stability of WAVE2 complex components (WAVE2, Nap1, Abi-1 and Sra-1) in Abi-1 deficient bone marrow. Bone marrow from three different Abi-1<sup>WT</sup> and Abi-1<sup>KO</sup> gender-matched 20-week old animals was used. (E) Volcano plot of fold change (log 2) versus q-value (-log<sub>10</sub>) of peak-area for 12,103 peptides identified in Abi-1<sup>KO</sup> (n=3) respective to

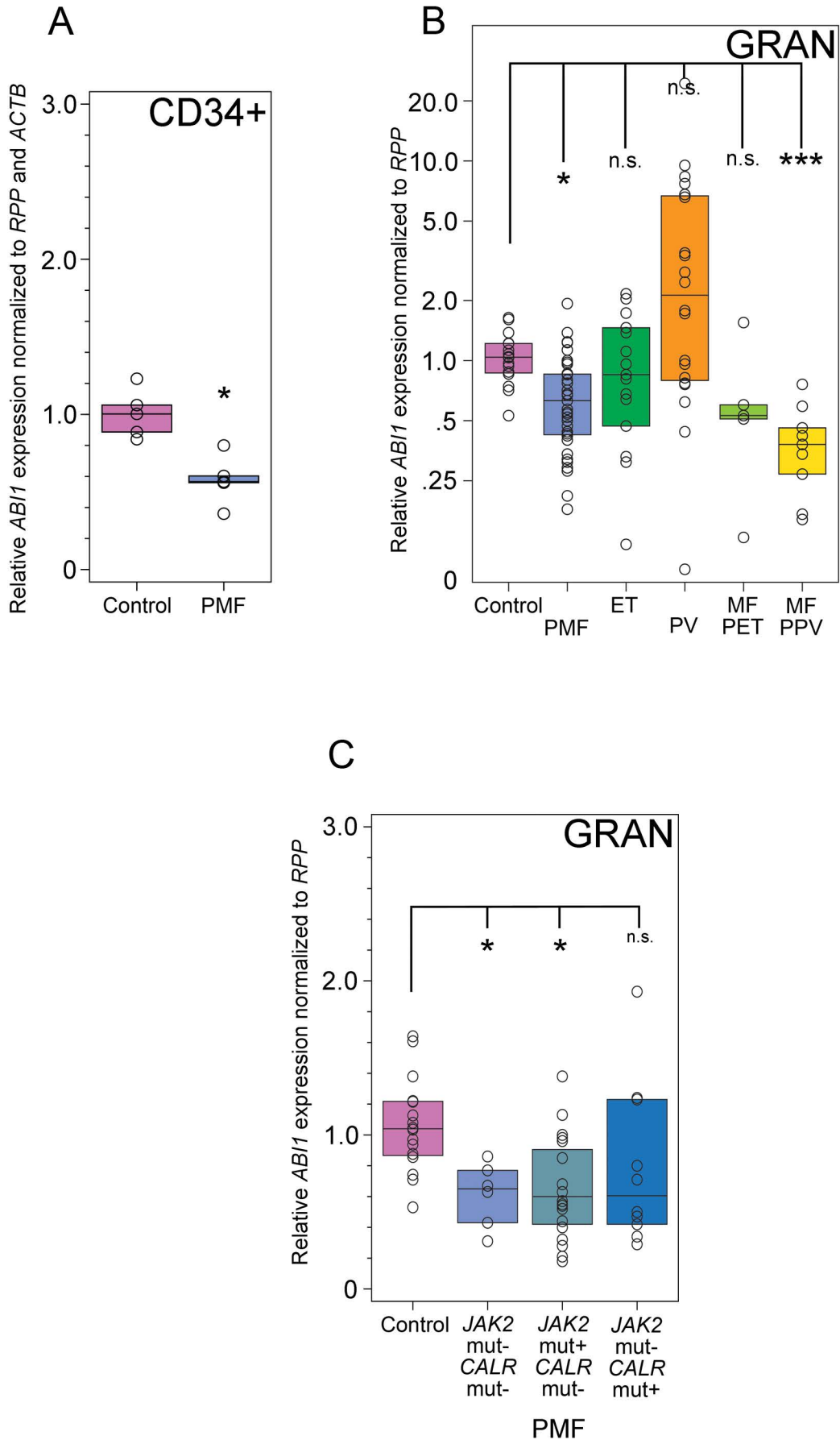
Abi-1<sup>WT</sup> (n=3) bone marrow isolated from gender-matched 20-weeks old animals. Green represents number of Peptides with 2-fold decrease in abundancies are presented in green, peptides with 2-fold increase in abundancies are presented in red. (F) Heatmap showing significantly changed peptides-derived from target proteins known to be associated with MPNs identified in LC-MS/MS analysis performed on Abi-1<sup>WT</sup> or Abi-1<sup>KO</sup> bone marrow samples. (G) Immunoblotting assessment of the levels of Abi-1 and activity status of SFKs, STAT3 and NF-κB in CD34+ cells isolated from the bone marrow of patients with PMF and compared to gender and age-matched healthy controls (sample details are presented in Supplementary Table 1). Whole cell lysates were used for immunoblotting analyses. (H) A hypothetical schematic depicting the effect of Abi-1 on SFK, STAT3 and NF-κB signaling. Abi-1 acts as a negative regulator of SFK activity. Its absence leads to overactive SFK signaling to STAT3, which becomes activated and cross-activates NF-κB. Overactivity of STAT3 and NF-κB results in cells acquiring inflammatory molecular signature, positively affecting proliferation, survival ultimately leading to transformation.

**Figure 6. Loss of one copy of Abi-1 accelerates disease development in the bone marrow**

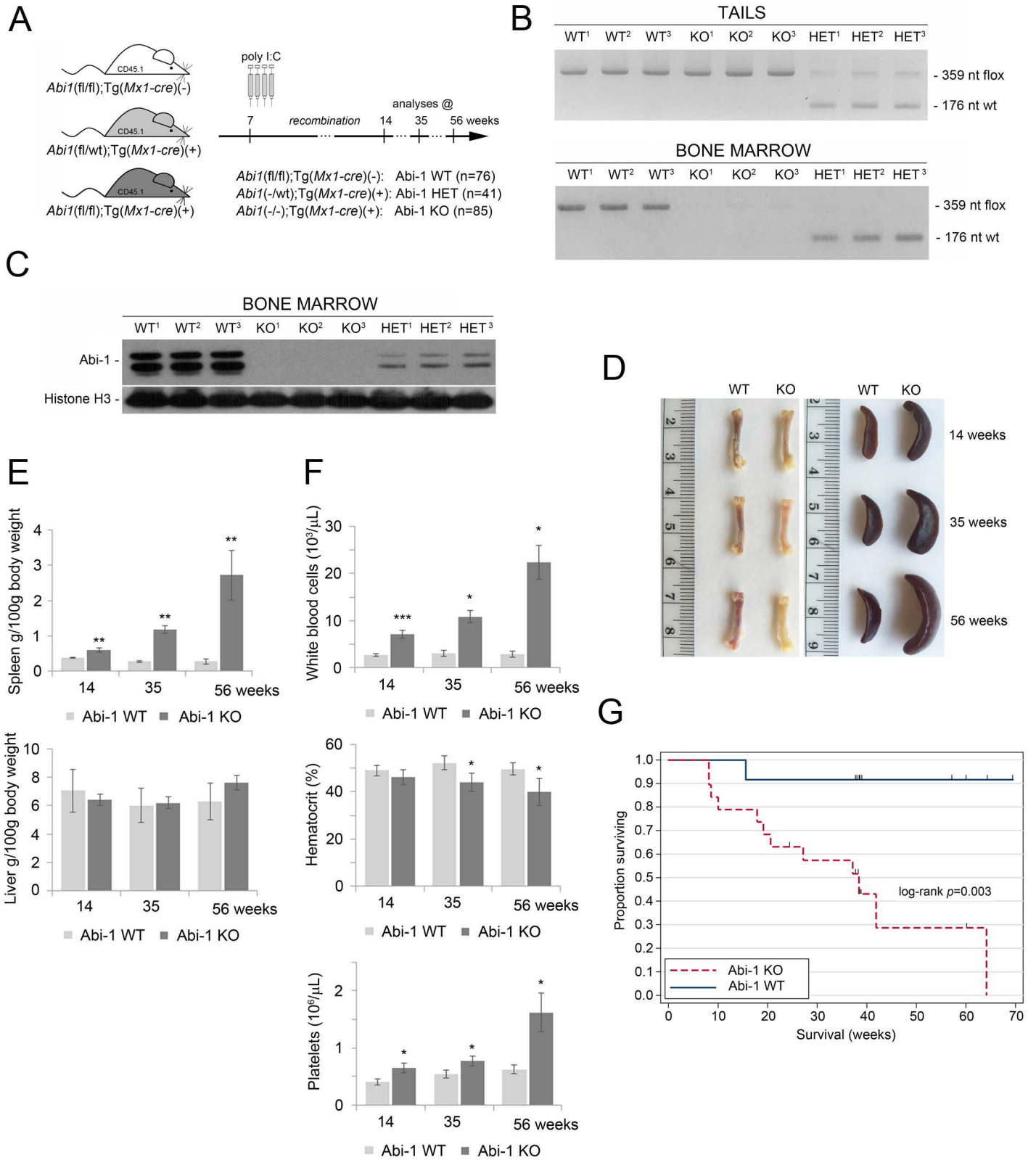
**transplantation model of MPL<sup>W515L</sup>-mediated myeloproliferative neoplasm.** (A) Schematic representation of Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> model. Bone marrow was obtained from 14-week-old gender matched Abi-1<sup>WT</sup> (n=3) or Abi-1<sup>HET</sup> (n=3) animals and transduced with retrovirus encoding Abi-1<sup>WT</sup> or W515L mutated GFP-tagged *MPL*. Infected bone marrows representing four experimental groups Abi-1<sup>WT</sup>/MPL<sup>WT</sup>, Abi-1<sup>HET</sup>/MPL<sup>WT</sup>, Abi-1<sup>WT</sup>/MPL<sup>W515L</sup> or Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> were next transplanted to C57BL/6J females (n=6 per experimental group). Analyses were conducted 60-days post-transplant. (B). Representative gross pathology images of femurs, spleens and livers of the animals from each experimental group are shown. (C) Average spleen and liver sizes of Abi-1<sup>WT</sup>/MPL<sup>WT</sup>, Abi-1<sup>HET</sup>/MPL<sup>WT</sup>, Abi-1<sup>WT</sup>/MPL<sup>W515L</sup> or Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> mice. Relative organ weight was calculated as an absolute organ weight(g)/body weight on sacrifice day (g) X100. Organs from six animals per group were evaluated. (D) Average white blood cells count, hematocrit and platelet count of Abi-1<sup>WT</sup>/MPL<sup>WT</sup>, Abi-1<sup>HET</sup>/MPL<sup>WT</sup>, Abi-1<sup>WT</sup>/MPL<sup>W515L</sup> or Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> mice. Peripheral blood from six animals per group

was analyzed. (E) Average frequencies of Mac1+/Gr-1+, CD41+, CD71+/Ter119+ or CD71+/Ter119- as well as B220+ and CD3+ cells in the bone marrow or peripheral blood obtained from Abi-1<sup>WT</sup>/MPL<sup>WT</sup>, Abi-1<sup>HET</sup>/MPL<sup>WT</sup>, Abi-1<sup>WT</sup>/MPL<sup>W515L</sup> or Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> mice. Samples from six animals per group were analyzed. (F) Gomori reticulin staining of the bone marrow from femurs and spleen sections of Abi-1<sup>WT</sup>/MPL<sup>W515L</sup> or Abi-1<sup>HET</sup>/MPL<sup>W515L</sup> animals (black stain). Bars correspond to 100  $\mu$ m. Images were obtained using Zeiss Axiophot microscope with Zeiss Pan-Apochromat 20x/1.0 lens. \*p<0.05

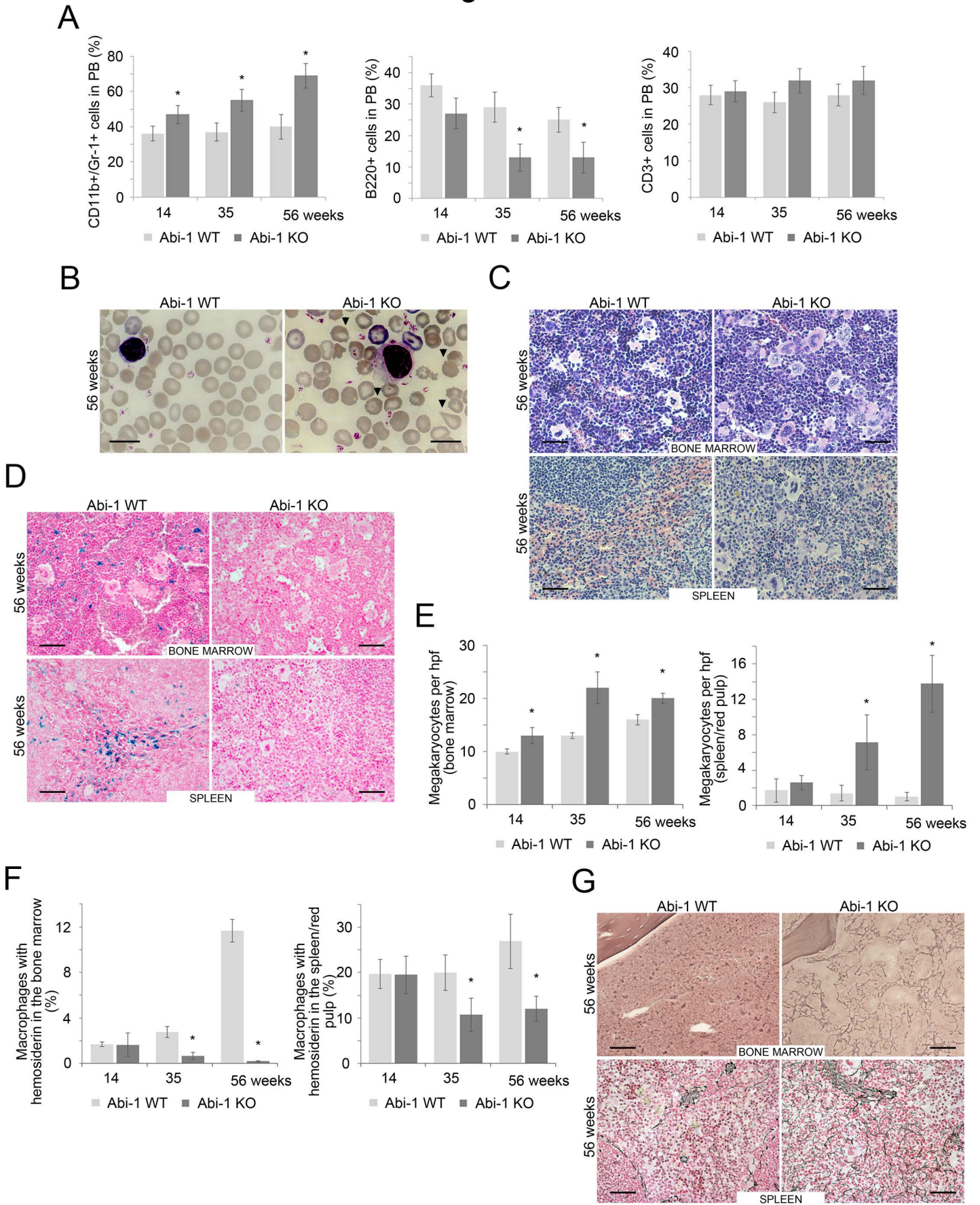
Figure 1



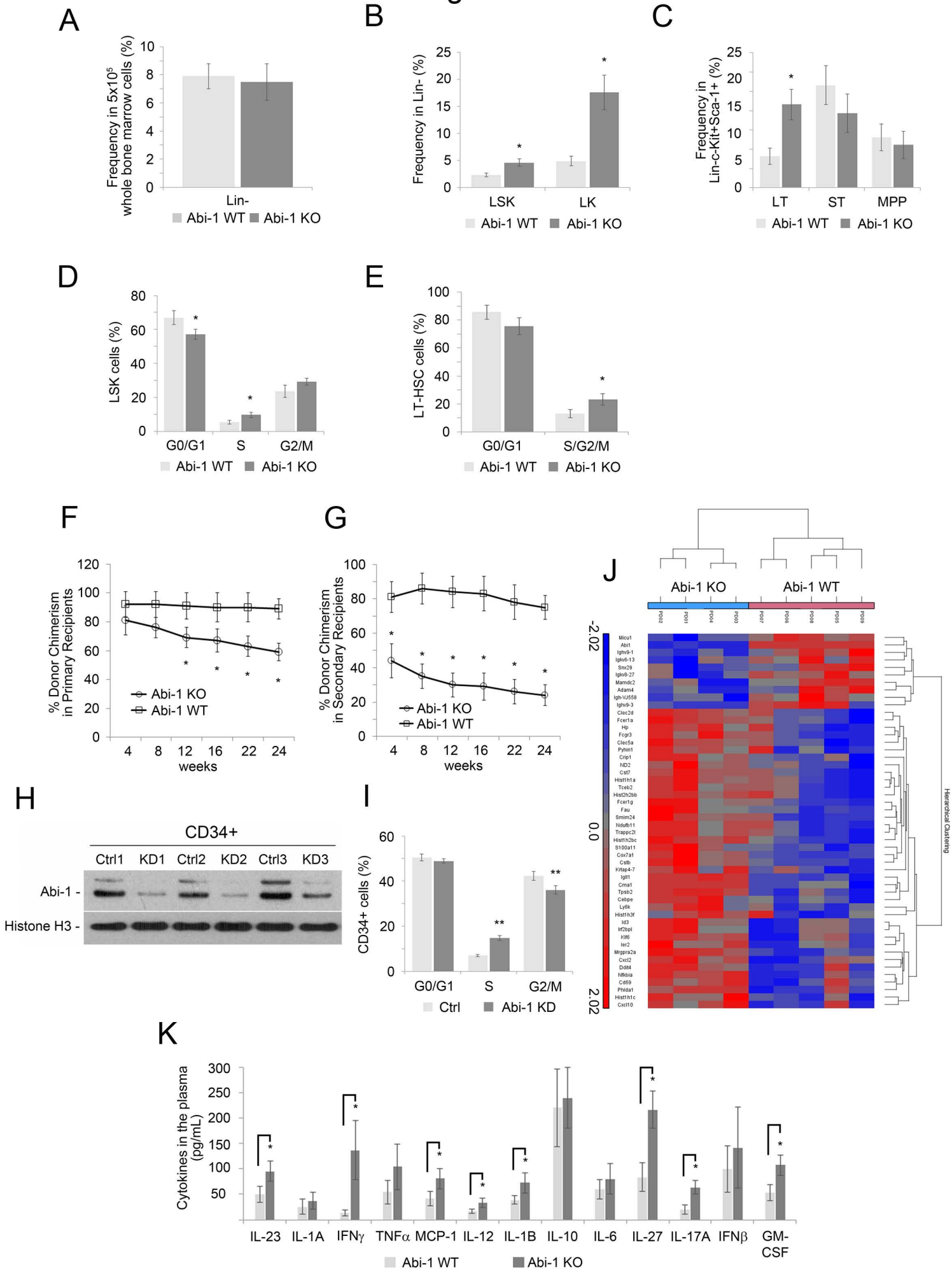
## Figure 2



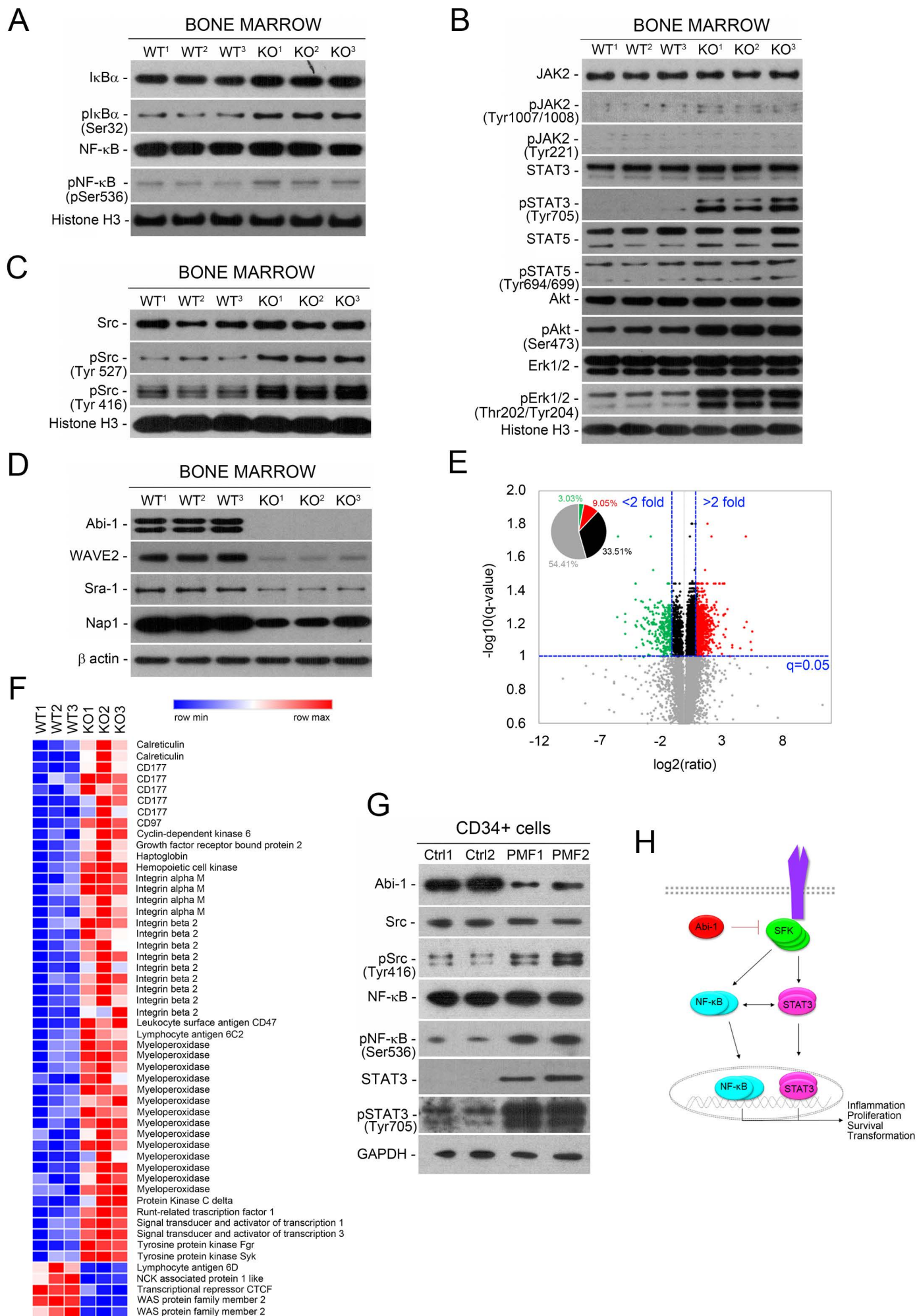
### Figure 3



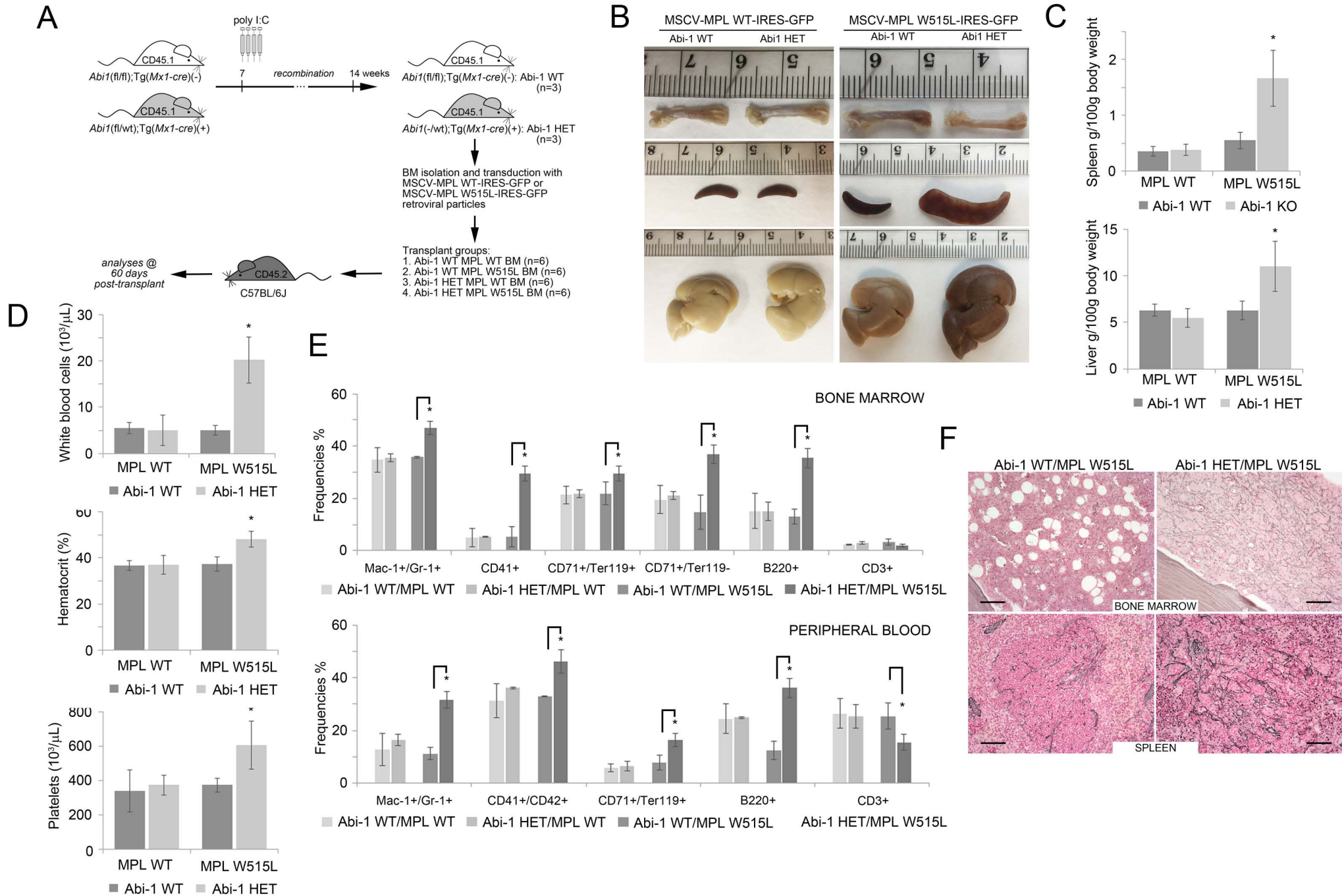
# Figure 4



# Figure 5



# Figure 6





**blood**<sup>®</sup>

Prepublished online September 13, 2018;  
doi:10.1182/blood-2018-05-848408

## **Bone marrow-specific loss of *ABI1* induces myeloproliferative neoplasm with features resembling human myelofibrosis**

Anna Chorzalska, John Morgan, Nagib Ahsan, Diana O. Treaba, Adam J. Olszewski, Max Petersen, Nathan Kingston, Yan Cheng, Kara Lombardo, Christoph Schorl, Xiaoqing Yu, Roberta Zini, Annalisa Pacilli, Alexander Tepper, Jillian Coburn, Anita Hryniewicz-Jankowska, Ting C. Zhao, Elena Oancea, John L. Reagan, Olin Liang, Leszek Kotula, Peter J. Quesenberry, Philip A. Gruppuso, Rossella Manfredini, Alessandro Maria Vannucchi and Patrycja M. Dubielecka

---

Information about reproducing this article in parts or in its entirety may be found online at:  
[http://www.bloodjournal.org/site/misc/rights.xhtml#repub\\_requests](http://www.bloodjournal.org/site/misc/rights.xhtml#repub_requests)

Information about ordering reprints may be found online at:  
<http://www.bloodjournal.org/site/misc/rights.xhtml#reprints>

Information about subscriptions and ASH membership may be found online at:  
<http://www.bloodjournal.org/site/subscriptions/index.xhtml>

---

Advance online articles have been peer reviewed and accepted for publication but have not yet appeared in the paper journal (edited, typeset versions may be posted when available prior to final publication). Advance online articles are citable and establish publication priority; they are indexed by PubMed from initial publication. Citations to Advance online articles must include digital object identifier (DOIs) and date of initial publication.